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PHARMACOGNOSTIC STUDIES AND QUALITATIVE ANALYSIS FOR PHYTOSTEROLS OF THE LEAVES OF *BOMBAX CEIBA* LINN.

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ABSTRACT: Bombax ceiba Linn. is one of the important plant species used in various system of medicine. Almost every part of plant is used as medicine and its roots and flowers are used for curing maximum number of ailments. Leaves of the plants are also containing different important secondary metabolites. For the use as safe herbal medicine the plant material (leaves) has to be standardized with the establishment of a consistent biological activity. Pharmacognistic studies includes different Physicochemical parameters like Proximate analysis of plant material such as Fluorescence Analysis, percentage of loss on drying (Moisture content), Total ash, Acid Insoluble ash, Water Soluble ash, used for the standardization, as per guideline of the in Indian Pharmacopoeia. Evaluation of drug in this ways was done for the confirmation of its identity and determination of its quality and purity and detection of nature of adulteration. Sterols present in plant called as Phytosterols. Phytoesterols with the principle advantage that inhibit the absorption of cholesterol and lower serum cholesterol levels was detected by basic techniques like Histochemical Studies, Preliminary Phytochemical analysis and Thin Layer Chromatography in the leaves of Bombax ceiba Linn.

INTRODUCTION: Herbal medicine is one of the oldest forms of medical treatment in human history, and could be considered one of the forerunners of the modern pharmaceutical trade. The herbal products today symbolize safety in contrast to the synthetics that are regarded as unsafe to human and environment. The clinical applications of these natural drugs have helped to revive an interest in higher plants as source of new drug¹.

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WHO guidelines include an important norm of standardization of herbal drugs. Standardization is a system that ensures a predefined amount of quantity, quality and therapeutic effect of ingredients in each dose². Herbal product cannot be considered scientifically valid if the drug tested has not been authenticated and characterized in order to ensure reproducibility in the manufacturing of the product. Moreover, many dangerous and lethal side effects have recently been reported, including direct toxic effects, allergic reactions, effect from contaminants and interactions with 3 . In view of the above. drugs herbal standardization thus is an important step for the establishment of a consistent biological activity, a consistent chemical profile or simply a quality

assuarance program for production and manufacturing of an herbal drug ⁴. Some Physicochemical parameters like Histochemical analysis, Proximate analysis of plant material such as the percentage of loss on drying (Moisture Content), total ash, acid insoluble ash, water soluble ash were determined as per guideline of the Indian Pharmacopoeia.

Plants contain an enormous number of biologically active compounds with various chemical structures. These phytochemicals, often secondary metabolites present in smaller quantities in higher plants include the alkaloids, flavonoids, tannins, terpenoids, sterols, and many others. Out of all these, Sterols present in many plants as an imperative phytoconstituent. Generally, phytosterols thought to stabilize plant membranes, with an increase in the sterol / phospholipid ratio leading to membrane rigidification ⁵. Phytosterols can be found at widely varying concentrations in the fatsoluble fractions of roots, stems, branches, leaves, seeds and blossoms etc. They are constituents of both edible and ornamental plants, including herbs, shrubs and trees ⁶.

Pharmacognostic studies of plant material for standardization and qualitative analysis for the presence of phytosterol by different tests is the basic aim of the present investigation.

MATERIALS AND METHODS:

Plant Material: The leaves of *Bombax ceiba* were collected, identified and authenticated from Flora of Maharashtra State -Vol. I and II⁷ and Flora of Nagpur District⁸.

Plant Profile: Taxonomical Classification:

Kingdom: Plantae Division: Angiospermae Class: Dicotyledoneae Order: Malvales Family: Malvaceae Juss. Genus: Bombax L. Species: ceiba Burm. f.

Vernacular Name: Semal, Silk Cotton tree, Shalmali.

Parts Used: Leaves.

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FIG. 1: PLANT OF BOMBAX CEIBA LINN.

Plant Description: Deciduous tree with cylindrical stem and horizontally spreading branches in whorls. The tree reaches upto 40 meter in height. Young stem and branches covered with sharp, stout prickeles. Leaves are palmately compound, glabrous with common petiole. Flowers are bright red in colour. Fruit is Capsule with many seeds.

Pharmacognistic Studies: 9

Proximate Analysis: Shade dried leaves of *Bombax ceiba* ground in a grinder and powder stored in a dry place for further processes.

Fluorescence Analysis: ¹⁰ 200 mg powder of Shade dried leaves of *Bombax ceiba* were taken in 10 test tubes and dissolved in 10ml of 1N NaOH (dissolved in Distilled water), 1N NaOH (dissolved in Alcohol), 10% HCl, Conc. HCl, 10% H₂SO₄, Conc. H₂SO₄, 10% HNO₃, Conc. HNO₃, Acetone and Distilled water, respectively. Colour of the solution was observed in the visible and UV light (Shorter wavelength – 254 nm and Longer wavelength – 365 nm)

Moisture Content: 2g powder of plant powder was taken in the weighed petriplates and the initial weight was taken. Petriplates with powder then kept in the oven for 90 mins at 100 - 105 °C. After 90 mins petriplates with powder was again weighed and kept for cooling. The loss of water was calculated. The % moisture content was determined by the following formulae:

Loss of water content $(W_0) = W_i - W_a$

Where, W_i = Weight of Petriplate with sample before drying; W_a = Weight of Petriplate with sample after drying.

% Moisture Content = $\frac{W0}{Ws} \times 100$ Where, W_s = Weight of Sample

Ash Content: ¹¹

Total Ash Content: 2g of accurately weighed quantity of powdered plant material (sample) was taken in a tarred silica crucible and incinerated at a temperature 600 ± 15 °C until free from carbon, cooled and weighed. % Total Ash was calculated by the following formulae:

Weight of Total ash $(W_t) = W_i - W_c$

Where, W_i = Weight of Crucible with sample after incineration; W_c = Weight of Crucible without sample before incineration

% Total Ash =
$$\frac{Wt}{Ws} \times 100$$

Where, $W_s =$ Weight of Sample

Acid Insoluble Ash Content: Total ash obtained was mixed and stirred well with 25 ml of boiled 2M Hydrochloric acid. The insoluble matter was collected on an ashless filter paper. Filter paper with insoluble matter was dried in oven at 100 °C for 15 mins % Acid insoluble ash was calculated by the following formula:

Weight of Acid Insoluble Ash $(W_a) = W_i - W_f$

Where, W_i = Weight of Filter paper with insoluble matter after drying; W_f = Weight of Filter paper

% Acid Insoluble Ash = $\frac{Wa}{Ws} \times 100$

Where, $W_s =$ Weight of Sample

Water Soluble Ash Content: Total ash obtained was mixed and stirred well with 25ml of boiled Distilled water. The insoluble matter was collected on an ashless filter paper. Filter paper with insoluble matter was dried in oven at 100 °C for 15 mins % Water Soluble ash was calculated by the following formulae:

Weight of water Insoluble Ash $(W_w) = (W_m - W_f)$ Where, Wm = Weight of Filter paper with insoluble matter after drying; $W_f =$ Weight of Filter paper.

% Water Soluble Ash = $\frac{Wt - Ww}{Ws} \times 100$ Where, W_s = Weight of Sample **Qualitative Analysis (Detection of Phytosterols): Histochemical Analysis:** Thin Tranverse sections of leaves were taken and mounted on glycerine and Antimony Trichloride (SbCl₃). Pinkish brown stained cells in the sections mounted in Antimony Tricholride were compared with sections mounted in glycerine for the presence of phytosterols¹².

Preliminary Phytochemical Analysis:

Standard Lab Tests: Two standard methods were done to determine the presence of sterols for the comparative confirmation with field tests ^{13, 14}

Salkowski Test: Salkowski test was done with the maceration of selected plants. 2ml extract taken in a test tube. 2ml Chloroform and 2ml conc. Sulphuric acid was added in it. Brown or red colored ring on the sulphuric acid layer given the confirmatory test ¹⁵.

Liebermann Burchard's Test: Liebermann and Burchurd's test was done after the extraction and reflux of the plant material. 2ml extract taken in a test tube. 2ml Chloroform, 2ml Aceticanhydride and 2ml conc. Sulphuric acid was added in it. Translucent green color given the confirmatory test ^{16, 17}.

Thin Layer Chromatography: The active extract (Hexane) obtained from the dried leaves of leaves of *Bombax ceiba* Linn. and was subjected to thin layer chromatography to find out the Phytosterol contents in it ^{18, 19}. The adsorbent used for preparation of thin layer plate as a stationary phase was Silica Gel G. 15 g powder of Silica Gel G was mixed with 30ml Distilled water. This Silica Gel G suspension was spread with a spreader on thin layer chromatographic glass plates fixed on a stage. The prepared plates were air-dried and activated in an oven at 110 °C for 30 min.

The activated plates then used for the application of samples and standard solutions [β – Sitosterol (M P Bio meditech), in Hexane with capillary tubes. The spots of samples and standard solutions were applied on plate, keeping distance of approximately 1cm. The chromatographic glass chamber was saturated with the moistened filter paper by dipping it in selected solvent system. The applied plate then kept in a mobile phase, Benzene : Ethyl Acetate (5: 1). The developed plate then derivatize with spraying reagent (20% Antimony Trichloride in

Chloroform) for the visualization of phytosterol spots. The $R_{\rm f}$ values of standard spots and sample spots were calculated.

 $R_f \ value = \qquad \frac{\text{Distance travelled by the Compound}}{\text{Distance travelled by the Solvent}}$

RESULTS AND DISCUSSIONS: Pharmacognistic Studies:

Proximate Analysis: Different solvents showed visibly different colour patterns with suspended powder (**Table 1**). Hence, fluorescence studies were undertaken the purity check of the plant materials.

This is so because if any adulteration is present in the plant material mixture, the color so obtained will be different from the pure one, proves the significance of the fluorescence studies. The values of all the Physicochemical determinants *viz*. Moisture Content, Ash Values etc. are summarized in **Table 2**. Moisture content examined in the intended plants was not too much so that less prone to microbial contamination and eventually prevent from decomposition. Total Ash Content of *Bombax ceiba* Linn. (Leaves) was found to be more which shows that these plants contain more carbonates, phosphates, silicates *i.e.* mineral contents and some inorganic elements.

 TABLE 1: FLUORESCENCE ANALYSIS OF BOMBAX CEIBA L. (LEAVES)

Sr.		Colour of the Powder with Solvent in the corresponding Wavelength		
No.	Powder + Solvent	Visible	254nm	365nm
1.	NaOH in Distilled Water	Dark Brown	Brownish Black	Blackish Brown
2.	NaOH in Alcohol	Dark Green	Blackish Brown	Black
3.	10% HCl	Yellowish Brown	Muddy Brown	Brown
4.	Conc. HCl	Brownish Yellow	Blackish Brown	Blackish Brown
5.	$10\% H_2SO_4$	Yellow	Blue	Fluorescent Blue
6.	Conc. H_2SO_4	Reddish Brown	Blackish Blue	Greenish Black
7.	10% HNO ₃	Orange	Greenish Blue	Dark Green
8.	Conc. HNO ₃	Dark Orange	Blackish Blue	Black
9.	Acetone	Green	Greenish Blue	Blackish Green
10.	Distilled Water	Muddy Yellow	Blackish Brown	Brownish Black

TABLE 2: VALUES OF PHYSICO-CHEMICALPARAMETERS OF LEAVES OF BOMBAX CEIBA L.

Sr.	Parameters	Values
No.		Mean ± SEM
1	Moisture Content	3.07 ± 0.202
2	Total Ash Content	15.72 ± 0.301
3	Acid Insoluble Ash Content	1.4 ± 0.15
4	Water Soluble Ash Content	0.85 ± 0.26

Qualitative Analysis (Detection of Phytosterols): Histochemical Analysis: The Reddish Brown colored granules was observed in the Palisade and Spongy Tissues, stained with Antimony Trichloride (SbCl₃) (Fig. 2). These granules were assumed to be stained steroidal compounds according to the study of Trease & Evans (1986). Thus this study had given a primary confirmation of location of phytosterols in the plant material.

Preliminary Phytochemical Analysis: Salkowski's Test and Liebermann Burchard's Test are the two important test for the detection of sterols from the plant material. Leaves of *Bombax ceiba* L. showed both the tests positive for the phytosterols (**Table 3**).

TABLE3:SALKOWSKIANDLIEBERMANBURCHARD TEST

Plant	Parts	Salkowski	Lieberman
Name	Used	Test	Burchard Test
Bombax	Leaves	+	+
ceiba L.			

'+' indicates the positive test for the presence of Phytosterols.

Thin Layer Chromatography: TLC is also used to support the identity of a compound in a mixture when the R_f of a compound is compared with the R_f of a known compound. Additional tests involve the spraying of phytochemical screening reagents, which cause color changes according to the phytochemicals existing in a plants extract; or by viewing the plate under the UV light ²⁰.

The leaves extract in the present investigation showed different spots, out of them 0.26 R_f value was much similar with that of the R_f value of standard β -Sitosterol which was 0.26 (**Table 4**).

Distance Travelled by Solvent (Solvent front) = 8.5 cm.



IN GLYCERINE (MID RIB REGION)

IN GLYCERINE (WING OF LEAF REGION)



IN SBCl₃ (MID RIB REGION)

IN SBCl₃ (WING OF LEAF REGION)

FIG. 2: BOMBAX CEIBA (LEAF SECTION)

TABLE 4: THIN LAYER CHROMATOGRAPHY

Sr. No.	Name of Extract/Standard	Distance Travelled by	R _f value of
		Bands (cm)	Bands
1.	β-Sitosterol (Standard)	2.2	0.26
2.	Bombax ceiba L.	0.9, 1.9, 2.2, 2.9, 3.6, 4.2,	0.11, 0.22, 0.26, 0.34, 0.42,
	(Leaves extract)	4.6, 5.3, 6.0	0.49, 0.54, 0.62, 0.71

The brownish colour appear in both β -Sitosterol (Standard) and *Bombax ceiba* Linn. (Leaves extract) at R_f value 0.26, after derivatization, confirms the presence of phytosterols in the leaves of *Bombax ceiba* Linn.

CONCLUSION: The information from pharmacognostic study will be helpful to differentiate *Bombax ceiba* Linn. from the closely related other species and varieties. Proximate analysis can be considered as standardized enough to identify and decide the authenticity of this drug. Qualitative analysis for phytosterols was found to be positive in all the selected parameters for study. The present physicochemical data thus emphasize the knowledge of quality and identity of the plant *Bombax ceiba* L. (Leaves).

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CONFLICT OF INTEREST: Nil.

REFERENCES:

- 1. Phillipson J: Phytochemistry and Medicinal Plants. Phytochemistry 2000; 56: 237-248.
- 2. Sagar BP, Zafar R and Panwar R: Herbal drug standardization. The Indian Pharmacist 2005; 4(35): 19-22.
- 3. George P: Concerns regarding the safety and toxicity of medicinal plants An overview Journal of Applied Pharmaceutical Science 2011; 1(6): 40-44.
- 4. Patra K, Pareta S, Harwansh R and Kumar JK. Traditional approaches towards standardization of herbal medicines A review. J Pharm Sci Technol 2010; 2(11): 372- 379.
- 5. Moreau RA, Whitaker BD and Hicks KB: Phytosterols, phytostanols, and their conjugates in foods: Structural diversity, quantitative analysis, and health-promoting uses. Progress in Lipid Research 2002; 41(6): 457-500.

- Clifton P, Noakes M, Sullivan D, Erichsen N and Ross D: Cholesterol-lowering effects of plant sterol esters differ in milk, yoghurt, bread and cereal. European Journal Clinical Nutrition 2004; 58: 503-509.
- 7. Singh N and Kartikeyan S: Flora of Maharashtra State, Botanical Survey of India 2000.
- 8. Ugemuge N: Flora of Nagpur District 1986.
- Mondal P, Sharan S, Md. Zaman K, Bhuyan N and Das S: Pharmacognostical Studies and Phytochemical Evaluation of the Stem Barks of *Embilica officinalis* Gaertn. Indo Global Journal of Pharmaceutical Sciences 2013; 3(1): 58-66.
- Chase C and Pratt R: Fluorescence of powdered vegetable drugs with particular reference to development of a system of identification. Journal of the American Pharmaceutical Association 2006; 38(6): 324-331.
- 11. Lee S: Encyclopedia of Chemical processing. (L. Lee, Ed.) CRC Press 2005.
- 12. Trease G and Evans W: Pharmacognosy. East Bourne: Bailliere Tindal 1986.

- E-ISSN: 0975-8232; P-ISSN: 2320-5148
- 13. Krishnaiah D, Sarbatly R and Bono A: Phytochemical antioxidants for health and medicine: A move towards nature. Biotechnol Mol Biol Rev 2007; 1: 97-104.
- Anasane P and Chatuevedi A: Field Test for the Detection of Phytosterols. International Research Journal of Pharmacy 2014; 5 (9): 734-736.
- 15. Salkowski E: Berichte der Deutschen Chemischen Gesellschaft 1872; 637.
- 16. Liebermann C: Ber. dtch. chem. Ge, 1803; 18.
- 17. Burchard: Dissertation, Rostock. Chem. Zbl 1890.
- Kagan IA and Flythe MD: Thin Layer Chromatography (TLC) Separations and Bioassays of Plant Extracts to Identify Antimicrobial Compounds. Journal of Visualized Experiments 2014; 85: 1-8.
- 19. Wagner H and Bladt S: Plant drug analysis, A thin layer Chromatography. CBS Publisher and Distributors 1995.
- 20. Sasidharan S, Chen Y, Saravanan D, Sundram K and Yoga LL: Extraction, Isolation and Characterization of Bioactive compounds from Plants extracts. Afr J Tradit Complement Altern Med 2011; 8(1): 1-10.

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