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# HEPATOPROTECTIVE EVALUATION OF *BIXA ORELLANA* L. SEED EXTRACT AGAINST ACUTE ETHANOL-INDUCED HEPATOTOXICITY IN RATS

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### **Keywords:**

Antioxidant,
Bixa orellana, Ethanol,
Hepatotoxicity, Hepatocellular
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**ABSTRACT:** Disorders caused by excessive use of alcohol known as alcohol use disorders (AUD) are the most common cause of liver diseases including cirrhosis. In adults, the most common cause of death is alcohol consumption which leads to an array of alcoholic liver diseases (ALD). Present study has been conducted to investigate the hepatoprotective activity of Bixa orellana L. (BO) seed extract against acute ethanol-induced hepatotoxicity in rats. The characterization of BO seed extract was performed using standard biochemical analysis. 50% ethanolic extract of Bixa orellana (BOE 200 and 400 mg/kg body weight) was administered daily for 8 days in experimental animals for the assessment of hepatoprotective activity. To develop hepatotoxicity, animals were orally administered with alcohol (40%) 12 ml/kg at 2 h after the doses of BOE every day for eight consecutive days except the rats of normal group. The hepatoprotective activity was assessed using various biochemical parameters like aspartate aminotransferase, alanine aminotransferase, alkaline phosphatise, bilirubin, albumin, cholesterol and lactate dehydrogenase. However, the results demonstrated that the treatment with Bixa orellana seed extract significantly (P<0.05 - P<0.001) and dose-dependently prevented alcohol-induced increase in serum levels of hepatic enzymes such as AST, ALT, ALP, BIL, ALB, CHL and LDH. Histopathological study further attenuated the hepatocellular necrosis and led to reduction in inflammatory cells infiltration. The present study concludes and suggests that Bixa orellana seed extract significantly protected the liver from alcohol induced toxicity. It also showed that the extract contains numerous antioxidant compounds with hepatoprotective effect.

**INTRODUCTION:** Alcohol has been the most frequently abused drug for centuries, but it was not until the 1960s that it was recognized as a direct hepatotoxin <sup>1</sup>. In today's scenario, the third most common cause of mortality and morbidity in the Indian society remains alcohol which also contributes to one of the major socially acceptable harmful agent <sup>2</sup>.



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Long-term excess alcohol exposure leads to alcoholic liver disease (ALD) <sup>3</sup> - a global health problem without effective therapeutic approach <sup>4</sup> and is one of the leading causes of liver cancer, cirrhosis, fibrosis, acute and chronic liver failure which can therefore lead to significant morbidity and mortality <sup>5</sup>. ALD is increasingly considered as a complex and multifaceted pathological process, involving oxidative stress, inflammation and excessive fatty acid synthesis <sup>3</sup>.

However, the reports might differ in the data, but the current consensus states that about 100 - 120 g of ethanol is ingested by alcoholic hepatitis patients every day since last 10 to 20 years.

This quantity is measured to be standard with the drink of pure alcohol containing 14 g, which is considered equivalent to 1.5 ounces (44.36 mL), 5 ounces (147.87 mL) of wine, 12 ounces (354.88 mL) of beer and a "shot" of 80-proof liquor <sup>6</sup>. In the initial stage of ALD, long-term heavy alcohol exposure leads to fatty liver (hepatic steatosis), characterized by triglyceride (TG) accumulation in hepatocytes, which has been widely assumed to be a benign and reversible condition, on continued alcohol consumption hepatic steatosis progresses to advanced stages of ALD, such steatohepatitis, fibrosis. cirrhosis, and even hepatocellular carcinoma, particularly in presence of co-factors including hepatitis virus infection, smoking and diabetes <sup>7</sup>. Fatty liver, an early response to alcohol consumption, develops in most (more than 90%) heavy drinkers, with earlymild steatosis in zone 3 (perivenular) hepatocytes; it can also affect zone 2 and even zone 1 (periportal) hepatocytes when liver injury is more severe<sup>3</sup>.

As per the World Health Organization (WHO) fact sheet published in January 2015, around 3.3 million deaths all over the world are observed every year due to harmful use of alcohol which reflects an average of 5.9% of total deaths. Consumption of alcohol is the most prominent root cause for more than hundreds of diseases and injuries. According to the Disability- Adjusted Life Years (DALYs) measured value, the burden of different types of injury and diseases all over the world due to consumption of alcohol is about 5.1%. Often it is observed that about 25% of individuals having age group between 20 - 39 years approximately die due to alcohol consumption <sup>8</sup>. Worldwide about 16.0% of drinkers aged 15 years or older are engaged in heavy episodic drinking. Liver-related mortality from alcohol contributes to 4% of mortality and 5% of disability adjusted life years (DALY) globally, with highest impact in Europe, where these figures are 7% and 12%, respectively <sup>1</sup>. However, in 2016 all over the world the alcohol consumption level was 6.4 litres of pure alcohol per person with the age of 15 years or more, with suitable variations between the regions of WHO<sup>9</sup>.

Bixa orellana L., commonly known as Annatto (also called as Chinese dye tree or the lipstick tree), belongs to the family Bixaceae and is native to

tropical and subtropical regions of the world <sup>10</sup>, but now cultivated in many tropical countries of the world including India. Bixa orellana is an evergreen shrub or small tree, 2 - 8 m high bark light to dark brown, tough, smooth sometimes. Flowers in terminal branched panicles, 8 - 50 flowered, covered with reddish brown scales; petals 4 - 7, obovate,  $2-3 \times 1$  - 2 cm, pinkish, whitish. Fruit a spherical or broadly elongated ovoid capsule,  $2 - 4 \times 2 - 3.5$  cm, flattened, green, greenish-brown or red when mature; seeds numerous, with bright orange-red fleshy coats 11. Natural colorants obtained from the seeds of achiote plant (annatto) have been used since pre-Hispanic times <sup>12</sup>. Its leaves have antiseptic, antibacterial, and antiemetic effects, whereas the seeds were used for fever, buccal tumors, jaundice. Many pharmacologic studies have been conducted on its potential as a source of medicine  $^{13}$ . B. orellana was shown activity against protozoan, helminths and had platelet anti-aggregant activity <sup>14</sup>. Annatto tocotrienols (AnT3), which contain approximately 90% d-tocotrienol (d- T3) 15.

Twigs and leaves of Bixa orellana (Bixaceae) are claimed to be used traditionally as a herbal drug in north Kerala in treatment of liver disorders as an emollient and seeds have been proved to possess hepatoprotective activity as is used in many marketed herbal formulations <sup>16</sup>. Bixa orellana is a hepatoprotective against safe agent various tetrachloride hepatotoxins (carbon acetaminophen) in albino rats and act similar to that of silymarin as hepatoprotective and is more inducer of biochemical effective indicating the hepatoprotective property 17, 18. Hence, an attempt has been made in this study to investigate the hepatoprotective activity of seed extract of Bixa orellana against acute ethanolinduced liver injury in Wistar rats.

## **MATERIALS AND METHODS:**

Chemicals and Reagents: All the chemicals and reagents used were of analytical grade, and procured from Sigma chemicals Co., USA and Qualigens fine chemicals, Mumbai, India. Organic solvents such as formic acid, n-hexane, petroleum ether, chloroform, ethyl acetate, and methanol (high-pressure liquid chromatography grade, Merck) were used.

The drug used as standard was Silymarin (Sigma Chemicals, USA (BCBF6608V) Made by china). All the enzymatic kits used were obtained from Span Diagnostics Ltd. Surat (76LS200-60).

Plant Collection and Authentication: The ripped fruits of *Bixa orellana* Linn. (Bixaceae) were collected from local market of Lucknow, in the month of January 2014. The plant material was authenticated by Mr. Muhmmad Arif (Assistant Professor) and Dr. Arshan Hussain (Associate Professor & Head) Department of Pharmacognosy and Phytochemistry, Faculty of Pharmacy, Integral University, Lucknow - 226022, A voucher specimen of *Bixa orellana* Linn. (IU/PHAR/HRB/14/02) was deposited in the institute for further reference.

Extract Preparation for Animal Study: The freshly collected fruits of Bixa orellana Linn. were washed with distilled water to remove dirt and soil and shade dried in a ventilated place at room temperature. Dried seeds were cut into small pieces and reduced to coarse powder by mechanical grinder and further extraction was carried out with 50% hydroalcoholic by cold percolation method to avoid damage due to heat. The extract was filtered and concentrated under reduced pressure below 40 ± 1 °C using rotevavaccum rotary evaporator (Model no- UDOIAB-2391 Medica instrument) to dryness to get a constant weight. The % yield was found to be 20.25% w/w. The extract was stored in -20 °C freezer and used for Pharmacological investigation.

**Preparation of TLC Sample:** Coarsely powdered sample of fruit seeds 10 g was extracted with 300 ml methanol in a Soxhlet extractor until discolouration of the sample. The extract was filtered and concentrated to dryness under reduced pressure below 40 ± 1 °C using a rotary vacuum evaporator (Model no- UDOIAB-2391 Medica instrument). Percentage (%) yield was found to be 13.69 % w/w. A preparative TLC plate was cut into small pieces and activated at 110 °C for 30 min. Single spot of the sample was applied on the plate using the capillary. Further the plate was developed in a chamber having desired solvent systems for bixin and amino acids as mentioned below in Table 1. The ratios of the distances travelled by the samples compared to the distances travelled by solvents were identified on the TLC plates as colored spots. Each plate was then placed inside a closed container with iodine vapour which helped to specifically identify each and every spot more clearly for better visualization.

**Animal Experiments:** Adult female Wistar rats weighing 160 ± 20 g procured from National Laboratory Animal Center, Central Drug Research Institute (CDRI), Lucknow were used for the study. animals housed separately The were polypropylene cages for acclimitization at a temperature of 23  $\pm$  2 °C and 50 - 60% relative humidity, with a 12 hr light/dark cycle one week before and during the commencement of the experiment. Animal were kept on standard pellet diet (Dayal animal feed, Unnao, India) and drinking water ad libitum throughout the housing period.

All experimental procedures involving animals were conducted in accordance with the guidelines of Committee for the Purpose of Control and Supervision on Experiments on Animals (CPCSEA). The study protocols were approved by the Institutional Animal Ethics Committee (IAEC) of Integral University, Faculty of Pharmacy, Lucknow, India (Reg. no. 1213/GO/ac/08/CPCSEA).

**Experimental Protocol:** Totally 25 experimental animals (female Wistar rats) randomly divided into five groups consisting of five rats (n=5) per group were used in this study as follows:

**Group I:** Control received 0.3% CMC (vehicle) orally for 8 days throughout the study.

**Group II:** Female Wistar rats received 12 ml/kg b.wt of 40% ethanol which served as acute ethanolintoxicated control.

**Group III:** Rats were treated with *Bixa orellana* seed extract (BOE) dose 200 mg/kg b.wt before the dose of ethanol orally for 8 consecutive days

**Group IV:** Rats received *Bixa orellana* fruit seed extract (400 mg/kg b.wt) as in group III 2 h before the ethanol dose (12 ml/kg b.wt)

**Group V:** Rats received standard drug Silymarin (100 mg/kg b.wt) as in group III followed by the dose of ethanol for 8 continuous days <sup>16</sup>.

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**Serum Collection and Organ Isolation:** At the end of the treatment *i.e.*, six hours after the administration of last single dose of 40% ethanol, rats were anaesthetized and blood was collected by retro-orbital plexus followed by heart puncture as per the regulations of Good Laboratory Practices. After withdrawal of blood the animals were sacrificed. The serum was separated by centrifugation at 3000 rpm at 4 °C for 20 min and

stored at -80 °C for the analysis of various biochemical parameters. The isolated liver tissue was washed twice with ice cold saline, blotted, dried and then weighed. The relative liver weight was calculated as the percentage ratio of liver weight to the body weight. A small portion of the tissue was fixed in formalin for histopathalogical examination.

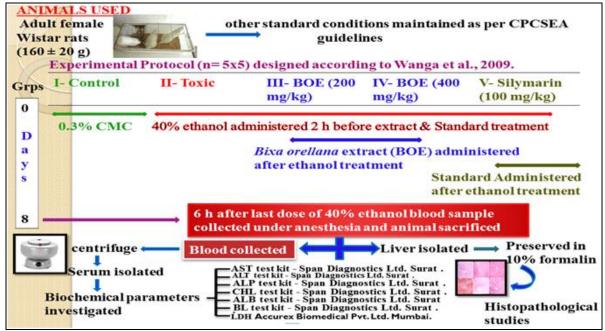


FIG. 1

Assessments of Liver Function Test (L.F.T): Serum enzymes like, Serum alanine transaminase (ALT), Alkaline Phosphate (ALP), Aspartate Transaminase (AST), Bilurubin, Total Cholesterol and Albumin (ALB) were determined by using standard kits from Span diagonistic ltd, Surat, India. Serum lactate dehydrogenase (LDH) was estimated by using standard kits from Accurex biomedical Pvt., Ltd., Mumbai. India. All estimations were carried out using UV spectrophotometer (Shimadzu, India) per standard kit methods.

Histopathological Assessment: For histologic studies, the liver tissues were fixed with 10% phosphate buffered neutral formalin, dehydrated in graded (50 - 100%) alcohol and embedded in paraffin. Thin sections (5 M) were cut stained with routine hematoxylin and eosin stain for photo microscopic assessment. The examination was qualitative, with the purpose of determining histopathological lesions in liver tissue.

**Statistical Analyses:** The data were represented as mean  $\pm$  standard error of mean (S.E.M.) for five rats. Student t-test was followed by individual comparison by Newman- Keuls test using GraphPrism Pad software (Version 6.05, GraphPad Software, Inc. USA) for the determination of level of significance. The value of probability less than 5% (P<0.05) was considered statically significant.

#### **RESULTS:**

Standardization of Plant Extract by Thin Layer Chromatography: A number of spots on the TLC plates of the plant extract identified in Fig. 2 clearly indicate the presence of various chemical compounds. Retention factors  $(R_f)$  was calculated. Different ratios of n-hexane and ethyl acetate solvent system for bixin and Chloroform, ethyl acetate and formic acid for amino acids (Glycine, Leucine and Proline) gave many  $R_f$  values for the plant extract of  $Bixa\ orellana$  as mentioned in Table 1.

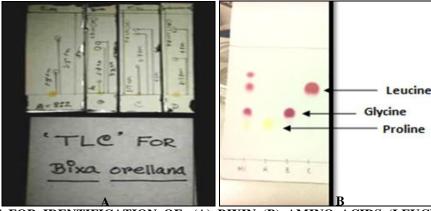


FIG. 2: TLC PLATE FOR IDENTIFICATION OF: (A) BIXIN (B) AMINO ACIDS (LEUCINE, GLYCINE AND PROLINE) FROM BIXA ORELLANA EXTRACT

TABLE 1: VARIOUS  $R_f$  VALUES CALCULATED FOR DIFFERENT SOLVENTS RATIOS FOR B. ORELLANA EXTRACT

Extract of Bixa orellana for identification	Solvent system	Ratio	$R_f$ values
Bixin	n-hexane: ethyl acetate	8:2, 7:3, 6:4, 5:5	0.3, 0.7; 0.2, 0.5, 0.6;
			0.3, 0.7, 0.8; 0.4, 0.7
Amino acid (Leucine, Glycine, Proline)	chloroform: ethyl acetate:	5:4:1	0.6; 0.3; 0.2
	formic acid		

Animal Experiment: Administration of ethanolic BOE extract in rats without introducing ethanol was performed to determine the BOE effect alone if it can cause hepatic damage. The effect of 50% *Bixa orellana* ethanolic extract on liver weight, body weight and serum marker enzymes was found to attenuate the toxic effect of 40% ethanol on rats, thereby contributing to its antihepatotoxic potential.

Effect of 50% Ethanolic Extract of *Bixa orellana* on Body Weight, Liver Weight and Relative Liver Weight in Control and 40% Ethanol - Induced Hepatotoxicity in Rats: The final body weight of normal group I rats showed  $201.2 \pm 11.3$  g which was significantly decreased to  $165.7 \pm 10.4$  g (P<0.01) compared to group II rats following 40% ethanol treatment. In BOE-treated Group III (200 mg/kg) and IV (400 mg/kg) rats, the final

body weights significantly increased to 191.5  $\pm$  12.4 g (P<0.05) and 203.7  $\pm$  13.5 g (P<0.05) when compared to Group II treated with 40% ethanol, respectively.

40% ethanol treatment in Group II significantly increased the relative liver weight to (P<0.001) 5.44  $\pm$  0.52 / 100g body weight when compared to Group I (Control) 3.05  $\pm$  0.27/100 g body weight. Administration of 200 and 400 mg/kg BOE significantly reduced (P<0.01 and P<0.001) the relative liver weight to 3.85  $\pm$  0.43 and 2.69  $\pm$  0.25 g body weight, respectively, compared to 5.44  $\pm$  0.52 g in 40% ethanol treatment. BOE treated group IV activity was less to standard silymarintreated group V rats at the concentration used **Table 2**.

TABLE 2: EFFECT OF BIXA ORELLANA SEED EXTRACT ON BODY WEIGHT, LIVER WEIGHT AND RELATIVE LIVER WEIGHT OF CONTROL AND ETHANOL INDUCED HEPATOTOXICITY IN RATS

Treatment	Initial body	Final body weight	Liver weight (g)	Relative liver wt. (liver
	weight (g)	(g)		weight/100g b.w.)
Control	$184.2 \pm 6.9$	$201.2 \pm 11.3$	$6.15 \pm 0.42$	$3.05 \pm 0.27$
40% ethanol	$180.5 \pm 8.4$	$165.7 \pm 10.4^{\#}$	$9.02 \pm 0.64^{\#}$	$5.44 \pm 0.52^{##}$
12ml/kg				
B. orellana	$179.4 \pm 9.3$	$191.5 \pm 12.4^*$	$7.39 \pm 0.58^*$	$3.85 \pm 0.43^{**}$
200 mg/kg				
B. orellana	$186.6 \pm 8.5$	$203.7 \pm 13.5^*$	$6.21 \pm 0.42^{**}$	$2.69 \pm 0.25^{***}$
400 mg/kg				
Silymarin	$183.2 \pm 6.7$	$202.8 \pm 14.7^{**}$	$6.11 \pm 0.57^{**}$	$3.01 \pm 0.32^{***}$
	Control 40% ethanol 12ml/kg B. orellana 200 mg/kg B. orellana 400 mg/kg	weight (g)       Control $184.2 \pm 6.9$ $40\%$ ethanol $180.5 \pm 8.4$ $12\text{ml/kg}$ $179.4 \pm 9.3$ $200 \text{ mg/kg}$ $186.6 \pm 8.5$ $400 \text{ mg/kg}$ $186.6 \pm 8.5$	weight (g)(g)Control $184.2 \pm 6.9$ $201.2 \pm 11.3$ $40\%$ ethanol $180.5 \pm 8.4$ $165.7 \pm 10.4^{\#}$ $12\text{ml/kg}$ $179.4 \pm 9.3$ $191.5 \pm 12.4^{*}$ $200 \text{ mg/kg}$ $186.6 \pm 8.5$ $203.7 \pm 13.5^{*}$ $400 \text{ mg/kg}$ $183.2 \pm 6.7$ $202.8 \pm 14.7^{**}$	weight (g)         (g)           Control $184.2 \pm 6.9$ $201.2 \pm 11.3$ $6.15 \pm 0.42$ 40% ethanol $180.5 \pm 8.4$ $165.7 \pm 10.4^{\#}$ $9.02 \pm 0.64^{\#}$ $12\text{ml/kg}$ $B. \text{ orellana}$ $179.4 \pm 9.3$ $191.5 \pm 12.4^{*}$ $7.39 \pm 0.58^{*}$ $200 \text{ mg/kg}$ $B. \text{ orellana}$ $186.6 \pm 8.5$ $203.7 \pm 13.5^{*}$ $6.21 \pm 0.42^{**}$ $400 \text{ mg/kg}$

Values are expressed as mean  $\pm$  SEM of 5 rats in each group. P values:  $^{*}$ <0.01,  $^{**}$ <0.001 compared with respective control group I. P values:  $^{*}$ <0.05,  $^{**}$ <0.01 and  $^{***}$ <0.001 compared with group II (Ethanol).

Effect of 50% Ethanolic Extract of Bixa orellana on Liver Injury in Control and Ethanol -Induced Hepatotoxicity in Rats: The effect of BOE on liver injury is shown in **Fig. 3**, **4** and **5**.

Ethanol-treated Group II rats showed increased serum aspartate transaminase (AST- 319.89 ± 30.57 U/l, P<0.001), alanine transaminase (ALT - $185.66 \pm 13.71 \text{ U/l}, \text{ P} < 0.001), \text{ Alkaline Phosphate}$ (ALP- 201.51  $\pm$  13.54 U/l, P<0.001), Lactate Dehydrogenase (LDH- 701.36  $\pm$  26.33 U/l, P<0.001), Cholesterol (CHL-  $76.57 \pm 5.75 \text{ mg/dl}$ , P<0.001), Bilirubin (2.78  $\pm 0.27$ , P<0.001) and decreased Albumin level (ALB- 2.88 ± 0.09 gm/dl, P<0.001) when compared to control Group I rats  $(93.45 \pm 10.84 \text{ U/I}, 42.34 \pm 7.34 \text{ U/I}, 74.58 \pm 9.51)$ 

U/l,  $371.14 \pm 31.02$  U/l,  $4.06 \pm 0.12$  gm/dl, 32.14 $\pm 6.11$  mg/dl and  $0.61 \pm 0.13$  mg/dl), respectively.

The BOE treated groups III and IV rats at 200 and 400 mg/kg significantly decreased AST (241.87 ± 19.87 and 120.23  $\pm$  12.85 U/l, P<0.01 and P<0.001), ALT (98.07  $\pm$  10.23 and 50.84  $\pm$  5.41 U/l, P<0.01 and P<0.001), ALP (102.12  $\pm$  11.21 and 90.17  $\pm$  70.23 U/l, P<0.01 and P<0.001), LDH  $(521.45 \pm 14.24 \text{ and } 397.51 \pm 17.15\text{U/I}, P<0.01 \text{ and }$ P<0.001), CHL (52.18  $\pm$  5.89 and 39.28  $\pm$  3.01 mg/dl, P<0.01 and P<0.001) and Bilirubin (1.37  $\pm$ 0.17 and  $0.86 \pm 0.12$  mg/dl, P<0.01 and P<0.001), and significantly increased the ALB (3.72  $\pm$  0.11 and  $43.93 \pm 0.08$  gm/dl, P<0.01 and P<0.001) respectively, when compared to Group II animals.

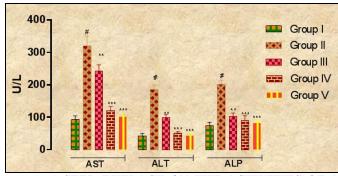


FIG. 3: EFFECT OF BIXA ORELLANA SEED EXTRACT ON THE ACTIVITIES OF ASPARTATE TRANSAMINASE (U/L), ALANINE TRANSAMINASE (U/L) AND ALKALINE PHOSPHATASE (U/L) AGAINST ETHANOL INDUCED **HEPATOTOXICITY IN RATS.** Values are expressed as mean ± S.E.M. of 5 rats in each group. P values: #<0.001 compared with respective control group I. P values: \*<0.05, \*\*<0.01, \*\*\*<0.001 compared with group II (ethanol).

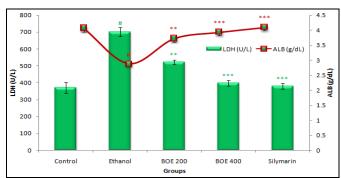
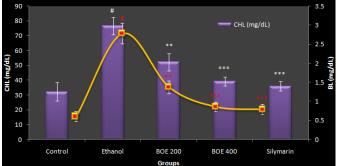


FIG. 4: EFFECT OF BIXA ORELLANA FRUIT SEED FIG. 5: EFFECT OF BIXA ORELLANA SEED EXTRACT ON EXTRACT ON LACTATE DEHYDROGENASE (LDH) AND THE ALBUMIN (ALB) AGAINST **ETHANOL** HEPATOTOXICITY IN RATS. Values are expressed as mean ± ETHANOL INDUCED HEPATOTOXICITY IN RATS. Values S.E.M. of 5 rats in each group. P values: #<0.001 compared with are expressed as mean ± S.E.M. of 5 rats in each group. P values: respective control group I; \*<0.05, \*\*<0.01, \*\*\*<0.001 compared #<0.001 compared with respective control group I; \*<0.05, with group II (ethanol).

Histopathological Studies: The histopathological evaluation of liver tissues of experimental groups (I-V) of rats (hematoxylin and eosin) was observed as described. (A) Liver section of normal control rat shows uniform hepatocytes with small vesicular nuclei and architecture of liver is well maintained.



ACTIVITY OF CHOLESTEROL (CHL) INDUCED BILIRUBIN (BLB) IN THE SERUM CONTROL AND \*\*<0.01, \*\*\*<0.001 compared with group II (ethanol).

(B) Liver section of ethanol treated rats showed smaller hepatocytes and eosinophilic cytoplasmic with indistinct cell boundaries showing proliferation. Architecture not well maintained but vascularity has relatively increased (indicated by arrow). (C) Liver section of rats treated ethanol and

200 mg/kg of BOE indicates less inflammatory cells, mild focal necrosis with sinusoidal dilatation. (D) Liver section of rats treated ethanol and 400 mg/kg of BOE showed normal vesicular nuclei and abundant eosinophilic cytoplasm with distinct cell

boundaries. Architecture well maintained with normal interstitial cells and vascularity. (E) Liver section of rats treated ethanol and 100 mg/kg of Silymarin showing normal morphology of hepatocyte.

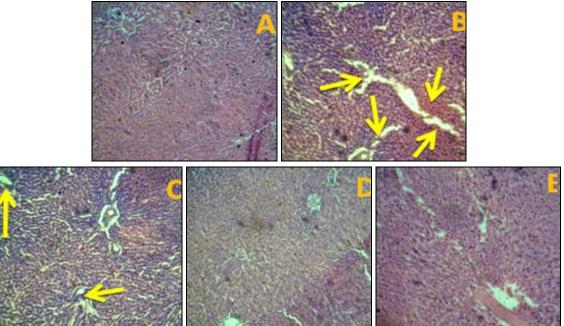


FIG. 6: HISTOPATHOLOGY OF LIVER TISSUES (HEMATOXYLIN AND EOSIN). (A) LIVER SECTION OF NORMAL CONTROL (B) LIVER SECTION OF ETHANOL TREATED RATS GROUP II (C) LIVER SECTION OF RATS TREATED ETHANOL AND 200 mg/kg OF BOE GROUP III (D) LIVER SECTION OF RATS TREATED ETHANOL AND 400 mg/kg OF BOE GROUP IV(E) LIVER SECTION OF RATS TREATED ETHANOL AND 100 mg/kg OF SILYMARIN GROUP V

**DISCUSSION:** Liver disease due to alcohol consumption is a common cause of death in adults; medicinal support to alcohol-induced liver dysfunction is indeed very meager. In order to reflect the protective effect of *Bixa orellana*, we established the alcohol-induced acute liver injury model in experiment. In the present study, the *Bixa orellana* extract (BOE) was observed to exhibit hepatoprotective effect by using 40% ethanol induced hepatotoxicity in Wistar rat.

Ethanol-induced acute liver injury demonstrated that there was a significant increase in aspartate aminotransferase (AST), alanine aminotransferase (ALT), alkaline phosphatase (ALP), Bilirubin (BIL), Cholesterol (CHL), lactate dehydrogenase and decrease in albumin (LDH) (ALB) concentrations in ethanol induced toxicity (Group II) animals. An augment in the serum levels of these marker enzymes occurred due to the enzymes leakage from liver into the circulation as a consequence of liver damage <sup>19</sup>.

The AST and ALT actions are responsive indicators of acute hepatic necrosis, and the ALP level is indicator of hepatobiliary disease <sup>20</sup>. Significant reduction in the levels of AST, ALT, ALP, LDH, CHL, Bilirubin and increase in the level of ALB towards the normal value was an indication of the stabilization of plasma membrane and the repair of hepatic tissue. Histological examination of the liver sections revealed lipid change of hepatocytes treated with 40% alcohol, while in the sections obtained from the rats treated with BOE and with alcohol, the lipid change of hepatocytes was alleviated.

Ethanol consumption leads to the production of excessive free radical inside the living system. This is due to the fact that ethanol is extensively metabolized by the microsomal oxidizing system to acetaldehyde and 1-hydroxyethyl radicals by cytochrome  $P_{450}$  II E1. These free radicals cause peroxidative degradation in the adipose tissue resulting in fatty infiltration of the hepatocytes.

Ethanol is extensively metabolized to acetaldehyde in the liver by the enzyme alcohol dehydrogenase. Acetaldehyde is further oxidized to acetate by acetaldehyde dehydrogenase/oxidase, leading to the generation of reactive oxygen species (ROS) <sup>21</sup>. Further oxidations in alcohol metabolism are accompanied by an excessive reduction of nicotinamide adenine dinucleotide (NAD), with a shift in NADH / NAD ratio. Under normal circumstances, reduction of NAD (changing to NADH) is finely regulated by the cell Krebs cycle. These reactive species oxidize cellular biomolecules, such as proteins and DNA and initiate membrane peroxidative degradation in the adipose tissue resulting in fatty infiltration of the hepatocytes and causing impairment of gluconeogenesis and diversion of metabolism to ketogenesis and fatty acid synthesis <sup>22</sup>.

The toxic effect of ethanol was controlled in the animals treated with ethanolic seed extract of B. orellana at the dose of 200 and 400 mg/kg respectively, by restoring the levels of liver function, indicating its protective activity against liver damage Fig. 3, 4, 5. This protective effect could be possibly due to the reduction in the tissue damage brought by the ethanolic extract of B. orellana. The results were compared with the standard drug Silymarin (100 mg/kg). The increase in the levels of serum bilirubin reflects the depth of jaundice as the clear indication of cellular leakage and loss of functional integrity of cell membrane. Bilirubin is the conventional indicator of liver diseases, restoration of total bilirubin levels may be due to the inhibitory effects on cytochrome P<sub>450</sub> resulting in the hindrence of the formation of hepatotoxic free radicals <sup>21</sup>. A marked elevation was observed in serum bilirubin level (SBL) of ethanol treated groups, whereas albumin level in the serum was markedly decreased. A reduction in synthesizing proteins was seen intoxication of the liver with hepatotoxicants. As seen in the silymarin treated group and ethanolic extract of B. orellana, all studied parameters were restored to normal conditions from the abnormal ones.

The protective effect of *B. orellana* may also be attributed to its antioxidant activity as evaluated by Chisté *et al.*, and Bell *et al.*, <sup>23, 24</sup> for acetaminophen induced- oxidative stress causing hepatic damage in

rats. Therefore, from the results it is clear that *Bixa orellana* seed extract has shown dose dependent activity among which at the dose level of 400mg/kg, p.o. shows greater activity that is comparable with the control and Silymarin treated groups (Standard).

CONCLUSION: Concluding from the above results it indicates that the ethanolic seed extract of Bixa orellana supplementation could antagonize the development of oxidative liver injury induced by acute ethanol exposure in Wistar rats. BOE (200 and 400 mg/kg) administration significantly the ethanol-induced liver injury. attenuated Furthermore, BOE supplementation prevented the acute ethanol-induced enhancement of hepatic enzymes. Liver histopathology images evidenced that BOE attenuated the hepatocellular necrosis and led to reduction in inflammatory cells infiltration, which may be attributed to its hepatoprotective effects.

The hepatoprotective effects of the ethanolic seed extract of *Bixa orellana* possesses antioxidant activities as proved earlier and ameliorate hepatic function of hepatic injury induced by alcohol. Clinically, application of *B. orellana* extract for liver protection against liver injury induced by ethanol in patient demands further investigation, yet proving to be the promising herbal drug to be included in the list of herbal drugs against alcohol induced hepatotoxicity.

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