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FORMULATION AND CHARACTERIZATION OF CALCIUM SILICATE BASED FLOATING MICROSPHERES OF FAMOTIDINE FOR THE TREATMENT OF PEPTIC ULCER

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ABSTRACT: The aim of the present study was to prepare and evaluate floating microspheres consisting of (i) calcium silicate (CS) as porous carrier; (ii) famotidine and (iii) hydroxypropyl methylcellulose (HPMC) and ethylcellulose (EC) as polymers. The floating microspheres were evaluated for particle size, micromeritic properties, percent drug content, in vitro floating behavior, and in vitro drug release. The % yield of formulations (FM1 to FM9) was found to be in the range of 79.51 \pm 3.71 to 93.48 \pm 0.94 %. Percentage drug content of floating microspheres formulations (FM1 to FM9) was found in the range of 77.25 ± 0.36 to 86.14 ± 2.04 %. In Vitro Buoyancy percentage of the microspheres was found to be 97.5 ± 1.53 %. At pH 1.2, the best formulation FM4 showed maximum drug release (99.26 \pm 1.14 %) at the end of 12 hr. The SEM photographs of formulation FM4 showed that the fabricated microspheres were spherical with a smooth surface and exhibited a range of sizes within each batch. The in vivo evaluation for the determination of pharmacokinetic parameters was performed in albino rats. Higher plasma concentration was maintained throughout the study period from the floating microspheres of famotidine. The enhanced bioavailability and elimination half-life observed in the present study may be due to the floating nature of the dosage form. The results suggested that Calcium Silicate is a useful carrier for the development of floating and sustained release preparations. The results suggested that Calcium Silicate is a useful carrier for the development of floating and sustained release preparations.

INTRODUCTION: Oral delivery of drugs is by far the most preferable route of drug delivery due to ease of administration, patient compliance and flexibility in formulation etc. From immediate release to site-specific delivery, oral dosage forms have really progressed. However, it is a well accepted fact that it is difficult to predict the real *in vivo* time of release with solid, oral controlled release dosage forms.



Thus, drug absorption in gastrointestinal (GI) tract may be very short and highly variable in certain circumstances ¹. Several difficulties are faced in designing controlled release systems for better absorption and enhanced bioavailability. Various attempts have been made to prolong the retention time of the dosage form in the stomach.

One such method is the preparation of a device that remains buoyant in the stomach contents due to its lower density than that of the gastric fluids ². Single-unit formulations are associated with problems such as sticking together or being obstructed in the gastrointestinal tract, which may have a potential danger of producing irritation. On the other hand, a floating system made of multiple unit forms has relative merits compared to a single unit preparation. Indeed, the gastric emptying of a multiparticulate floating system would occur in consistent manner with small individual variations. On each subsequent gastric emptying, sink particles will spread out over a large area of absorption sites, increasing the opportunity for drug release profile and absorption in a more or less predictable way. Moreover, since each dose consists of many subunits, the risk of dose dumping is reduced ³.

For the present investigation, the drug used in the treatment of peptic ulcer i.e. famotidine, a histamine H2 -receptor antagonist was selected for its formulation as floating drug delivery system. The recommended oral dosage of famotidine is 20 mg twice daily or 40 mg once daily. Conventional dosage of 20 mg can inhibit gastric acid secretions up to 5 hours but not up to 10-12 hours and leads to fluctuation in plasma level and requires frequent dosing. Famotidine is absorbed only in the stomach and in the initial part of small intestine and has 40 -50 % absolute bioavailability. Therefore sustained release formulation of famotidine is desirable. The short biological half-life (2.5 to 3.5 hr.) also favors development of sustained release formulation. The traditional oral sustained release formulation releases most of the drug in the intestine and colon, thus the drug has a narrow absorption window ⁴.

The objective of the present work was to develop and characterize floating microspheres of famotidine, which after oral administration could prolong the gastric residence time and increase the drug bioavailability.

2. MATERIAL AND METHODS:

2.1 Materials: Famotidine was received as a gift from Cadila Pharmaceutical Ltd., sample Ahmedabad. Sigma Aldrich Laborchemikalien GMBH Bombay India provided Porous calcium silicate (Florite® RE, FLR). Ethyl cellulose was procured from Himedia Laboratories Ltd., Mumbai, India. Hydroxypropyl methylcellulose was Chemical from purchased G.S. Testing Laboratories, New Delhi, India. Polyvinyl alcohol, hydrochloric acid and tween 80, Dichloromethane, ethanol. All the other chemicals used were of analytical grade.

2.2. Preparation of fam absorbed CS: CS (1.0 g) was dispersed in 10 ml ethanolic solution of famotidine to prepare a slurry. The slurry was ultrasonicated for 10 min in an ice bath at 40% voltage frequency using a probe sonicator (Soniweld, Imeco Ultrasonics, India) to entrap the drug solution and reducing agent inside the pores of porous carrier. The excess ethanolic solution was removed by filtration and then drying in vacuum, which resulted in famotidine absorbed CS powders.

2.3. Preparation of floating microspheres: Microspheres were prepared using a modified emulsion solvent diffusion technique ⁵ Briefly, the drug absorbed CS was added into the polymer solution of ethyle cellulose and HPMC in ethanol and dichloromethane (2:1) and sonicated using probe sonicator. The resulting suspension was poured into 200 ml aqueous solution of PVA (0.75% w/v) at 40 °C. The emulsion or suspension was stirred at 500 rpm for 3 h employing a propeller type agitator. The microspheres were separated by filtration, washed with water and dried at room temperature in a desiccator for 24 h.

2.4 Size and shape of microspheres: The size of microspheres was determined using microscope (Olympus NWF 10x, Educational Scientific Stores, India) fitted with an ocular micrometer and stage micrometer. Scanning electron microscopy (SEM) (Leo 430, Leo Electron Microscopy Ltd, Cambridge, England) was performed to characterize the surface of the formed microspheres. Microspheres were mounted directly onto sample stub and coated with gold film (~200 nm) under reduced pressure (0.133 Pa).

2.5 Flow properties: The flow properties of microspheres were characterized in terms of angle of repose, carr index and hausner ratio ⁶. For determination of angle of repose (θ), the microspheres were poured through the walls of a funnel, which was fixed at a position such that its lower tip was at a height of exactly 2.0 cm above hard surface. The microspheres were poured till the time when upper tip of the pile surface touched the lower tip of the funnel. The tan-1 of the height of the pile / radius of its base gave the angle of repose. Microspheres were poured gently through a glass funnel into a graduated cylinder cut exactly to 10ml mark. Excess microspheres were removed using a

spatula and the weight of the cylinder with pellets required for filling the cylinder volume was calculated. The cylinder was then tapped from a height of 2.0 cm until the time when there was no more decrease in the volume. Bulk density (ρ b) and tapped density (ρ t) were calculated. Hausner ratio (HR) and carr index (IC) were calculated according to the two equations given below:

$$HR = \rho t / \rho b$$
$$IC = (\rho t \% \rho b) / \rho t$$

2.6 *In vitro* **buoyancy:** Microspheres (300mg) were spread over the surface of a USP XXIV dissolution apparatus type II filled with 900 mL of 0.1 N hydrochloric acid containing 0.02% tween 80. The medium was agitated with a paddle rotating at 100 rpm for 12 h. The floating and the settled portions of microspheres were recovered separately. The microspheres were dried and weighed. Buoyancy percentage was calculated as the ratio of the mass of the microspheres that remained floating and the total mass of the microspheres⁷.

2.7 Incorporation efficiency (IE): To determine incorporation efficiency floating microspheres were dissolved in a minimal amount of dichloromethane and the drug was extracted into a suitable aqueous media (0.1 N Hcl) by evaporating dichloromethane. The solution was filtered through 0.45 m membrane, diluted suitably and analyzed for drug content spectrophotometrically at 292 nm using 0.1 N HCl as blank.

2.8 *In vitro* **drug release studies:** The drug release was studied using a USP 24 dissolution apparatus type I (Veego Scientific, Mumbai) at 100 rpm in 0.1N Hcl as dissolution medium (900 mL) maintained at $37\pm1^{\circ}$ C. A sample (10 mL) of the solution was withdrawn from the dissolution apparatus hourly and the samples were replaced with fresh dissolution medium. The samples were filtered through a 0.45 μ membrane filter and diluted to a suitable concentration with 0.1 N Hcl. Absorbance of these solutions was measured at 292 nm using a SYSTRONICS 2202 UV/visible spectrophotometer. Cumulative percentage drug release was calculated using an equation obtained from a standard curve.

2.9 *In vivo* **Radiographical Study:** ⁸ In order to assess the gastro retentive efficacy of those floating formulations, the % buoyancy was determined by using barium sulphate X- ray contrast medium. 10% w/v polymer solution (composition ratio of 1:2 HPMC and EC) floating microsphere containing 15% barium sulphate as a contrast agent were prepared for radio graphical study. The study was carried with three healthy male rabbits free of detectable gastrointestinal diseases or disorders. The rabbits were fasted overnight. The rabbits were administered suspension of formulations (FM4) with 25 ml of water and X- ray photograph was taken immediately after administration.

2.10 Anti Secretory And Ulcer Protective Effect of Formulation In Ethanol Induced Gastric Ulcers^{9, 10} The studies were performed on two groups of animal (each group having six rats). Animals of group I were served as control and received 2 ml vehicle [1% CMC] whereas, animals of group II were served as test and administered with 2 ml suspension of formulation (equivalent to 3 mg/kg famotidine) orally. A three dose treatment was given to all the animal at a interval of 12 hrs and last dose was given one hour The percentage protection was calculated by following formula.

RESULTS AND DISCUSSION: The floating microspheres were prepared by emulsion solvent diffusion technique. A solution or suspension of polymer (HPMC and EC) and famotidine absorbed porous calcium silicate (CS) in ethanol and dichloromethane was poured into an agitated aqueous solution of polyvinyl alcohol (PVA). The evaporation subsequent of the entrapped dichloromethane led to the formation of internal cavities within the micro particles. The incorporation of drug absorbed CS into the formulation might have produced porous structure within the microspheres. (Table 1) (Fig.1 and 2) The flow properties of microspheres were characterized in terms of angle of repose, bulk density, tapped density and carr index. All formulation showed excellent flowability as expressed in term of angle of repose ($< 40^{\circ}$).

Angle of repose of the developed formulations FM1 to FM9 varied from 19.44 ± 0.48 % to 22.91 ± 0.65 %. Very good microspheres were produced when the polymeric phase was mixture of EC: HPMC at ratio: 2:1 FM4 has the lowest angle of repose (19.44 ± 0.48 %). Carr's Index was found in

the range of 18.94 ± 1.56 % to 23.07 ± 0.95 % for formulations FM1 to FM9. Angle of repose and carr index were determined to predict flow ability. Therefore all prepared floating microspheres showed better floability. (**Table 2**)

TABLE 1: FORMULATION CHART FLOATING MICROSPHERES OF FAMOTIDINE

Formulation	Amount of Drug (mg)	Total Amount of	otalAmount of EthylAmount ofount ofCelluloseHPMClymer(mg)(mg)		Amount of calcium	Amount of	Amount of		
Code		Polymer			(mg)		silicate	Ethanol/	PVA
	Diug (ing)	(mg)	%	mg	%	mg	(gm)	DCM 2:1 (ml)	(ml)
FM1	500	500	100	500	0	0	2.5	30	200
FM2	500	1000	100	1000	0	0	2.5	30	200
FM3	500	1500	100	1500	0	0	2.5	30	200
FM4	500	500	66.66	333.3	33.33	166.6	2.5	30	200
FM5	500	1000	66.66	666.6	33.33	333.3	2.5	30	200
FM6	500	1500	66.66	999.9	33.33	499.9	2.5	30	200
FM7	500	500	50	250	50	250	2.5	30	200
FM8	500	1000	50	500	50	500	2.5	30	200
FM9	500	1500	50	750	50	750	2.5	30	200

TABLE 2: MICROMERITIC PROPERTIES OF FLOATING MICROSPHERES OF FAMOTIDINE

Formulation Code	Angle of repose (°)	Carr's index (%)	Percentage Yield* ± S.D.	Percentage Drug Entrapment Efficiency* ± S.D.	Percentage Buoyancy* ± S.D.	Particle size (µm)
FM1	20.58 ± 1.10	18.94 ± 1.56	79.51 ± 3.71	78.71 ± 1.80	95.22 ± 1.55	95.73
FM2	20.27 ± 0.35	21.40 ± 3.22	88.85 ± 0.99	83.23 ± 1.78	96.72 ± 1.55	98.13
FM3	21.11 ± 0.26	$23.07{\pm}~0.95$	93.48 ± 0.94	86.02 ± 1.16	97.50 ± 1.53	103.02
FM4	19.44 ± 0.48	20.36 ± 2.20	85.83 ± 6.09	81.72 ± 1.04	93.15 ± 1.85	82.13
FM5	20.21 ± 0.51	21.44 ± 1.13	87.08 ± 2.24	84.26 ± 2.80	95.06 ± 3.21	88.20
FM6	21.11 ± 0.84	21.38 ± 1.13	92.11 ± 1.90	86.14 ± 2.04	96.37 ± 2.71	97.13
FM7	20.89 ± 0.60	19.67 ± 1.02	81.20 ± 1.48	77.25 ± 0.36	88.36 ± 4.85	79.01
FM8	21.51 ± 0.89	19.88 ± 0.83	84.84 ± 1.32	80.40 ± 1.42	94.39 ± 1.42	84.88
FM9	22.91 ± 0.65	20.17 ± 2.39	91.66 ± 1.68	83.18 ± 0.60	96.10 ± 2.44	92.95



FIG.1: SEM VIEW OF POROUS CALCIUM SILICATE PARTICLES FLR



FIG. 2: SEM VIEW OF CALCIUM SILICATE BASED FLOTING MICROSPHERES OF FAMOTIDINE (FM4)

The percentage yields of floating microspheres of formulations (FM1 to FM9) were found to be in the range of 79.51 ± 3.71 to 93.48 ± 0.94 %. Very good microspheres were produced when the polymeric phase was mixture of EC: HPMC at ratio: 2:1 FM6 has the highest yield $(92.11 \pm 1.68 \%)$. The size of microspheres formed may be a function of many factors such as stirring speed, viscosity of the dispersed phase and dispersion medium, temperature, conc. of polymer, amount and size of porous carrier. Mean particle size of the microspheres prepared with ethyl cellulose formulation FM1 was found to be 95.73 µm while it was significantly increased to 103.02 µm as the concentration of total amount of polymer was increased, % yield and drug entrapment (drug content) were also increased.

Percentage drug content of floating microspheres formulations (FM1 to FM9) were found in the range of 77.25 ± 0.36 to 86.14 ± 2.04 %. The high entrapment efficiency of drug may be attributed to their poor aqueous solubility. All formulation of drug content revealed above 77%. Formulation FM6 showed good % drug entrapment efficiency $(86.14 \pm 2.04 \%)$. (**Table 2**) In vitro Buoyancy percentage of the microspheres famotidine was in the range of 88.36 ± 4.85 to 98.75 ± 3.62 % at the end of 12 h (above 88 %). The nature of the polymer influenced the floating behavior of the microspheres. Good in vitro floating behavior was observed for all the microsphere formulation. This may be attributed to the low density CS within the system. (Fig. 3)



FIG. 3: PHOTOGRAPH OF *IN VITRO* BUOYANCY OF FLOATING MICROSPHERES IN SGF (pH 1.2) AFTER 0 AND 2 HR. (FIG A), 4 AND 6 HR. (FIG B), 8 AND 12 HR. (FIG C).

The releases of famotidine from calcium silicate based microspheres were studies in SGF (pH 1.2) and PBS (pH 7.4). Results revealed that the release rate depends upon the polymer concentration, amount of adsorbed polymer as well as the composition ratio of the HPMC and EC in the polymer solution. The cumulative release of famotidine significantly decreased with increasing ethyl cellulose concentration due to enteric property of ethyl cellulose polymer. There was no burst effect from any of these formulations. The increased density of the polymer matrix at higher concentrations results in an increased diffusion path length. This may decrease the overall drug release from the polymer matrix. At pH 1.2, the best formulation FM4 showed maximum drug release (99.26 \pm 1.14 %) at the end of 12 hr. While at pH 7.4, Formulation FM4 showed 98.13 \pm 1.30 % at the end of 12 hr. respectively. The results of formulations (FM1 to FM9) are shown in **Table 3** and **4** and graphically represented in **Fig. 4-9**.

TABLE 3: *IN-VITRO* DRUG RELEASE PROFILE OF FAMOTIDINE FROM FLOATING MICROSPHERES OF VARIOUS FORMULATIONS (pH 1.2)

Time		% Cumulative drug release*							
(h)	FM1	FM2	FM3	FM4	FM5	FM6	FM7	FM8	FM9
1	15.16 ± 4.03	11.59 ± 3.01	9.58 ± 2.78	28.98 ± 1.37	22.32 ± 1.68	19.34 ± 1.34	37.24 ± 4.07	$30.54{\pm}1.68$	27.86 ± 1.68
2	25.75 ± 4.39	21.97 ± 4.37	16.44 ± 3.41	39.76±1.70	32.12 ± 1.95	27.32 ± 2.20	52.81 ± 5.07	46.49 ± 2.69	42.45 ± 2.69
3	37.65 ± 4.69	31.13 ± 5.79	25.25 ± 3.75	48.34 ± 2.57	39.23 ± 2.30	32.12 ± 2.38	68.33 ± 5.79	61.94 ± 2.74	56.29 ± 3.39
4	45.22 ± 4.76	39.51±6.16	32.21±4.14	56.89 ± 2.07	47.27 ± 2.05	38.34 ± 2.07	78.77 ± 5.78	69.52±3.75	63.82 ± 4.11
5	53.07±6.15	48.41±2.24	40.15 ± 5.16	67.23 ± 1.42	55.23 ± 2.41	46.54 ± 2.43	86.66 ± 2.29	77.63±4.15	70.08 ± 3.81
6	62.16±4.27	54.72±1.57	44.36 ± 5.22	75.88 ± 1.78	61.65 ± 2.91	51.23 ± 2.14	91.58 ± 2.14	87.16±2.93	77.96±3.93
7	67.28 ± 3.32	61.33±1.69	49.51±5.30	80.48 ± 1.45	67.36 ± 2.46	59.45 ± 2.80	95.02 ± 1.60	91.43 ± 2.54	86.59 ± 3.35
8	72.01±2.68	67.11±1.77	54.49 ± 6.36	86.66 ± 2.17	76.34 ± 2.17	67.23 ± 2.20	$98.90{\pm}1.94$	95.74±1.10	92.62 ± 2.92
9	75.89 ± 3.39	71.65±3.29	60.42 ± 5.46	89.79±1.57	81.23 ± 1.50	74.34±2.19	-	-	-
10	79.47 ± 4.29	77.30 ± 4.07	66.85 ± 5.88	92.95±1.17	86.45 ± 0.94	79.12±1.88	-	-	-
11	86.21±3.87	82.34±2.61	72.98 ± 5.62	95.91±1.18	91.23±1.44	86.11±1.89	-	-	-
12	90.12 ± 2.46	86.09 ± 3.56	78.87 ± 4.47	99.26±1.14	95.89 ± 1.26	94.12±1.92	-	-	-

TABLE 4: *IN-VITRO* DRUG RELEASE PROFILE OF FAMOTIDINE FROM FLOATING MICROSPHERES OF VARIOUS FORMULATIONS (pH 7.4)

Time	% Cumulative drug release*								
(h)	FM1	FM2	FM3	FM4	FM5	FM6	FM7	FM8	FM9
1	16.10±1.89	12.47±2.34	11.77±1.66	19.34±1.65	13.32 ± 1.41	15.13±1.07	39.82±2.66	31.16±1.09	29.23±1.43
2	26.85 ± 2.41	25.12±2.7	19.89 ± 1.71	31.23 ± 1.64	24.17 ± 2.06	19.89 ± 2.14	52.69 ± 1.40	49.25 ± 1.63	43.11±2.57
3	36.83 ± 2.44	35.32 ± 2.6	25.43 ± 0.72	41.11±0.67	34.13±1.57	25.43 ± 1.51	$69.39{\pm}1.88$	65.43 ± 1.92	56.62 ± 3.33
4	47.91±2.63	38.94 ± 0.71	34.28 ± 1.54	48.19 ± 0.88	41.56±2.15	35.11 ± 0.00	$81.44{\pm}1.84$	71.70 ± 1.79	63.57 ± 2.05
5	55.76 ± 1.91	47.69±1.53	42.50 ± 2.44	56.11 ± 1.00	49.93 ± 1.14	42.50 ± 1.65	90.58 ± 2.26	81.94 ± 2.16	73.07 ± 1.51
6	65.61±2.16	54.01 ± 2.22	$45.84{\pm}1.47$	64.76±0.96	56.23±0.61	48.75 ± 0.81	$94.47{\pm}1.86$	89.48 ± 1.84	78.83 ± 1.03
7	68.89 ± 1.33	62.08 ± 1.91	51.37 ± 1.17	71.32±1.33	62.08 ± 2.03	53.32 ± 0.93	95.57±1.38	92.55 ± 1.97	87.59 ± 1.54
8	74.19 ± 2.05	69.31±2.09	55.76±1.91	79.23±0.54	71.21±1.67	$59.54{\pm}1.59$	97.45 ± 0.47	95.7 ± 2.60	92.89 ± 2.03
9	79.28 ± 3.10	74.61±1.34	64.08 ± 2.75	86.13±1.51	76.76±0.96	64.08 ± 1.71	-	-	98.4 ± 3.24
10	$83.85{\pm}1.81$	78.08 ± 1.52	67.78 ± 1.07	91.76±1.38	81.98 ± 1.74	68.43 ± 1.28	-	-	-
11	87.48 ± 1.77	82.55 ± 1.79	71.16±1.31	95.31±0.51	86.53 ± 1.56	$77.14{\pm}1.06$	-	-	-
12	90.83 ± 1.10	86.30 ± 2.63	78.17 ± 1.61	98.13±1.30	$92.32{\pm}1.28$	86.32 ± 2.01	-	-	-

The *in-vivo* study with X-ray contrast medium adsorbed on the floating microspheres were conducted to determine the *in-vivo* floating performance of 10% w/v polymer solution (composition ratio of 1:2 HPMC and EC) floating microspheres containing 15% barium sulphate as a contrast agent. It is clear from the X- ray photographs that microspheres remained buoyant even after 3 hr. (**Fig. 11**) The bioavailability study in animals showed that in animals of group I which received plain drug (famotidine), C_{max} was found to be 0.69 \pm 0.02 µg/ml after 4 hr. (t_{max}), AUC was 2.82 µg hr / ml. In second group of animals, which received formulation FM4, the serum concentration C_{max} of famotidine was found to be 0.84 \pm 0.04 µg/ml after 5 hr. (t_{max}), AUC was 3.58 µg hr/ ml.

These results showed that formulation FM4 released famotidine. (**Fig 10**) In the study of anti secretory and ulcer protective effect of FM4 formulations in absolute ethanol induced gastric ulcer model. In animals of group I acute ulcers were produced by the use of absolute alcohol (1ml) body weight. The animals of group II, which were administered suspension of formulations FM4 (2ml) before 30 min administration of absolute alcohol were found to protect gastric mucosa from ulcer. The suspensions of the formulation reduced the volume of gastric juice secreted by 37.5 %.

The total acidity was also reduced from 60 ± 2.1 to 35 ± 1.5 meq/100gm by the suspension formulations FM4. The ulcer index reduced significantly (P<0.01) from 32.8 ± 0.92 to 9.45 ± 0.51 by the formulation and showed the % inhibition of 71.18%. Therefore it is clear from this study that the floating formulations FM4 prevented the ulceration effectively for a longer period of time. (**Table 5-6** and **Fig. 12-14**)



FIG. 4: *IN-VITRO* DRUG RELEASE PROFILE OF FAMOTIDINE FROM FLOATING MICROSPHERES FORMULATIONS FM1 TO FM3 (pH 1.2)



FIGURE 5: *IN-VITRO* DRUG RELEASE PROFILE OF FAMOTIDINE FROM FLOATING MICROSPHERES FORMULATIONS FM4 TO FM6 (pH 1.2)



FIG. 6: *IN-VITRO* DRUG RELEASE PROFILE OF FAMOTIDINE FROM FLOATING MICROSPHERES FORMULATIONS FM7 TO FM9 (pH 1.2)



FIG. 7: *IN-VITRO* DRUG RELEASE PROFILE OF FAMOTIDINE FROM FLOATING MICROSPHERES FORMULATIONS FM1 TO FM3 (pH 7.4)



FIG. 8: *IN-VITRO* DRUG RELEASE PROFILE OF FAMOTIDINE FROM FLOATING MICROSPHERES FORMULATIONS FM4 TO FM6 (pH 7.4)



FIG. 9: *IN-VITRO* DRUG RELEASE PROFILE OF FAMOTIDINE FROM FLOATING MICROSPHERES FORMULATIONS FM7 TO FM9 (pH 7.4)



FIG. 10: PLOT OF MEAN SERUM LEVEL CONCENTRATION OF FAMOTIDINE VERSUS TIME AFTER ORAL ADMINISTRATION OF ITS FLOATING MICROSPHERE (FM4)



FIG. 11: X-RAY IMAGES OF RABBIT'S ABDOMEN AFTER ORAL ADMINISTRATION OF FM4 AND AM4 CONTAINING 15 % BARIUM SULPHATE AS A CONTRAST AGENT



FIG. 12: HISTOPATHOLOGY OF ULCERATED STOMACH OBTAINED FROM RATS OF CONTROL GROUP (0.5% CMC) TREATED WITH 1ML ABSOLUTE ETHANOL AFTER 1HR OF ADMINISTRATION (N=6). 45X, 10X



FIG. 13: HISTOPATHOLOGY OF ULCERATED STOMACH OBTAINED FROM RATS TREATED WITH FORMULATION AND ETHANOL AFTER 1HR OF ADMINISTRATION (N=6). 45X, 10X



FIG. 14: STOMACH OF CONTROL (A) AND (B) TREATED RATS AS SHOWN IN VISUAL INSPECTION

TABLE 5: MEAN PHARMACOKINETIC PARAMETERS OF FAMOTIDINE OBTAINED FOLLOWING ORAL ADMINISTRATION OF FORMULATION FM4

S. no	Formulation code	Dose administrated (mg/kg)	$C_{max}(\mu g/ml)$	T _{max} (hr.)	AUC ₀₋₇ (µghr/ml)
1	Plain drug (famotidine)	3	0.69 ± 0.02	4	2.82
2	FM4	3	0.84 ± 0.04	5	3.58

TABLE 6: EFFECT OF FORMULATION ON ULCERATION AND GASTRIC SECRETION USING ETHANOL INDUCED GASTRIC LESIONS IN RATS (N=6)

Treatment	Dose	Vol. of gastric juice ml/100gm	pН	Total acidity (meq/100g)	Ul	% Inhibition
Control	1ml ethanol	3.2 ml	1.1	60	32.8 ± 0.92	
Treated	3 mg/kg (Fam)	1.2 ml	1.6	35	9.45 ± 0.51 **	71.18

Results were expressed in Mean±SD. **P<0.01

CONCLUSION: Floating microspheres of famotidine was prepared by emulsion solvent evaporation method. Calcium silicate has been used as carrier. On the basis of results obtained in these investigations, the following conclusions may be drawn:

- It is possible to prepare an intragastric floating and sustained release preparation using calcium silicate (FLR) as the floating carrier by covering the pores of the FLR particles with adsorbed drug by a polymer solution containing both of HPMC and EC in suitable proportions.
- FLR based floating drug delivery system provides the possibility of enhancing the bioavailability and control the release of famotidine exhibiting absorption window by prolonging the gastric emptying time of the dosage form, ensuring availability of drug at the absorption site for the desired period of time.
- As the FLR microsphere with adsorbed drug and polymer coating showed a good floability

and drug release, it has a great potential for its use powder form encapsulation.

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