IJPSR (2014), Volume 5, Issue 9



INTERNATIONAL JOURNAL



Received on 04 March 2014; received in revised form, 15 April 2014; accepted, 07 June 2014; published 01 September 2014

ANTICANCER POTENTIAL OF SOLANUM JASMINOIDES

O. Porwal^{1, *4}, M. J. Nanjan¹, M. J. N. Chandrasekar², R. Srinivasan² and S. Gupta³

TIFAC CORE in Herbal Drugs¹, Department of Pharmaceutical Chemistry's², JSS College of Pharmacy, Ootacamund - 643001, Tamil Nadu, India.

Department of Pharmacology ³, Indore Institute of Pharmacy, Indore - 453331, Madhya Pradesh, India. National Institute of Pharmaceutical Education and Research (NIPER-A)⁴, Ahmedabad - 380054, Gujarat, India.

Keywords:

Solanum jasminoides, Solanaceae, Cytotoxicity, Anticancer

Correspondence to Author: Omji Porwal

Scientific Assistant, Natural Product, National Institute of Pharmaceutical Education and Research (NIPER-A), Ahmedabad - 380054, Gujarat, India.

E-mail: omporwal@gmail.com

ABSTRACT: Objective: To investigate the anticancer potential of the extracts and the alkaloidal and nonalkaloidal fractions of Solanum jasminoides Paxt using in-vitro and in-vivo models. Method: The petroleum ether, methanol, and water extracts of the whole plant were prepared by successive soxhlet method. The methanol extract was treated with solvents to separate alkaloidal and non-alkaloidal fractions. All the samples were subjected to in-vitro anticancer studies using Vero, BRL-3, HEp-2, HeLa, MDCK, and A-549 cell lines. Based on the in-vitro results, the alkaloidal fraction was selected and subjected to in-vivo anticancer activity against DLA and EAC induced ascities in mice. Results: The results reveal that the extracts and the fractions show moderate to good *in-vitro* anticancer activity, and the alkaloidal fraction shows superior activity. In the in-vivo study, the alkaloidal fraction shows a dose dependent effect on changes in the body weight, mortality, and clinical symptoms induced by cancer cells. Conclusion: The results of the present study indicate that the alkaloidal fraction of the plant possesses good, both in-vitro and in-vivo, anticancer potential.

INTRODUCTION: Considerable efforts are being made to identify naturally occurring anticancer agents who would prevent, slow and reverse cancer induction and its subsequent development ¹. Plants contain abundant quantities of these compounds, and several plant-derived compounds are currently employed successfully in treating cancer. A major group of these compounds is the powerful antioxidants. And the rest include reactive groups that confer the protective properties.



Anticancer agents from plants currently in clinical use can be categorized into four main classes, namely *Vinca* alkaloids, epipodophyllotoxins, taxanes and camptothecin². The plant *Solanum jasminoides* Paxt (Family: Solanaceae) is a native of South America, introduced and found in the roadsides and waste places in the tempest regions of the Nilgiris, Tamil Nadu, India, at 1500-2500 m altitude regions. The leaves and fruits contain glycoalkaloids³. The present study was undertaken to investigate the anticancer potential of successive extracts and the isolated fractions of the plant.

MATERIALS AND METHODS:

Plant Material: The whole plant was collected from in and around the Nilgiris forests, Tamilnadu, India. In June 2004, and authenticated by the Survey of Medicinal Plants and Collection Unit,

Govt. of India, Ootacamund, India. A voucher specimen (TIFAC 03) has been deposited for future reference at the college herbarium.

Chemicals: 3-(4,5 Dimethyl thiazole-2-yl)-5diphenyl tetrazolium bromide (MTT), and sulphorhodamine B (SRB) were obtained from Sigma–Aldrich Co., St. Louis, USA. 5-Fluorouracil (purity≥99%) was obtained from Ranbaxy Ltd., Gurgaon, India. All other chemicals used in the study were of analytical grade and purchased from E-Merck, Ltd., Mumbai, India.

Animals: Healthy adult Swiss albino mice weighing (25-30 g) were obtained from the animal house, J.S.S. College of Pharmacy, Ootacamund, India, and maintained under standard environmental conditions (22–28 °C, 60–70% relative humidity, 12-h dark: 12-h light cycle). They were fed with standard rat feed (M/S Hindustan Lever Ltd., Bangalore, India) and water *ad libitum*. The animals were housed in large cages.

The experiments were conducted with prior approval of the Institutional Animal Ethics Committee (approval no. JSSCP/IAEC/Ph.D/PH. Chemistry/01/2005–2006).

Extraction Procedure and Phytochemical Analysis: The whole plant was chopped to small pieces and dried in the shade. The dried sample (100 g) was extracted successively with 600 mL each of petroleum ether (60–80 °C), methanol and water in a Soxhlet extractor for 18–20 h. The successive petroleum ether, methanol, and water extracts will henceforth be called SJP, SJM, SJW, respectively. All the extracts were subjected to qualitative phytochemical analysis using standard tests ⁴.

Isolation of Total Alkaloids from SJM: The *Solanaceae* family is known to possess glycoalkaloids ³. The total alkaloidal fraction was, therefore, isolated from SJM. The SJM was treated with hydrochloric acid (3%, HCl) and kept overnight. It was then filtered through a Whatman filter paper, and the next day the residue was washed several times with 3% HCl (100 ml). The filtrate was extracted with chloroform (300 ml, 4 times), and the chloroform extracts were combined and dried (non alkaloid fraction). The acid layer was treated with ammonium hydroxide solution

(50%) to make it alkaline (>pH 10). The precipitated alkaloids were further extracted in chloroform (250 ml each, 5 times). The chloroform extracts were combined, filtered, and concentrated to dryness under reduced pressure and controlled temperature (40–50 °C). A pale yellow powder (0.5312 g, 0.53 %) was obtained (alkaloid fraction). Henceforth, the total alkaloidal and non-alkaloidal fraction of SJM will be called as SJM-A and SJM-NA, respectively.

Cell Lines and Culture Medium: Vero (normal fibroblast-like African green monkey cell), BRL-3 (normal fibroblast-like rat liver cell) and HEp-2(human laryngeal epithelial carcinoma) cell cultures were procured from National Centre for Cell Sciences (NCCS), Pune, India. HeLa (Epithelial human cervix cancer) and MDCK (dog kidney) cell lines of both normal and cancerous cell lines were used for the comparison purposes, are procured from V.P. Chest Institute, New Delhi, India. A-549 (human non-small cell lung carcinoma) cells were obtained from Christian Medical College, Vellore, India. Stock cells of all cell lines were cultured in DMEM supplemented with 10% inactivated FBS/NBCS, penicillin (100 streptomycin IU/mL), $(100 \mu g/mL)$ and amphotericin B (5µg/mL) in a humidified atmosphere of 5% CO₂ at 37 °C until confluent.

The cells were dissociated with TPVG solution (0.2% trypsin, 0.02% EDTA, 0.05% glucose in PBS). The stock cultures were grown in 25 cm² tissue culture flasks and all the experiments were carried out in 96 well microtitre plates. DLA and EAC cells were procured from Amla Cancer Research center, Thrissur, Kerala, India. The cells were maintained *in-vivo* in Swiss albino mice by intraperitoneal transplantation. EAC cells aspirated from the peritoneal cavity of mice were washed with saline and given intraperitoneally to develop an ascitic tumor.

Preparation of Suspension and Solutions: For *invitro* studies each extract (10mg) was separately dissolved in 1 mL of dimethyl sulphoxide (DMSO), and the volume was made up to 10 mL with maintenance medium to obtain a stock solution of 1 mg/mL concentration and sterilized by filtration. Further dilutions were made from the stock and used for short and long term cytotoxicity studies. For *in-vivo* studies SJP, SJM, SJW, SJM-A, SJM-NA, and standard 5-FU were suspended in sodium carboxymethyl-cellulose (CMC, 0.3%) and administered orally to the animals with the help of an intragastric catheter.

Cytotoxicity Studies: The cytotoxicity assay was carried out using 0.1 mL of cell suspension, containing 10,000 cells seeded in each well of a 96 well microtitre plate (Tarson India Pvt., Ltd., Kolkata). Fresh medium containing different concentrations of the test sample was added after 24 h of seeding. Control cells were incubated without the test sample. The microtitre plate was incubated at 37 °C in a humidified incubator with 5% CO₂ for a period of 72 h. Sixteen wells were used for each concentration of the test sample. The CTC₅₀ was determined by the standard MTT ⁵ and SRB assays ⁶.

Short Term Cytotoxicity Studies: The short term toxicity studies were carried out against DLA and EAC cells using standard procedures 7 . The long term toxicity study was carried out by the method of Freshney⁸. A series of cultures were prepared in 25 cm^2 tissue culture flasks, three for each concentration and three control of seeding 2×10^5 cells in 4 mL of growth medium and incubating at 37 °C for 48 h. Long term survival studies against normal (Vero) and cancer (HEp-2) cell cultures were carried out, and the regeneration capacity of the cells was estimated by standard clonogenic assay. Three nontoxic concentrations of each extract were tested for their reduction in regenerative capacity, in comparison with untreated cell control. Among all the tested samples, SJM-A showed well *in-vitro* cytotoxicity against all cancer cells in-vitro. SJM-A was, therefore, selected to study it's in-vivo anticancer activity in different tumor models.

Effect of SJM-A on Mice Bearing DLA Cells Swiss Albino mice were divided into five groups with six animals in each group. All the animals were injected with DLA cells (1×10^6) intraperitoneally except the normal group. This was taken as day zero. Group, I served as normal control and group II as tumor control. These two groups received sodium CMC suspension (0.3%) administered orally. Group III served as a positive control and was treated with standard 5Fluorouracil (5-FU) at 20 mg/kg per oral route. Group IV and V were treated with SJM-A at a dose of 25 and 100 mg/kg per oral route, respectively. All these treatments were given 24 h after the DLA cells were inoculated, once daily for 10 days. The clinical signs, mortality, and body weight changes were recorded throughout the study period.

Effect of SJM-A on Mice Bearing EAC Cells: Swiss Albino mice were divided into five groups, with six animals in each group. All the animals were injected with EAC cells (2×10^6) intraperitoneally except the normal group. Group, I served as normal control and group II as tumor control. These two groups received sodium CMC suspension (0.3%), orally. Group III served as a positive control and was treated with standard, 5-Fluorouracil (5-FU) at 20 mg/kg p.o. Group IV and V were treated with SJM-A at a dose of 25 and 100 mg/kg p.o., respectively. All these treatments were given 24 h after the tumor inoculation, once daily for 10 days. The clinical signs, mortality, and body weight changes were recorded throughout the study period.

Statistical Analysis: The *in-vivo* data was analyzed by One-Way Analysis of Variance (ANOVA) followed by Turkey-Kramer multiple comparison tests, and p<0.05 was considered significant.

RESULTS: Qualitative phytochemical analysis revealed that SJP contains steroids, tannins, and fatty acid. SJM contains alkaloids, carbohydrates, flavonoids, glycosides, tannins, and triterpenoids. SJW contains carbohydrates, glycosides, and tannins. The results of the *in-vitro* cytotoxicity studies are shown in **Table 1**.

The results reveal that among the samples tested SJM-A shows significant cytotoxicity with CTC_{50} value 64.84, 71.28, 78.16, 111.29, 116.62, 132.84 µg/mL towards A-549, HEp-2, BRL-3A, HeLa, Vero, and MDCK, respectively. The results of the *in-vitro* short term toxicity studies are shown in **Table 2**.

The data reveal that SJM-A has moderate toxicity against DLA and EAC cells, whereas others do not show any significant toxicity even at higher concentrations. The results of the long term survival studies are shown in **Table 3**. The data reveal that in Vero cell culture, treatment with 100 μ g/ml of SJM-A for 72 h, the total number of colonies reduces to 14 ± 0.11 when compared to cell control (62 ± 3.25), but treatment with 100 μ g/mL of SJM-NA the total number of colonies reduces to only 21 ± 1.83. In HEp-2 cell culture. After 72 h of treatment with 50 μ g/mL of SJM-A, the total number of colonies reduces to 16 ± 0.11 when compared to cell control (63 ± 2.60), but

treatment with SJM-NA the total number of colonies reduces to only 19 ± 1.83 . SJM-A thus shows good cytotoxicity against all cancer cells cultures. SJM-A also shows moderate activity in long term toxicity studies by clonogenic assay. SJM-A was, therefore, selected for *in-vivo* anticancer activity in different tumor models.

TABLE 1: CYTOTOXIC ACTIVITY OF SJP, SJM, SJW, SJM-A, AND SJM-NA AGAINST NORMAL AND CANCER CELL CULTURES

Cell	SJP				SJM			SJW			SJM-A			SJM-NA		
Cultures	MTT	SRB	Avg.	MTT	SRB	Avg.	MTT	SRB	Avg.	MTT	SRB	Avg.	MTT	SRB	Avg.	
Vero	656.58	595.60	626.09	325.34	345.01	335.18	332.85	349.46	341.16	123.12	110.11	116.62	183.32	191.63	187.48	
	± 19.29	± 13.50		± 14.38	± 20.45		± 16.34	± 17.76		±5.37	± 4.93		±6.26	± 7.27		
BRL-3A	589.32	565.00	577.16	297.92	304.8	301.37	165.35	173.46	169.41	81.56	74.75	78.16	182.23	176.22	179.23	
	± 12.64	±7.56		±13.17	± 10.87		±7.36	± 8.76		±3.16	±2.27		±5.14	± 5.24		
MDCK	472.00	501.85	486.92	245.91	271.82	258.86	172.28	181.91	177.10	124.35	141.32	132.84	166.49	166.19	166.34	
	±3.56	± 18.65		± 10.14	± 14.78		± 8.42	± 7.21		±4.37	±5.95		±4.32	± 7.44		
HeLa	512.85	494.76	503.80	132.83	123.86	126.89	165.23	156.93	161.08	113.33	109.26	111.29	148.16	151.20	149.68	
	±6.89	± 12.50		±4.39	±5.72		±7.13	± 6.74		±5.36	± 4.57		±6.37	± 7.27		
HEp-2	456.80	479.2	468.02	133.49	125.40	129.46	153.36	162.23	157.79	76.29	66.27	71.28	90.93	102.89	96.91	
	±11.58	±6.72		±4.28	±5.26		±7.23	±6.86		±3.42	±3.45		±3.92	± 4.11		
A-549	549.50	515.32	532.41	312.43	321.82	317.13	134.92	144.99	146.99	59.25	70.46	64.86	88.35	82.13	85.24	
	± 4.02	±1.52		±4.36	± 15.22		± 5.88	± 6.27		±3.11	±3.91		±4.63	± 3.74		

Values are mean \pm SEM, n=6

TABLE 2: SHORT TERM IN VITRO CYTOTOXICITY OF SJP, SJM, SJW, SJM-A, AND SJM-NA ON DLA AND EAC CELLS

	DLA		EAC					
Extract	Conc. used	CTC ₅₀	Extract	Conc. used	CTC ₅₀			
& Fraction	(µg/mL)		& Fraction	(µg/mL)				
	1000			1000				
SJP	500	>500	SJP	500	>500			
	250			250				
	500			500				
SJM	250	>500	SJM	250	>500			
	125			125				
	500			500				
SJW	250	>500	SJW	250	>500			
	125			125				
	500			500				
SJM-A	250	175.15 ± 7.83	SJM-A	250	265.15 ± 5.09			
	125			125				
SJM-NA	500		SJM-NA	500				
	250	>500		250	>500			
	125			125				

Values are mean \pm SEM, n=6

TABLE 3: CYTOTOXICITY OF SJM-A AND SJM-NA BY LONG TERM SURVIVAL STUDY ON NORMAL VERO CELL CULTURES AND CANCER HEP-2 CELL CULTURES

	Normal Ver	o cell cultures		Cancer HEp-2 cells cultures					
Isolated	Conc.	Avg. No. of	CTC ₅₀	Isolated	Conc.	Avg. No. of	CTC ₅₀		
Fraction	Used	Colonies ±		Fraction	Used	Colonies ±			
	(µg/mL)	S.E.M. *			(µg/mL)	S.E.M. [*]			
	100	14 ± 0.11			50	16 ± 0.11			
SJM-A	50	27 ± 1.42	116.62	SJM-A	25	34 ± 1.42	71.28		
	25	55 ± 2.29			12.5	55 ± 2.29			
	200	$15\ \pm 0.04$			100	17 ± 0.04			
SJM-NA	100	21 ± 1.83	187.48	SJM-NA	50	19 ± 1.83	96.91		
	50	43 ± 2.37			25	37 ± 2.37			
Cell control		62 ± 3.25		Cell	control	63 ± 2.60			

*Values are mean ± SEM, n=6

International Journal of Pharmaceutical Sciences and Research

The effects of SJM-A on the average life span of DLA bearing mice are shown in **Fig. 1**.



FIG. 1: EFFECT OF SJM-A ON AVERAGE LIFE SPAN OF DLA TUMOR BEARING MICE. Values are mean ±SEM, * p<0.05 when compared to DLA control, n=6

The data reveal that treatment with SJM-A at 100 mg/kg significantly ($p \le 0.05$) increases the average life span (ALS) of DLA bearing mice from 17.59 \pm 0.82 to 23.86 \pm 0.99 days. The standard 5-FU at 20 mg/ kg significantly ($p \le 0.001$) increases the life span to 31.36 \pm 1.59 days. SJM-A at low doses does not show any significant increase in ALS. The effect of SJM-A on the % increase in body weight of DLA bearing mice is shown in **Fig. 2**.



FIG. 2: EFFECT OF SJM-A ON % INCREASE BODY WEIGHT OF DLA TUMOR BEARING MICE. Values are mean \pm SEM, * p<0.05 when compared to DLA control, n=6

The data reveal that treatment with SJM-A has a significant reduction in the percentage increase in body weight (p<0.001) when compared to DLA control. The standard 5-FU, however, is more potent in inhibiting the increase in body weight of tumor-bearing mice. The data reveal that the treatment with SJM-A at 100 mg/kg significantly (p≤0.01) increases the average life span (ALS) of EAC bearing mice when compared to EAC tumor control group. This is comparable to standard 5-FU at 20 mg/kg. SJM-A at 25 mg/kg also shows a moderate increase (p≤0.01 to p≤0.05) in ALS, namely, 20.55 ± 1.75.

The effect of SJM-A on the average life span (ALS) of EAC tumor-bearing mice is shown in **Fig. 3**.



FIG. 3: EFFECT OF SJM-A ON AVERAGE LIFE SPAN OF EAC TUMOR BEARING MICE. Values are mean \pm SEM, *p<0.05 when compared to DLA control, n=6

The effect of SJM-A on the percentage increase in body weight of EAC tumor-bearing mice is given in **Fig. 4.**



FIG. 4: EFFECT OF SJM-A ON % INCREASE BODY WEIGHT OF EAC TUMOR BEARING MICE. Values are mean \pm SEM, * p<0.05 when compared to DLA control, n=6

The data reveal that treatment with SJM-A (at both dose 25, 100 mg/kg) has a significant reduction in percent increase in body weight ($p \le 0.001$) when compared to EAC control. The standard 5-FU, however, is found to be more potent in inhibiting the percentage increase in body weight of tumor-bearing mice.

DISCUSSION: Alkaloids represent a very extensive group of secondary metabolites, with diverse structures, distributed in nature with important biological activities. The cyclic compound containing nitrogen in a negative oxidation state is distributed to a limited extent in living organisms ⁹. However, strong physiological effects of some alkaloids present opportunities for

utilizing them in human medicine. Although a number of the polyhydroxylated alkaloids have been reported to show anticancer activity, recent research has concentrated on developing alkaloids as drug candidates for the management of human malignancies. There is considerable evidence that alkaloids enhance the natural antitumor defenses of the body ^{10, 11}. Merschjohann has analyzed the cytotoxic potential of 34 alkaloids of various structural types against T. brucei and T. congolense ¹². Many alkaloids, of which more than 21,000 structures have been found in plants, show a high degree of toxicity towards animals. Though many alkaloids are neurotoxins that interfere with neuroreceptors, ion channels or other parts of the neuronal signal chain, a substantial number of alkaloids are cytotoxic.

Cytotoxicity is the result of the molecular interactions of an alkaloid with one or several important targets present in a cell. The main targets include DNA, RNA, and the associated enzymes and related processes (*i.e.*, replication, repair, transcription, DNA polymerase, RNA polymerase, transcriptase, repair reverse enzymes. topoisomerase, telomerase), protein biosynthesis, protein conformation, biomembrane integrity, and membrane proteins ^{13, 14, 15, 16, 9}. Cytotoxicity is one of the chemotherapeutic targets of antitumor activity ¹⁷. Most of the clinically used antitumor agents possess significant cytotoxic activity in cell culture systems ^{18.} Although the mechanism of tumor-inhibiting the activity of alkaloids is not clear, they are reported to decrease the high glycolytic activity of Ehrlich's ascites cells by inhibition of (Na-K)- ATPase in the plasma membrane¹⁹. This might be responsible for the inhibition of tumor growth.

The plants of the Solanaceae family are known to possess glycoalkaloids. The alkaloidal fraction of the methanolic extract of the plant, *Solanum jasminoides* (SJM-A) was, therefore, tested for its anticancer activity. The reliable criteria for judging the value of the anticancer potential of a sample is the prolongation of the life span of animals. In the present study, treatment with of the alkaloidal fraction SJM-A increases the life span of animals significantly with significant prevention of body weight changes when compared to tumor-bearing mice indicating its potent anticancer property. Ascitic tumor implantation is known to induce per se local inflammatory reactions with an increase in vascular permeability, resulting in an intense edema formation, cellular migration, and a progressive ascetic fluid formation ²⁰. The ascitic fluid is essential for tumor growth since it constitutes the direct nutritional source for tumor cells ²¹. The prevention of the body weight changes in the treated animals may be due to the reduction in ascitic fluid volume, which in turn may have restricted the tumor progression.

CONCLUSION: The results indicate the strong anticancer property of the alkaloidal fraction of the plant, *Solanum jasminoides*. Alkaloids are known to be widely distributed in the plant kingdom, and many of them have shown good anticancer activity ²². It is not surprising; therefore, the alkaloidal fraction of the plant, *Solanum jasminoides* shows anticancer potential. Further studies are in progress on the isolation of the alkaloids, their structure elucidation, and biological evaluation.

ACKNOWLEDGEMENT: The authors thank the Department of Science and Technology, Government of India for financial support.

COMPETING INTEREST: The authors declare that they have no competing interest.

REFERENCES:

- 1. Newman DJ, Cragg GM and Sander KM: Natural products as sources of new drugs over the period 1981-2002. Journal of Natural Product 2003; 66: 1022-37.
- Sriram D, Yogeeswari P, Thirumurugan R and Bal TR: Camptothecin and its analogs: A Review on their chemotherapeutic potential. Natural Product Research 2005; 19: 393-12.
- 3. Jain SC, Sharma GL: Screening of some Solanaceous plants for glycoalkaloids. Planta Medica 1977; 31: 212-13.
- 4. Harborne JB: Phytochemical Methods. Chapman and Hall, New York 1984; 123-75.
- 5. Francis D and Rita L: Rapid colorimetric assay for cell growth and survival: modifications to the tetrazolium dye procedure giving improved sensitivity and reliability. Journal of Immunology Methods 1986; 89: 271-77.
- 6. Philip S, Rista S, Dominic S, Anne M and James M: New colorimetric cytotoxicity assay for anticancer- drug screening. Journal of Natural cancer Institute 1990; 82: 1107-12.
- Unnikrishnan MC and Kuttan R: Cytotoxicity of an extract of species to cultured cells. Nutrition and Cancer 1988; 11: 251-57.
- Freshney IR: Culture of Animal cells A manual of Basic Technique. 4th edition, New York, USA, John Wiley and sons. Inc. Publication 2000.

- Roberts MF and Wink M: Biochemistry, ecology, and medicinal applications. In: Roberts, M.F., Wink, M. (Eds.), Alkaloids. Plenum Press, New York, London 1998: 1-7.
- Kino T, Inamura N, Nakahara K, Kiyoto S, Goto T, Terano H, Kohsaka M, Aoki H and Imanaka H: Studies of an immunomodulator swainsonine II. Effect of swainsonine on mouse immunodeficient system and experimental murine tumor. The Journal of Anti-biotics 1985; 38: 936-40.
- 11. Humphries MJ, Matsumoto K, White SL, Molyneux RJ and Olden K: Augmentation of murine natural killer cell activity by swainsonine, a new antimetastatic immunomodulator. Cancer Research 1988; 48: 1410-15.
- 12. Merschjohann K, Sporer F, Steverding D and Wink M: *Invitro* effects of alkaloids on bloodstream forms of *Trypanosoma brucei* and *T. congolense. Planta Meica* 2001; 67: 623-27.
- Wink M: Allelochemical properties and the raison d'être of alkaloids. In The Alkaloids, G. Cordell; Academic Press: Orlando FL 1993; 43: 1-118.
- 14. Wink M: Interference of alkaloids with neuroreceptors and ion channels. In Bioactive natural products; Elsevier: Amsterdam, New York 2000; 11: 3-129.
- 15. Wink M and Schimmer O: Modes of action of defensive secondary metabolites. In Function of Plant secondary metabolites and their exploitation in biotechnology; Wink, M., Ed.; Annual Plant Reviews; Sheffield Academic Press: Sheffield 1999; 3: 17-133.

- Wink M: Molecular modes of action of cytotoxic alkaloids

 From DNA intercalation, spindle poisoning, topoisomerase inhibition to apoptosis and multiple drug resistance. In The Alkaloids; Cordell, G., Ed.; Elsevier: Amsterdam, New York; 2007; 64: 1-48.
- 17. Suffness M and Pezzuto JM: Methods in Plant Biochemistry, Vol. V1. Academic Press, New York 1991 71.
- Carlo GD, Mascolo N, Izzo AA and Capaso F: Flavonoids and Alkaloids: old and new aspects of a class of natural therapeutic drugs. Life Science 1999; 65: 337-53.
- Pomilio AB, Sola GAR, Mayer AMS and Rumi LS: The antitumor and cytotoxic screen of 5,6,7-trisubstituted flavones from *Gomphrena martiana*. Journal of Ethnopharmacology 1994; 44: 25-33.
- Badami S, Moorkoth S, Rai SRMS, Kannan E and Bhojraj S: Antioxidant activity of Caesalpinia sappan heartwood. Biological and Pharmaceutical Bulletin 2003; 26: 1534– 37.
- 21. Gupta M, Mazumder UK, Kumar RS, Sivakumar, T and Vamsi MLM: Antitumour activity and antioxidant status of *Caesalpinia bonducella* against Ehrlich Ascites Carcinoma in Swiss albino mice. Journal of Pharmaceutical Science 2004; 94: 177-84.
- 22. Patel JR, Choulisa B and Dholakiya BZ: Plant-derived compounds having activity against P388 and L1210 Leukemic cells. Journal of Chemical Science 2011; 1: 1-16.

How to cite this article:

Porwal O, Nanjan MJ, ChandrasekarMJN, Srinivasan R and Gupta S: Anticancer potential of *Solanum jasminoides*. Int J Pharm Sci & Res 2014; 5(9): 3768-74. doi: 10.13040/JJPSR.0975-8232.5(9).3768-74.

All © 2013 are reserved by International Journal of Pharmaceutical Sciences and Research. This Journal licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported License.

This article can be downloaded to **ANDROID OS** based mobile. Scan QR Code using Code/Bar Scanner from your mobile. (Scanners are available on Google Playstore)