



Received on 16 June, 2017; received in revised form, 10 August, 2017; accepted, 17 September, 2017; published 01 March, 2018

PROTECTIVE EFFECT OF QUERCETIN ON BISPHENOL A - INDUCED ENZYMATIC CHANGES IN TESTIS OF MICE

Sanman Samova*, Hetal Doctor and Ramtej Verma

Department of Zoology, School of Sciences, Gujarat University, Ahmedabad - 380009, Gujarat, India.

Keywords:

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Correspondence to Author:

Sanman Samova

Department of Zoology,
School of Sciences, Gujarat
University, Ahmedabad - 380009,
Gujarat, India.

E-mail: samova.sanman@gmail.com

ABSTRACT: Bisphenol A is well known endocrine disturbing chemical. Evidence has demonstrated that exposure to certain levels can cause toxic effect in humans and laboratory animal models. Quercetin is consistently the most active of the flavonoids in experimental studies. The present investigation was an attempt to evaluate the effect of Bisphenol A on testis and its amelioration by quercetin. Inbred Swiss strain male albino mice were orally administered with BPA (80, 120 and 240 mg/kg body weight/day) for 45 Days. On the completion of treatment, animals were sacrificed, testis was isolated and used for evaluation of energy metabolism (SDH and ATPase activity) and phosphatases activity (ACP, ALP). The results revealed that oral administration of BPA for 45 days caused significant dose-dependent reduction in activities of ATPase and SDH. However, ACP and ALP activities were significantly dose-dependent increased in BPA treated mice. Oral administration of Quercetin (30, 60 and 90 mg/kg body weight/day) along with high dose of BPA (240 mg/kg body weight/day) for 45 days caused significant amelioration in all biochemical parameters as compared to high dose of BPA treated group. Present study concludes that BPA causes reduction in energy metabolism and induces the activity of phosphatases. The effect was significant and dose-dependent. This amelioration in toxic effect of BPA might be due to antioxidant properties of quercetin.

INTRODUCTION: Interest in dietary phenolics has increased recently, due to their antioxidant properties (free radical scavenging and metal chelating) and their possible beneficial implications in human health. Quercetin (3, 3', 4', 5, 7-pentahydroxyflavone) is a natural polyphenolic flavonoid widely found in edible plants such as fruits, vegetables, herbs, grains and beverages (*i.e.* tea, red wine)¹.

It is a versatile molecule, owing to its specific chemical structure, counteracts oxidative stress generated as a result of reactive oxygen species (ROS), have many pharmacological properties including antioxidant²⁻⁴, neurological, antiviral^{5, 6}, anticancer^{7, 8}, cardiovascular^{9, 10}, antimicrobial¹¹, anti-inflammatory agent¹², Hepatoprotective^{13, 14} and fertility^{15, 16}. Bisphenol - A [2, 2-bis (4-hydroxyphenyl) propane] (BPA) is an important environmental contaminant¹⁷.

Exposure to BPA is widespread and associated with a wide range of health outcomes in animal and human studies^{18, 19}. BPA is an important component of polycarbonate and epoxy resins. It is mainly found in the composition of a wide variety of polycarbonate plastic, flame retardants, dental

<p>QUICK RESPONSE CODE</p> 	<p>DOI: 10.13040/IJPSR.0975-8232.9(3).1256-62</p>
<p>Article can be accessed online on: www.ijpsr.com</p>	
<p>DOI link: http://dx.doi.org/10.13040/IJPSR.0975-8232.9(3).1256-62</p>	

sealant resins, and liners for food packaging^{20, 21}. It is a known endocrine disruptor is a monomer of polycarbonate plastic and a constituent of epoxy and polystyrene resins^{18, 22}. Food is considered as the main source of exposure to BPA as it leaches out from coatings^{16, 23}.

Various studies suggested that trace amounts of BPA also show estrogenic activity even at concentrations as low as 1 ng/l level²⁴⁻²⁶. Since a weak estrogen-like activity of BPA was reported by researchers, its effects on human health have become of emergent concern. BPA orally administered could easily cross the placental barrier and enter the fetus in animal experiment^{27, 28}.

Bisphenol A shows potential acute, short-term and sub-chronic toxicity²⁹⁻³¹. Studies have shown that BPA can cause injury in the liver, kidney, brain and other organs in rodents³²⁻³⁴. Moreover, BPA easily crosses the blood-brain and placental barriers; it can affect the developing nervous system in fetuses, infants and young children³⁵. Oral exposure to BPA has been reported to interfere with proliferation of the cell cycle in intermediate (neural) progenitor cells (IPC) and neurogenesis in developing neocortex³⁶. Therefore, the present study was designed to investigate the possible sub chronic toxic effect of BPA on testis of mice and mitigation of its toxicity by quercetin in testis of mice.

MATERIALS AND METHODS:

Chemicals: Quercetin and bisphenol A were acquired from Hi Media Laboratories Pvt. Ltd., Mumbai, India. Olive oil was obtained from Figaro, Madrid, Spain. All the other chemicals used in the study were of AR grade.

Experimental Animals: In this study, inbred healthy adult Swiss strain male albino mice weighing 35 - 40 gm were obtained from Cadila pharmaceutical Research Center, Ahmedabad, India. Animals were kept in the Animal House of Zoology Department of Gujarat University, Ahmedabad, India. They were housed in an air - conditioned room at a temperature of 25 ± 2 °C and 50 - 55% relative humidity with a 12 h light / dark cycle throughout the experiment. Animals were fed with certified pelleted rodent feed supplied by Amrut Feeds, Pranav Agro Industries Ltd., Pune,

India and potable water *ad libitum*. All the experimental protocols were approved by the Committee for the Purpose of Control and Supervision of Experiment on Animals (Reg. - 167/1999/CPCSEA), New Delhi, India. Animals were handled according to the guidelines published by Indian National Science Academy, New Delhi, India (1991).

Dose Selection: Different doses of BPA used for treatment were based on its LD₅₀ value³⁷ (240, 120 and 80 mg/kg bw/day respectively). Animals were treated orally for 45 days. For the amelioration of BPA induced toxicity, different doses of quercetin (30, 60 and 90 mg/kg bw/day) were selected based on the study of Sangai and Verma (2014)³⁸.

Experimental Design: Experimental Protocol is shown in **Table 1**. Mice were randomly divided into nine groups, each containing 10 animals. Treatment schedule of the animals was as follows. Animals from group I (untreated control) were kept untreated and given free access to feed and water. Group II (vehicle control) animals were administered with olive oil (0.2 ml/animal/day), which has been used as vehicle to dissolve bisphenol A. Group III (antidote control) animals were treated with quercetin (90 mg/kg bodyweight/day), which has been used for amelioration of BPA - induced toxicity.

Animals of group IV, V and VI received three different doses of bisphenol A (80, 120 and 240 mg/kg bw/day) for 45 days. Animals of group VII, VIII and IX received three different doses of quercetin (30, 60 and 90 mg/kg bw/day) along with high dose of BPA (240 mg/kg bw/day). All treatments were given orally using a feeding tube attached to hypodermic syringe for 45 days. Animals were sacrificed on 46th day by using mild anaesthesia and the testis was dissected out, blotted free from blood and used for biochemical analysis.

Biochemical Analysis:

Analysis of Energy Metabolism: The ATPase (E.C.3.6.1.3) activity in testis of control and all treated groups of animals were assayed by the method of Quinn and White (1968)³⁹; while inorganic phosphate liberated was estimated using the method of Fiske and Subbarow (1925)⁴⁰. The enzyme activity was expressed as μ moles

inorganic phosphate released/mg protein/30 min. The activity of SDH (E.C.1.3.99.1) was estimated in the testis of control and all treated groups of animals according to the method of Beatty *et al.*,

(1966)⁴¹. The enzyme activity was expressed as μg formazon formed/mg protein/15 min.

TABLE 1: EXPERIMENTAL PROTOCOL

Experimental groups	Numbers of animal	Duration (days)	Necropsy
Control groups			
1 Untreated control	10	45	46 th
2 Vehicle control (0.2 ml olive oil/animal/day)	10	45	46 th
3 Antidote control (90 mg quercetin/kg body weight/day)	10	45	46 th
BPA- treated groups			
4 BPA-low dose (80 mg/kg body weight/day)	10	45	46 th
5 BPA-medium dose (120mg /kg body weight/day)	10	45	46 th
6 BPA-high dose (240 mg/kg body weight/day)	10	45	46 th
BPA-HD + quercetin- treated groups			
7 BPA HD + quercetin (30 mg/kg body weight/day)	10	45	46 th
8 BPA HD + quercetin (60 mg/kg body weight/day)	10	45	46 th
9 BPA HD + quercetin (90 mg/kg body weight/day)	10	45	46 th

Phosphatases Activity: Acid phosphatase (E.C.3.1.3.2) and alkaline phosphatase (E.C. 3.1.3.1) activity was estimated in testis by the method of Bessey *et al.*, (1946)⁴². The phosphatases activity in testis was expressed as μmoles p-nitrophenol released/mg protein / 30 min.

Statistical Analysis: The results were expressed as the mean \pm SEM. The data were statistically analyzed using one way analysis of variance (ANOVA) followed by Tukey's post hoc test in Graph pad prism 6 (graph pad, software, USA). Statistically significance was accepted with $p < 0.05$. Correlation Coefficient was measured to estimate the strength of linear association between two Variables.

RESULTS: The effect of BPA on activities of enzymes in testis and its possible amelioration by pre-treatment with quercetin is shown in **Table 2**. No significant difference was observed in activities of enzymes in testis of different control groups of animals (Groups I - III). BPA treatment caused significant ($p < 0.05$) dose-dependent decrease in ATPase ($r = 0.926$) and SDH ($r = 0.962$) activities

as compared to untreated control. Maximum alteration observed was up to 48.70% and 55.44% for ATPase and SDH respectively in high dose BPA-treated animals (Group VI) compared to vehicle control (**Graph 1**). On the other hand, BPA treatment (Group IV - VI) caused significant ($p < 0.05$) increase in ACP (BPA-LD: 40.68%, BPA-MD: 102.98%, BPA-HD: 179.77%) and ALP (BPA-LD: 72.54%, BPA-MD: 173.87%, BPA - HD: 327.83%) activities in testis of BPA - treated mice which was dose-dependent ($r = 0.899, 0.938$ respectively) (**Graph 2**). Oral administration of quercetin treatment along with BPA-HD caused significant ($p < 0.05$), dose-dependent increase in ATPase ($r = 0.972$) and SDH ($r = 0.926$) activities. In all these enzyme activities, maximum amelioration was observed in 90 mg/kg bw/day dose of quercetin along with BPA - HD as per calculated by organoprotective index (**Table 2**). Similarly, Oral administration of quercetin along with BPA-HD (Group VII - IX) caused significant ($p < 0.05$) decrease in ACP and ALP activities as compared to BPA - HD (Group VI) which was dose-dependent ($r = 0.886, 0.885$ respectively).

TABLE 2: AMELIORATIVE EFFECT OF BPA-INDUCED CHANGES ON LIPID PEROXIDATION AND ENZYMATIC ANTIOXIDANTS IN TESTIS OF MICE

S. no.	Experimental group	Energy metabolism		Phosphatase	
		ATPase	SDH	ACP	ALP
Control groups					
1	Untreated control	2.505±0.032	23.143±0.195	1.247±0.085	0.634±0.034
2	Vehicle control (0.2 ml olive oil / animal / day)	2.500±0.071	23.066±0.504	1.253±0.149	0.622±0.036
3	Antidote control (90 mg quercetin / animal / day)	2.480±0.044	22.979±0.113	1.257±0.055	0.637±0.012
BPA-treated groups					
4	BPA-Low dose (80mg/kg bodyweight/day)	2.124±0.041 ^a (15.03)	17.874±0.297 ^a (22.51)	1.763±0.071 ^a (40.68)	1.073±0.034 ^a (72.54)
5	BPA-Medium dose (120 mg/kg bodyweight/day)	1.586±0.063 ^a (36.54)	13.848±0.481 ^a (39.96)	2.543±0.097 ^a (102.98)	1.703±0.052 ^a (173.87)
6	BPA-High dose (240 mg/kg bodyweight/day)	1.282±0.040 ^a (48.70)	10.278±0.257 ^a (55.44)	3.505±0.138 ^a (179.77)	2.661±0.120 ^a (327.83)
HD BPA (240 mg/kg body weight) + quercetin treated groups					
7	BPA-HD + quercetin (30 mg/kg body weight/day)	1.543±0.022 ^b (21.39)	14.815±0.515 ^b (35.48)	3.062±0.039 ^b (19.66)	2.107±0.111 ^b (27.16)
8	BPA-HD + quercetin (60 mg/kg body weight/day)	2.066±0.026 ^b (64.35)	17.358±0.468 ^b (55.37)	2.371±0.078 ^b (50.36)	1.491±0.039 ^b (57.36)
9	BPA-HD + quercetin (90 mg/kg body weight/day)	2.388±0.017 ^b (90.84)	21.196±0.546 ^b (85.38)	1.513±0.096 ^b (88.45)	0.780±0.022 ^b (92.24)

Values are mean ± SEM; n = 10.

Level of significance: ^ap < 0.05, as compared to vehicle control; ^bp < 0.05, as compared to BPA-HD-treated.

No significant difference was noted between different control groups (Groups I - III).

Units: ACP- μ moles p-nitrophenol released/mg protein / 30 min; ALP- μ moles p-nitrophenol released / mg protein / 30 min; ATPase - μ moles inorganic phosphate released/mg protein / 30 min; SDH- μg formazon formed/mg protein / 15 min.

DISCUSSION: Bisphenol A is well-known xenoestrogenic chemical released in environment. Earlier studies have revealed that BPA treatment caused significant decrease bodyweights as well as testis weight ⁴³. Histopathological alteration was found in testis of BPA treated mice testis ⁴⁴. The reproductive toxicity of BPA is caused by interaction with androgen and estrogen receptors ⁴⁵, ⁴⁶. Testicular toxicity of BPA has been reported by Takahashi and Oishi (2003) ⁴⁷.

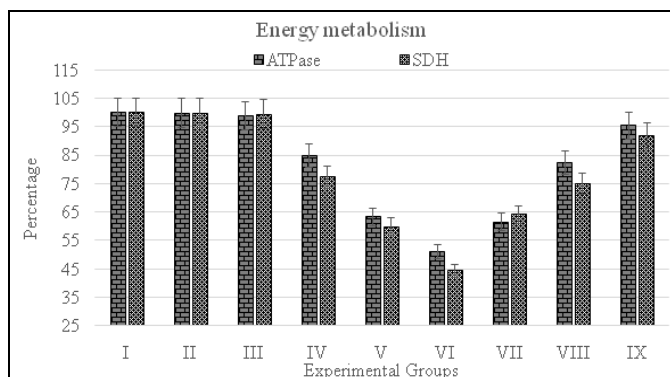
In present study we have administered Bisphenol A for 45 days to study sub-chronic exposure. We studied that Bisphenol A reduces the energy metabolism by reducing the activities of enzymes involved in energy metabolism. In present study we studied that BPA treatment for 45 days caused significant, dose-dependent decrease in ATPase and SDH activities in testis of mice. Mitochondria are the targets of several environmental toxicants including environmental estrogens ⁴⁸. SDH is a key enzyme of mitochondrial Krebs cycle and it is mainly concerned with aerobic oxidation of acetyl coA and generation of ATP. Among Krebs cycle dehydrogenases, SDH is very active than any other

enzyme ⁴⁹. ATPase is required for enzymatic hydrolysis of ATP which is important for intracellular transfer of energy. Reduction in ATPase and SDH activity could be due to alteration in mitochondria. A study by Nakagawa and Tayama ⁵⁰, revealed that the incubation of hepatocytes with BPA elicited a concentration and time-dependent cell death, accompanied by loss of intracellular ATP and total adenine nucleotide pools. Co-administration of quercetin along with BPA significantly increased ATPase and SDH activities in tissues might be preventing mitochondrial dysfunction from ROS due to its antioxidative property. It has been previously reported that quercetin potentially reduce BPA-induced oxidative stress and mitochondrial dysfunction in mice ^{51, 52}.

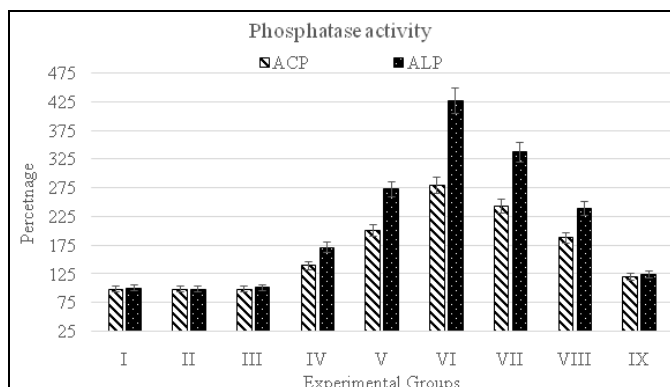
Acid phosphatase is a marker enzyme for the lysosomal integrity and important for the tissue reorganization and tissue repair ⁵³. Oral administration of BPA for 45 days caused significant dose - dependent increase in ACP activity in testis of mice. This might be due to increased release of lysosomal enzyme by

damaging lysosomal integrity causing lysis of cell. Activity of ALP was also significantly increased by BPA treatment in testis of mice. Alkaline phosphatase is a marker enzyme for plasma and endoplasmic reticulum. Bisphenol A treatment causes alteration in endoplasmic reticulum^{45 - 56}. This could be the reason for increasing ALP activity. However, quercetin treatment significantly decreased ACP and ALP activities by preventing damage to the tissue⁵⁷.

Different concentrations of quercetin caused significant reduction in ACP and ALP activities as well as increase the activities of ATPase and SOD and restored these levels close to corresponding control values. Quercetin increases the body's endogenous antioxidants to reduce oxidative damage. The anti-oxidant properties are largely a function of the chemical structure of quercetin, particularly the presence and location of the hydroxyl (-OH) substitutions and the catechol-type B-ring^{58 - 62}. Similar investigations have also been reported antioxidant properties of quercetin^{63 - 65}.



GRAPH 1: SHOWING THE PHOSPHATASE ACTIVITY (%) ATPase - Adenosine triphosphatase; SDH - Succinate dehydrogenase



GRAPH 2: SHOWING THE PHOSPHATASE ACTIVITY (%) ACP - Acid phosphatase; ALP - Alkaline phosphatase

CONCLUSION: Present study revealed that treatment of BPA for 45 days causes lysosomal and mitochondrial enzymatic changes in experimental animals by disturbing the balance between ROS and antioxidant defenses system in testis. However co-administration of quercetin for 45 days caused significant amelioration in enzymatic activities. This study concludes that bisphenol A causes toxicity in testis of mice by disturbing mitochondrial and lysosomal integrity; it can be mitigated by the quercetin.

ACKNOWLEDGEMENT: Nil

CONFLICT OF INTEREST: The authors do not have any conflict of interest.

REFERENCES:

1. Bravo L: Polyphenols: Chemistry, dietary sources, metabolism, and nutritional significance. *Nutrition reviews* 1998; 56(11): 317-33.
2. Alrawaiq NS and Abdullah A: A review of flavonoid quercetin: metabolism, bioactivity and antioxidant properties. *International Journal of Pharm Tech Research* 2014; 6(3): 933-41.
3. Zhang H and Tsao R: Dietary polyphenols, oxidative stress and antioxidant and anti-inflammatory effects. *Current Opinion in Food Science* 2016; 8: 33-42.
4. Moretti E, Mazzi L, Terzuoli G, Bonechi C, Iacoponi F, Martini S, Rossi C and Collodel G: Effect of quercetin, rutin, naringenin and epicatechin on lipid peroxidation induced in human sperm. *Reproductive Toxicology* 2012; 34(4): 651-7.
5. Coballase-Urrutia E, Pedraza-Chaverri J, Cárdenas-Rodríguez N, Huerta-Gertrudis B, García-Cruz ME, Montesinos-Correa H, Sánchez-González DJ, Camacho-Carranza R and Espinosa-Aguirre JJ: Acetonic and Methanolic Extracts of *Heterotheca inuloides*, and Quercetin, Decrease CCl₄-Oxidative Stress in Several Rat Tissues. *Evidence-Based Complementary and Alternative Medicine* 2013.
6. Choi GN, Kim JH, Kwak JH, Jeong CH, Jeong HR, Lee U and Heo HJ: Effect of quercetin on learning and memory performance in ICR mice under neurotoxic trimethyltin exposure. *Food Chemistry* 2012; 132(2): 1019-24.
7. Haleagrahara N, Siew CJ and Ponnusamy K: Effect of quercetin and desferrioxamine on 6-hydroxydopamine (6-OHDA) induced neurotoxicity in striatum of rats. *The Journal of toxicological sciences* 2013; 38(1): 25-33.
8. Gibellini L, Pinti M, Nasi M, Montagna JP, De Biasi S, Roat E, Bertocelli L, Cooper EL and Cossarizza A: Quercetin and cancer chemoprevention. *Evidence-Based Complementary Alternative Medicine* 15 (Article no. 591356), <http://dx.doi.org/10.1093/ecam/nea053> 2011.
9. Baghel SS, Shrivastava N, Baghel RS, Agrawal P and Rajput S: A review of quercetin: antioxidant and anticancer properties. *World J Pharm Pharmaceutical Sci.* 2012; 1(1): 146-60.
10. Russo M, Spagnuolo C, Tedesco I, Bilotto S and Russo GL: The flavonoid quercetin in disease prevention and

- therapy: facts and fancies. *Biochemical pharmacology* 2012; 83(1): 6-15.
11. Larson AJ, Symons JD and Jalili T: Therapeutic potential of quercetin to decrease blood pressure: review of efficacy and mechanisms. *Advances in Nutrition: An International Review Journal* 2012; 3(1): 39-46.
 12. Lin CF, Leu YL, Al-Suwayeh SA, Ku MC, Hwang TL and Fang JY: Anti-inflammatory activity and percutaneous absorption of quercetin and its polymethoxylated compound and glycosides: the relationships to chemical structures. *European Journal of Pharmaceutical Sciences* 2012; 47(5): 857-64.
 13. Ying HZ, Liu YH, Yu B, Wang ZY, Zang JN and Yu CH: Dietary quercetin ameliorates nonalcoholic steatohepatitis induced by a high-fat diet in gerbils. *Food and Chemical Toxicology* 2013; 52: 53-60.
 14. Lekić N, Canová NK, Hofínek A and Farghali H: The involvement of hemeoxygenase 1 but not nitric oxide synthase 2 in a hepatoprotective action of quercetin in lipopolysaccharide-induced hepatotoxicity of D-galactosamine sensitized rats. *Fitoterapia* 2013; 87: 20-6.
 15. Chapin RE, Adams J, Boekelheide K, Gray LE, Hayward SW, Lees PS, McIntyre BS, Portier KM, Schnorr TM, Selevan SG and Vandenberg JG: NTP-CERHR expert panel report on the reproductive and developmental toxicity of bisphenol A. *Birth Defects Research Part B: Developmental and Reproductive Toxicology* 2008; 83(3): 157-395.
 16. Tyl RW, Myers CB, Marr MC, Thomas BF, Keimowitz AR, Brine DR, Veselica MM, Fail PA, Chang TY, Seely JC and Joiner RL: Three-generation reproductive toxicity study of dietary bisphenol A in CD Sprague-Dawley rats. *Toxicological Sciences* 2002; 68(1): 121-46.
 17. Fernandez MF, Arrebola JP, Taoufiki J, Navalón A, Ballesteros O, Pulgar R, Vilchez JL and Olea N: Bisphenol-A and chlorinated derivatives in adipose tissue of women. *Reproductive toxicology* 2007; 24(2): 259-64.
 18. Huang YQ, Wong CK, Zheng JS, Bouwman H, Barra R, Wahlström B, Neretin L and Wong MH: Bisphenol A (BPA) in China: a review of sources, environmental levels, and potential human health impacts. *Environment international* 2012; 42: 91-9.
 19. Ranjit N, Siefert K and Padmanabhan V: Bisphenol-A and disparities in birth outcomes: a review and directions for future research. *Journal of perinatology: official journal of the California Perinatal Association* 2010; 30(1): 2.
 20. WHO: Food and Agriculture Organization of the United Nations, 2011. *Toxicological and Health Aspects of Bisphenol A: Report of Joint FAO/WHO Expert Meeting 2-5 November 2010 and Report of Stakeholder Meeting on Bisphenol A*. World Health Organization Press, Geneva 2010.
 21. Bae B, Jeong JH and Lee SJ: The quantification and characterization of endocrine disruptor bisphenol-A leaching from epoxy resin. *Water Science and Technology* 2002; 46(11-12): 381-7.
 22. Willhite CC, Ball GL and McLellan CJ: Derivation of a Bisphenol a Oral Reference Dose (RfD) and Drinking-Water Equivalent Concentration. *Journal of Toxicology and Environmental Health, Part B* 2008; 11(2): 69-146.
 23. Doerge DR and Fisher JW: Background paper on metabolism and toxicokinetics of bisphenol A: FAO/WHO expert meeting on bisphenol A (BPA).
 24. Kang JH, Kito K and Kondo F: Factors influencing the migration of bisphenol A from cans. *Journal of food protection* 2003; 66(8): 1444-7.
 25. Vom Saal FS, Cooke PS, Buchanan DL, Palanza P, Thayer KA, Nagel SC, Parmigiani S and Welshons WV: A physiologically based approach to the study of bisphenol A and other estrogenic chemicals on the size of reproductive organs, daily sperm production, and behavior. *Toxicology and Industrial Health* 1998; 14(1-2): 239-60.
 26. Fisher JS, Turner KJ, Brown D and Sharpe RM: Effect of neonatal exposure to estrogenic compounds on development of the excurrent ducts of the rat testis through puberty to adulthood. *Environmental Health Perspectives*. 1999; 107(5): 397.
 27. Steinmetz R, Brown NG, Allen DL, Bigsby RM and Ben-Jonathan N: The environmental estrogen bisphenol A stimulates prolactin release *in vitro* and *in vivo*. *Endocrinology* 1997; 138(5): 1780-6.
 28. Krishnan AV, Stathis P, Permuth SF, Tokes L and Feldman D: Bisphenol-A: an estrogenic substance is released from polycarbonate flasks during autoclaving. *Endocrinology* 1993; 132(6): 2279-86.
 29. Miyakoda H, Tabata M, Onodera S and Takeda K: Comparison of conjugative activity, conversion of bisphenol A to bisphenol A glucuronide, in fetal and mature male rat. *Journal of Health Science* 2000; 46(4): 269-74.
 30. Lin CY, Shen FY, Lian GW, Chien KL, Sung FC, Chen PC and Su TC: Association between levels of serum bisphenol A, a potentially harmful chemical in plastic containers, and carotid artery intima-media thickness in adolescents and young adults. *Atherosclerosis* 2015; 241(2): 657-63.
 31. Taepongsoat L, Tangpraputgul P, Kitana N and Malaivijitnond: Stimulating effects of quercetin on sperm quality and reproductive organs in adult male rats. *Asian journal of andrology* 2008; 10(2): 249-58.
 32. Tyl RW: Commentary to the CERHR expert panel report on bisphenol A. *Birth Defects Research Part B: Developmental and Reproductive Toxicology* 2008; 83(3): 152-.
 33. Sax NI and Lewis RJ: *Dangerous properties of industrial chemicals*. Toxicology of Phenol, New York 1975; 458-1008.
 34. Bindhumol V, Chitra KC and Mathur PP: Bisphenol A induces reactive oxygen species generation in the liver of male rats. *Toxicology* 2003; 188(2): 117-24.
 35. Kabuto H, Amakawa M and Shishibori T: Exposure to bisphenol A during embryonic/fetal life and infancy increases oxidative injury and causes underdevelopment of the brain and testis in mice. *Life sciences* 2004; 74(24): 2931-40.
 36. Braun JM, Kalkbrenner AE, Calafat AM, Yolton K, Ye X, Dietrich KN and Lanphear BP: Impact of early-life bisphenol A exposure on behavior and executive function in children. *Pediatrics* 2011; 128(5): 873-82.
 37. Komada M, Asai Y, Morii M, Matsuki M, Sato M and Nagao T: Maternal bisphenol A oral dosing relates to the acceleration of neurogenesis in the developing neocortex of mouse fetuses. *Toxicology* 2012; 295(1): 31-8.
 38. Hussein RM and Eid JI: Pathological mechanisms of liver injury caused by oral administration of bisphenol A. *Life Science Journal* 2013; 10(1).
 39. Sangai NP, Verma RJ and Trivedi MH: Testing the efficacy of quercetin in mitigating bisphenol A toxicity in liver and kidney of mice. *Toxicology and industrial health* 2014; 30(7): 581-97.
 40. Quinn PJ and White IG: Distribution of adenosinetriphosphatase activity in ram and bull

- spermatozoa. Journal of reproduction and fertility 1968; 15(3): 449-52.
41. Fiske CH and Subbarow Y: The colorimetric determination of phosphorus. J. Biol. Chem 1925; 66(2): 375-400.
 42. Beatty CH, Basinger GM, Dully CC and Bocek RM: Comparison of red and white voluntary skeletal muscles of several species of primates. Journal of Histochemistry and Cytochemistry 1966; 14(8): 590-600.
 43. Bessey OA, Lowky OH and Brock MJ: A method for the rapid determination of alkaline phosphatase with five cubic millimeters of serum. Journal of Biological Chemistry 1946; 164: 321-9.
 44. Sakae M, Ohsako S, Ishimura R, Kurosawa S, Kurohmaru M, Hayashi Y, Aoki Y, Yonemoto J and Tohyama C: Bisphenol-A affects spermatogenesis in the adult rat even at a low dose. Journal of occupational health 2001; 43(4): 185-90.
 45. Tan BL, Kassim NM and Mohd MA: Assessment of pubertal development in juvenile male rats after sub-acute exposure to bisphenol A and nonylphenol. Toxicology letters 2003; 143(3): 261-70.
 46. Tohmé M, Prud'homme SM, Boulahtouf A, Samarut E, Brunet F, Bernard L, Bourguet W, Gibert Y, Balaguer P and Laudet V: Estrogen-related receptor γ is an *in vivo* receptor of bisphenol A. The FASEB Journal 2014; 28(7): 3124-33.
 47. Matthews JB, Twomey K and Zacharewski TR: *In vitro* and *in vivo* interactions of bisphenol A and its metabolite, bisphenol A glucuronide, with estrogen receptors α and β . Chemical research in toxicology 2001; 14(2): 149-57.
 48. Takahashi O and Oishi S: Testicular toxicity of dietarily or parenterally administered bisphenol A in rats and mice. Food and chemical toxicology 2003; 41(7): 1035-44.
 49. Michelangeli F, Ogunbayo OA, Wootton LL, Lai PF, Al-Mousa F, Harris RM, Waring RH and Kirk CJ: Endocrine disrupting alkylphenols: structural requirements for their adverse effects on Ca^{2+} pumps, Ca^{2+} homeostasis and Sertoli TM4 cell viability. Chemico-biological interactions 2008; 176(2): 220-6.
 50. Racker E: Mechanisms in bioenergetics. Academic Press; 2014.
 51. Nakagawa Y and Tayama S: Metabolism and cytotoxicity of bisphenol A and other bisphenols in isolated rat hepatocytes. Archives of toxicology 2000; 74(2): 99-105.
 52. Moosmann B and Behl C: The antioxidant neuroprotective effects of estrogens and phenolic compounds are independent from their estrogenic properties. Proceedings of the National Academy of Sciences 1999; 96(16): 8867-72.
 53. Sangai NP and Verma RJ: Quercetin ameliorates bisphenol A-induced toxicity in mice. ActaPoloniae Pharmaceutical Drug Research 2012; 69(3): 557-63.
 54. Collins AJ and Lewis DA: Lysosomal enzyme levels in the blood of arthritic rats. Biochemical pharmacology 1971; 20(1): 251-3.
 55. Wright PJ and Plummer DT: The use of urinary enzyme measurements to detect renal damage caused by nephrotoxic compounds. Biochemical pharmacology 1974; 23(1): 65-73.
 56. Shahjahan M, Sabitha KE, Jainu M and Devi CS: Effect of *Solanum trilobatum* against carbon tetrachloride induced hepatic damage in albino rats. Indian Journal of Medical Research 2004; 120(3): 194.
 57. Gong Y, Wu J, Huang Y, Shen S and Han X: Nonylphenol induces apoptosis in rat testicular Sertoli cells *via* endoplasmic reticulum stress. Toxicology letters 2009; 186(2): 84-95.
 58. Williams C, Bondesson M, Kremontsov DN and Teuscher C: Gestational bisphenol A exposure and testis development. Endocrine disruptors 2014; 2(1): e29088.
 59. Rice-Evans C, Miller N and Paganga G: Antioxidant properties of phenolic compounds. Trends in plant science 1997; 2(4): 152-9.
 60. Moon YJ, Wang X and Morris ME: Dietary flavonoids: effects on xenobiotic and carcinogen metabolism. Toxicology *in vitro* 2006; 20(2): 187-210.
 61. Okamoto T: Safety of quercetin for clinical application. International journal of molecular medicine 2005; 16(2): 275-8.
 62. Kressler J, Millard-Stafford M and Warren GL: Quercetin and endurance exercise capacity: a systematic review and meta-analysis. Medicine and Science in Sports and Exercise 2011; 43(12): 2396-404.
 63. Luo M, Yang X, Hu JY, Jiao J, Mu FS, Song ZY, Gai QY, Qiao Q, Ruan X and Fu YJ: Antioxidant properties of phenolic compounds in renewable parts of *Crataegus pinnatifida* inferred from seasonal variations. Journal of food science 2016; 81(5).
 64. Moskaug JO, Carlsen H, Myhrstad M and Blomhoff R: Molecular imaging of the biological effects of quercetin and quercetin-rich foods. Mechanisms of ageing and development 2004; 125(4): 315-24.
 65. Jahan S, Ain QU and Ullah H: Therapeutic effects of quercetin against bisphenol A induced testicular damage in male Sprague Dawley rats. Systems biology in reproductive medicine 2016; 62(2): 114-24.
 66. Wahby MM, Abdallah ZM, Abdou HM, Yousef MI and Newairy AS: Mitigating potential of *Ginkgo biloba* extract and melatonin against hepatic and nephrotoxicity induced by Bisphenol A in male rats. Egyptian Journal of Basic and Applied Sciences 2017.

How to cite this article:

Samova S, Doctor H and Verma R: Protective effect of quercetin on Bisphenol A - induced enzymatic changes in testis of mice. Int J Pharm Sci & Res 2018; 9(3): 1256-62. doi: 10.13040/IJPSR.0975-8232.9(3).1256-62.

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