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EVALUATION OF NEPHROPROTECTIVE ACTIVITY OF ETHANOLIC EXTRACT OF *BAUHINIA PURPUREA* IN GENTAMICIN INDUCED NEPHROTOXICITY IN RATS

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ABSTRACT: Objective: To study the nephroprotective activity of *Bauhinia purpurea* unripe pods and bark against gentamicin-induced nephrotoxicity. **Materials and Methods:** Twenty four healthy adult albino rats of either sex (150-200 g) were randomly divided into four groups of six animals each. Group, I served as vehicle control and Group II as the negative control (gentamicin 80 mg/kg/d, i.p). Groups III and IV (test groups) were administered ethanolic extract of *Bauhinia purpurea* bark (BBE) and unripe pods (BPE) in the dose of 400 mg/kg/day, one hour before gentamicin administration. All the treatments were given for 8 days. On the 9th day, blood and urine samples were used for biochemical analysis and kidneys for histology and assessing anti-oxidant activity. **Results:** Gentamicin caused nephrotoxicity as evidenced by highly significant ($p < 0.001$) elevation of blood urea, serum creatinine and urine glucose in Group II as compared to Group I. Also administration of Gentamicin in group II caused oxidative stress in rats suggested by significant increase in MDA level and significant decrease in Catalase and Reduced GSH level. While administration of BBE and BPE in group III and IV respectively caused a significant decrease ($p < 0.01$) of blood urea, serum creatinine, and urine glucose as compared to group II. In case of antioxidant status there is significant increase ($p < 0.01$) in Catalase and Reduced GSH level while there is significant decrease ($p < 0.01$) in MDA level was observed in group III and IV. **Conclusion:** Ethanolic extract of *Bauhinia purpurea* unripe pods and bark has nephroprotective activity against gentamicin induced nephrotoxicity in rats.

INTRODUCTION: Gentamicin, a commonly used wide spectrum antibiotic is highly effective against severe gram-negative bacterial infections ¹. Unfortunately, its high efficacy is associated with the side effect of nephrotoxicity.

This nephrotoxicity in the form of acute renal failure occurs in 10-30% of patients receiving gentamicin ²⁻³. Although it is generally reversible upon drug discontinuation, but it complicates the patient's condition, prolongs the hospital stay and increases the medical expenditure ⁴. Proximal tubular necrosis (PTN) underlies the pathogenesis of gentamicin nephrotoxicity ⁵. But the mechanism of PTN is not understood clearly. However, several studies suggested that reactive oxygen species (ROS) may be an important contributor in the pathogenesis of gentamicin nephrotoxicity ⁶.

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Abnormal ROS production leads to proximal tubular necrosis *via* lipid peroxidation of membrane lipids, protein denaturation, and DNA damage⁷⁻⁸. From the last few years, tremendous interest has been shown on the role of herbal substances as an antioxidant for the management of various diseases.

Bauhinia purpurea, medium-sized deciduous tree belongs to family Caesalpiniaceae, found throughout India predominantly in sub-Himalayan forests⁹. It is commonly known as an orchid tree, lalkachnar, and kaniar. It has been used to treat various human ailments in folklore medicine such as pain, dropsy, rheumatism, convulsion, wound healing, delirium, and septicaemia¹⁰. Based on its folklore uses various pharmacological activities have been reported like analgesic and anti-inflammatory¹¹, antimalarial, antitubercular, anti-fungal and cytotoxic¹², cardiotoxic¹³, hypolipidaemic¹⁴, antioxidant¹⁵, hepatoprotective¹⁶ and antidiabetic¹⁷. There is single study, where nephroprotective activity¹⁸ of ethanolic extracts of unripe pods at the dose of 300 mg/kg against gentamicin-induced nephrotoxicity has been studied. In this study, we have evaluated the nephroprotective and antioxidant activities of unripe pods as well as the bark of *B. purpurea* against gentamicin-induced nephrotoxicity.

MATERIAL AND METHODS:

Plant Materials: The unripe pods and bark of *Bauhinia purpurea* were collected from the garden of Aligarh Muslim University, Aligarh during the month of April-May. Collected unripe pods and bark were identified and authenticated by Dr. Athar Ahmed, Assistant professor, Department of Botany, AMU., Aligarh and a voucher specimen with voucher no.DWS/VS/01 was submitted for future reference. Freshly collected unripe pods and bark in bulk were shade dried and pulverized to a coarse powder by the Soxhlet mechanical grinder. The powder was then subjected to extraction with absolute ethanol. The extract thus obtained was evaporated to dryness. Thus the yield obtained was 4.88% (w/v).

Experimental Animals: Adult albino rats of either sex were procured from central animal house, J.N Medical College, AMU, Aligarh (Registration no. 401/CPCSEA). They were housed in polypropylene

cages and maintained according to CPCSEA guidelines, *i.e.*, at room temperature of $27^{\circ} \pm 2^{\circ}\text{C}$ under 12 h light and dark cycle. Rats were acclimatized for one week before experimentation. Ethical clearance for the study was obtained from Institutional Animal Ethics Committee of Jawaharlal Nehru medical college, A.M.U, Aligarh.

Experimental Design: Twenty-four adult healthy albino rats of either sex were divided into four groups, each group containing six rats.

Group I served as the vehicle control group received normal saline intraperitoneally (1ml/kg) and distilled water per orum (1ml/kg) daily.

Group II served as negative control group received gentamicin at the dose of 80 mg/kg intraperitoneally¹⁹ and distilled water per orum daily (1ml/kg).

Group III (BBE) and Group IV (BPE) were administered *Bauhinia purpurea* unripe pods and bark extract in a dose of 400 mg/kg per orum respectively daily along with the same dose of gentamicin as in group II. The extract was given one hour before administration of gentamicin in the treatment group. All the groups were given treatment over 8 consecutive days. Following the last dose of treatment, animals were housed individually in separate metabolic cages to collect 24-hour urine. Twenty four hours after the last dosing, *i.e.*, on 9th-day rats were anesthetized with ketamine and sacrificed. Blood samples were collected by cardiac puncture method. Kidneys were dissected out and weighed. One kidney was preserved in 10% formalin for histological examination, and the other kidney was homogenized for biochemical analysis for oxidative stress and antioxidant activity.

Biochemical Analysis and Tissue Studies: Serum urea and creatinine levels, as well as urine glucose levels, were determined using diagnostic kits from span diagnostics, Hyderabad. For estimation of oxidative stress, the kidney tissue was cut into small pieces and homogenized in 10% phosphate buffer, by using homogenizer. Tissue lipid peroxidation level (MDA) was determined by the method described by Buege and Aust²⁰. According to this method, MDA condenses with two equivalents of thiobarbituric acid to give a

fluorescent red derivative which was assayed spectrophotometrically at 532nm. The results were expressed as nmol/g wet tissue weight. Catalase, an antioxidant enzyme level was estimated by the method described by Sinha²¹ and was expressed as Units of H₂O₂ consumed/min/g wet tissue. Reduced glutathione level estimation was performed according to the protocol of Ellman²².

Histological Examinations: Formalin preserved kidney tissue was embedded in paraffin, and 5-6 µm sections were cut using a rotary microtome and stained with hematoxylin and eosin (H&E)²³. All sections were examined with a light microscope for tubular degeneration, tubular necrosis, mononuclear cell infiltration, and hyaline casts.

Statistical Analysis: Results were expressed as the mean ± SD. A significant statistical difference was determined by one-way analysis of variance

(ANOVA) followed by post-hoc dunnett's test for multiple comparisons. Probability values (*P*) less than 0.05 were considered to be statistically significant.

RESULTS AND DISCUSSION: The nephrotoxicity of aminoglycoside antibiotics, especially that of the most commonly used antibiotic, gentamicin (GM) is well documented²⁴⁻²⁵. Several studies have reported that oxygen-free radicals are considered to be important mediators of GM-induced acute renal failure²⁶⁻²⁷. Therefore agents with antioxidant property can be used for amelioration of GM-induced nephrotoxicity. Results summarized in **Table 1** show the effect of GM alone and in simultaneous treatment with *Bauhinia purpurea* bark (BBE400) and unripe pods (BPE400) extract in the dose of 400 mg/kg on the physical parameters.

TABLE 1: EFFECT OF ETHANOLIC EXTRACTS OF UNRIPE PODS AND BARK OF *B. PURPUREA* ON PHYSICAL PARAMETERS IN GENTAMICIN INDUCED NEPHROTOXICITY

S. no.	Groups	% Change in body weight	Kidney weight per 100g B. wt (g)	Urine volume/day (ml)
I	Vehicle Control	(+) 3.76±0.54	0.40±0.017	9.83±0.77
II	Negative Control	(-) 9.51±0.47***	0.60±0.007***	18.75±1.13***
III	BBE 400	(-) 7.32±1.02	0.55±0.014*	15.00±0.28
IV	BPE 400	(-) 6.54±0.89	0.55±0.016*	16.00±0.76

BBE: *Bauhinia purpurea* bark extract, BPE: *Bauhinia purpurea* unripe pods extract; 400 denotes doses in mg/kg; Data were expressed in mean ± SEM (n=6 rats/group). The negative control group was compared with the Normal control group, and all other groups were compared to the Negative control group, ***p < 0.001, **p < 0.01, *p < 0.05 was considered significant.

GM treatment resulted in a significant decrease in % change in BW while there is a significant increase in kidney weight, and urine volume was observed compared to the vehicle control group. Decrease body weight resulted from the increased catabolism seen in acute renal failure accompanied by a decrease in food intake²⁸. Administration of ethanolic extract of *Bauhinia purpurea* bark (BBE) and unripe pods (BPE) in a dose of 400 mg/kg for 8 days caused mild protection from the deleterious effect of GM on above physical parameters. There was less reduction in body weight in group III and group IV as compared to the negative control group. But the improvement was less than in the normal control group, suggesting that supplementary energy must be added to the diet. The significant increase in normalized kidney weight of gentamicin treated negative control group (Group II) probably resulted from oedema due to drug-induced tubular necrosis. Also, the significant

increase in urine volume per day in gentamicin treated negative control group (Group II) compared to the normal control group (Group I) confirms the gentamicin-induced nonoliguric acute renal failure. However, the increase in kidney weight and urine volume in *B. purpurea* treated groups (Group III and Group IV) was significantly less as compared to the negative control group (Group II). But the *B. purpurea* treatment failed to completely prevent the edema caused by gentamicin administration. Probably, it might be due to the shorter duration of the study.

Nephrotoxicity, in the form of acute renal failure, was induced in the negative control group (Group II) according to the method described by Singh P *et al.*, 2009¹⁹. Gentamicin administration in the dose of 80 mg/kg/d for 8 days in the negative control group (Group II) significantly elevated blood urea, serum creatinine and urine glucose (p < 0.001) as

shown in **Table 2**. Eight days treatment of Ethanolic extract of *B. purpurea* bark and unripe pods in the dose of 400 mg/kg along with gentamicin, produced moderate protection compared to the negative control group. While *B. purpurea* bark extract (BBE400) reduced blood urea, serum creatinine, urine glucose by 22.42%, 27.80% and 22.10 % to a level of 156.8 ± 2.83 , 2.57 ± 0.052 , 74.77 ± 2.13 mg/dl respectively. *B. purpurea* unripe pods extract (BPE400) reduced the

blood urea, serum creatinine, and urine glucose by 23.29%, 28.25% and 21.22% to a level of 155.63 ± 2.47 , 2.56 ± 0.055 and 75.44 ± 2.38 respectively. Our experimental results indicate that BBE and BPE at the dose of 400 mg/kg have exerted protection against GM nephrotoxicity. The mechanism of this protective effect is not certain. However, BBE and BPE may have antagonized the oxidative stress of GM as explained subsequently.

TABLE 2: EFFECT OF ETHANOLIC EXTRACTS OF UNRIPE PODS AND BARK OF *B. PURPUREA* ON BIOCHEMICAL PARAMETERS OF GENTAMICIN INDUCED NEPHROTOXICITY

S. no.	Groups	Blood urea (mg/dl)	Serum creatinine (mg/dl)	Urine glucose (mg/dl)
I	Vehicle Control	55.8±3.92	0.96±0.076	15.72±1.17
II	Negative Control	186.00±14.45***	3.19±0.267***	91.53±5.19***
III	BBE 400	156.8±2.83**	2.57±0.052**	74.77±2.13**
IV	BPE 400	155.63±2.47**	2.56±0.055**	75.44±2.38**

BBE: *Bauhinia purpurea* bark extract, BPE: *Bauhinia purpurea* unripe pods extract; 400 denotes doses in mg/kg; Data were expressed in mean \pm SEM (n=6 rats/group). The negative control group was compared with the Normal control group, and all other groups were compared to Negative control group, ***p < 0.001, **p < 0.01, *p < 0.05 was considered significant.

Significant increase in lipid peroxidation indicated by elevated MDA level (p < 0.001) and reduction in GSH and CAT activity (p < 0.01) after the treatment of gentamicin in negative control group indicated the production of free radicals and involvement of oxidative stress to nephrotoxicity caused by gentamicin treatment **Table 3**. The same results have been reported earlier by Walker *et al.*, 1999²⁶. Administration of Ethanolic extract of *B. purpurea* bark and unripe pods produced protection against lipid peroxidation (p < 0.01) and increased the

activity of antioxidant reduced GSH and Catalase (p < 0.01) against gentamicin-induced oxidative stress **Table 3**. *B. purpurea* contains a major class of secondary metabolites like glycosides, flavonoids, saponins, triterpenoids, phenolic compounds, oxepins, fatty acids, and phytosterols^{12- 29- 30}. Therefore the nephroprotection observed with the treatment of BBE and BPE 400 mg/kg might be due to the presence of these polyphenolic compounds.

TABLE 3: EFFECT OF ETHANOLIC EXTRACTS OF PODS AND BARK OF *BAUHINIA PURPUREA* ON GENTAMICIN INDUCED OXIDATIVE STRESS

S. no.	Groups	MDA (nmoles/g wet tissue wt.)	Catalase (nmoles H ₂ O ₂ consumed/min/mg protein)	GSH (nmoles/mg protein)
I	Normal Control	58.69 \pm 1.94	124.81 \pm 2.20	20.62 \pm 0.25
II	Vehicle Control	117.19 \pm 1.83***	75.09 \pm 2.06***	10.16 \pm 0.22***
III	BBE 400	108.18 \pm 0.79**	84.79 \pm 1.31**	10.97 \pm 0.11**
IV	BPE 400	107.80 \pm 1.15**	85.04 \pm 1.67**	11.00 \pm 0.16**

BBE: *Bauhinia purpurea* bark extract, BPE: *Bauhinia purpurea* unripe pods extract; 400 denotes doses in mg/kg; Data were expressed in mean \pm SEM (n=6 rats/group). The negative control group was compared with the Normal control group, and all other groups were compared to Negative control group, ***p < 0.001, **p < 0.01, *p < 0.05 was considered significant

These findings correlated with the histological examination **Table 4**.

TABLE 4: GRADING OF HISTOLOGICAL FEATURES OF RAT KIDNEY TISSUE SECTIONS IN DIFFERENT GROUPS IN GENTAMICIN INDUCED NEPHROTOXICITY

Histopathological features	Vehicle control	Negative control	BBE400	BPE400
Glomerular congestion	-	+++	++	++
Mononuclear cells infiltration	-	+++	++	+
Tubular necrosis	-	+++	++	++
Tubular hyaline casts	-	++++	+	-

Light microscopic examination of stained kidney tissue slices from the vehicle control group (Group I) showed normal histological features with no structural alterations in glomeruli and tubules **Fig. 3A**, but the GM-treated group (Group II) revealed more extensive and marked tubular necrosis and numerous hyaline casts **Fig. 3B**. There were mononuclear infiltrations which are considered to be the response of the body tissue facing any injurious agent. These alterations could be due to the accumulation of free radicals resulting from an increased lipid peroxidation in the renal tissues of

the GM-treated group. Renal lesions were also characterized by vascular congestion as well as tubular obstruction. Similar changes were also reported by Yaman *et al.*,³¹ and Abdel-Raheem *et al.*³² who demonstrated structural changes in renal tissue of GM-treated animals and its reversal by various agents. Glomerular and tubular epithelial changes were considerably mild in the group treated with both GM along with *B. purpurea* unripe pods and bark extract **Fig. 3C** and **D**, thus showing the preventive effect of BPE400 and BBE400 against GM-induced tissue damage.

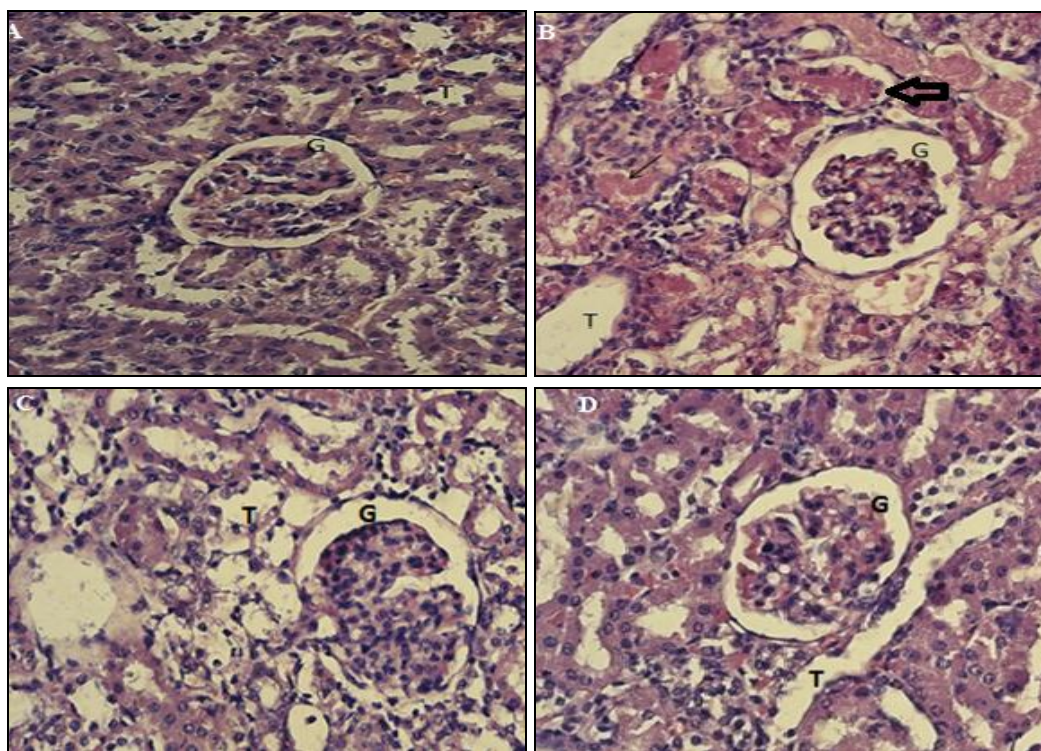


FIG. 1: PHOTOMICROGRAPH OF RAT KIDNEY TISSUE SECTION OF DIFFERENT GROUPS

(A) Normal histology of kidney tissue in the vehicle control group (Group I) showing normal glomerulotubular architecture (H&E $\times 400$). (B) Kidney tissue section of GM only treated negative control group (Group II) showing massive mononuclear cell infiltration, glomerular congestion, tubular necrosis, and numerous hyaline casts (shown in arrow) (H&E $\times 400$). (C) Kidney tissue section of rats treated with GM plus BBE (Group III) showing mild mononuclear cell infiltration with mild glomerular congestion (H&E $\times 200$). (D) Kidney tissue section of rat treated with GM plus BPE treated group (Group IV) showing almost complete prevention of histopathological alterations (H&E $\times 200$).

CONCLUSION: The results of our study showed that cotreatment with *Bauhinia purpurea* unripe pods and bark afforded significant protection against nephrotoxicity induced by gentamicin treatment. The beneficial effect of *Bauhinia purpurea* as suggested by biochemical findings and supported by histological evidence in gentamicin toxicity might be due to the scavenging effect of the extract. These findings indicate that *Bauhinia purpurea* ethanolic extract supplementation may reduce gentamicin-induced nephrotoxicity.

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CONFLICT OF INTEREST: Nil

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