



Received on 25 June 2018; received in revised form, 08 October 2018; accepted, 20 October 2018; published 01 February 2019

HIGH-DENSITY LIPOPROTEIN: ROLE IN REVERSE CHOLESTEROL TRANSPORT

D. M. Menge¹, N. K. Nair¹, T. P. Varghese¹ and P. R. A. Vijayakumar^{* 2}

Department of Pharmacy Practice¹, Department of Pharmacology², JSS College of Pharmacy, JSS Academy of Higher Education & Research, Ooty - 643001, Tamil Nadu, India.

Keywords:

Apolipoprotein A-I, ATP binding cassette transporter type B class 1, Cholesteryl ester transfer protein, High-density lipoprotein, Reverse cholesterol transport

Correspondence to Author:

P. R. Anand Vijayakumar

Professor,
Department of Pharmacology,
JSS College of Pharmacy,
JSS Academy of Higher Education
& Research, Ooty - 643001,
Tamil Nadu, India.

E-mail: ootyand2004@gmail.com

ABSTRACT: High-density lipoprotein (HDL) mediates reverse cholesterol transport (RCT) through its potential to accept excess cholesterol from extrahepatic tissues, and delivers it to the liver for breakdown and excretion. HDL also has pleiotropic properties such as anti-apoptosis, anti-inflammation, and capacity to remove oxidized sterols and phospholipids from the circulation. HDL is composed of apolipoprotein A-I, the major lipoprotein and apolipoprotein A-II, the minor lipoprotein. For HDL to carry out reverse cholesterol transport, it has to transform to make it suitable for acceptance of cholesterol and phospholipids. Therefore, in this review, we explore the composition and the various sizes of HDL. Further, we review the biosynthesis and remodeling of HDL by various proteins, enzymes and receptors such as ATP-binding cassette transporter class B-1 (ABCA1), endothelial lipase (EL), scavenger receptor class B type I (SR-BI), cholesteryl ester transfer protein (CETP), lecithin cholesterol acyltransferase LCAT), and phospholipids transfer protein (PLTP). Finally, we describe the pathways involved in the removal of cholesterol from the peripheral tissue and the current therapeutic strategies to increase levels of HDL-C as well as their outcomes.

INTRODUCTION: Reverse cholesterol transport (RCT) is a concept that was described by John Glomset as the removal of cholesterol from peripheral tissues back to the liver. In his work, he noticed that excess cholesterol from the peripheral tissues was delivered to the liver for excretion, which was attributed to protection conferred against cardiovascular disease¹⁻⁴. This idea concurs with studies that have linked low levels of HDL to increased incidence of cardiovascular disease^{5, 6}. Therefore, this review provides an insight into how high-density lipoprotein (HDL) undergoes different structural transformations while transporting cholesterol and triglycerides.

This review also highlights the various factors that are involved in remodeling HDL, as well as the different pathways that are involved in cholesterol efflux, which in turn may encourage research into how high-density lipoprotein could be used in future therapies.

High - Density Lipoprotein: High - density lipoproteins are a group of lipoproteins composed of lipids and lipoproteins with sizes ranging from 7.5 nm to 15 nm. HDLs can be classified into different categories based on their electrophoretic mobility, density, particle size, and apolipoprotein composition⁸, and may assume spherical or discoidal shapes⁹. The concentrations of lipid and apolipoprotein contents in HDL particles differ, thus, enabling them to interact with receptors and other transport proteins⁷. The number of apolipoproteins and the volume of cholesterol esters determine the size of HDL particle⁸. HDL₂ and HDL₃ are the main types of high-density lipoprotein found in circulation.

QUICK RESPONSE CODE 	DOI: 10.13040/IJPSR.0975-8232.10(2).481-88
	The article can be accessed online on www.ijpsr.com
DOI link: http://dx.doi.org/10.13040/IJPSR.0975-8232.10(2).481-88	

HDL₂ particles are larger and less-dense and have a density ranging between 1.063 g/mL - 1.125 g/mL while HDL₃ particles are smaller and denser and have a density ranging from 1.125-1.21 g/mL. HDL particles are heterogeneous and may carry other lipoproteins such as apo A-II, apo A-IV, apo E, and apo C. Lipid-poor apo A-I, and discoidal HDL particles have pre- β migration while spherical HDL particles have α -mobility.

Pleiotropic Activities of HDL:

- Removal of oxidized sterols and phospholipids^{10,11}.
- Endothelial cell protection and vasodilating effects^{12,13}.
- Anti-inflammatory properties¹⁴⁻¹⁶.
- Anti-apoptotic properties¹⁵.

HDL Remodelling: There are several proteins, enzymes, and receptors involved in remodeling HDL:

- Apolipoprotein A-I (apo A-I).
- Lecithin cholesterol acyltransferase (LCAT)
- ATP binding cassette transporter class A1 (ABCA1).
- Cholesterol ester transfer protein (CETP).
- Hepatic lipase (HL).
- Phospholipid transfer protein (PLTP).
- Scavenger receptor class B type I (SR-BI).

Apolipoprotein A-I: There are five major lipoproteins found in our bodies: chylomicrons, VLDL, IDL, LDL, and HDL. The high proportion of proteins in HDL make it dense and the smallest of the five lipoproteins⁶. Apolipoprotein A-I (apo A-I) is the most abundant apolipoprotein in HDL making up about 60% of the total volume of HDL. On the other hand, apo A-II is found Apo A-II is found in relatively smaller quantities⁸ see **Table 1**. Apo A-I secretion is often the first step in the biogenesis of HDL. Apo A-I is majorly synthesized in the liver, and a small quantity from the intestines then secreted into plasma. This apolipoprotein is involved in the initial stage of HDL synthesis¹⁷. It can also be dissociated from chylomicrons and very low-density lipoproteins during lipoprotein-mediated hydrolysis of triglycerides. Inter-conversion of HDL₂ and HDL₃ in the presence of proteins such as CETP, HL, and PLTP generates

apo A-I to^{18,19}. This apo A-I formed has a high affinity for cholesterol. It acquires cholesterol and phospholipids through cholesterol efflux. Lipolysis of triglycerides also provides additional cholesterol and phospholipids⁴.

ATP binding cassette transporter class A1(ABCA1) mediates the transfer of cholesterol on to apo A-I^{20,21}. Lecithin cholesterol acyltransferase (LCAT) converts this unesterified cholesterol into cholesteryl esters to form a hydrophobic core of HDL. ABCA1 enhances apo A-I binding and cell association in endothelial cells²²⁻²⁴. Interaction of this lipid-poor apo A-I and ABCA1 triggers cholesterol efflux in macrophages and fibroblasts, which internalize the lipid-poor apo A-I and resecret it as a lipidated, cholesterol-rich apolipoprotein, in a process known as retroendocytosis²⁵. Apo A-I and apo A-II makes up for about 70% and 20% respectively of the apolipoprotein content²⁶. Three forms of apo A-I have been identified. The first form exists as a component of spherical, α -migrating HDL particle, while the second form of LCAT which is a component of pre-beta migrating discoidal HDL. This form is rapidly converted, resulting in a very low concentration in plasma. The last form is the pre-beta migrating, lipid-poor apo A-I²⁶.

TABLE 1: FUNCTIONS AND MOLECULAR WEIGHTS OF VARIOUS APOLIPOPROTEINS

Apolipoprotein	The molecular weight (D)	Functions
Apo A-I	28,000	Activates LCAT The site of HDL binding to SR-BI A major component of HDL (approx.: 60%)
Apo A-II	17,000	Inhibits hepatic lipase The binding site of HDL A minor component of HDL (approx.: 30%)
Apo A-IV	46,000	Activates LCAT Stimulates cholesterol efflux Controls LPL
Apo C-I	6,600	Activates LCAT
Apo C-II	9,000	Inhibits hepatic TGRL uptake Activates LPL Inhibits hepatic uptake of TGRL
Apo D	33,000	Modulates the activity of LCAT
Apo E	34,000	The binding site of HDL Stimulates cholesterol efflux Mobilizes cholesterol in macrophages

Lecithin: Cholesterol Acyltransferase: LCAT is a glycoprotein found in the liver, testes and central nervous system²⁷, and it mediates the transfer of 2-acyl groups from lecithin to the free cholesterol, forming cholesteryl ester and lysolecithin²⁸. The cholesteryl ester formed partitions into the core of a discoidal HDL particle, thus triggers uptake of cholesterol from any cholesterol donor to form pre- β migrating HDL²⁹. The LCAT reaction accounts for most of the cholesteryl esters in circulation.

Discoidal HDL is a suitable substrate of LCAT and can accept more cholesterol from the peripheral tissues than other apolipoproteins. Also, cholesteryl esters are hydrophobic, and they migrate from the surface of the particle to the core. This migration results in the transition from discoidal particle to a spherical shaped particle²⁸.

ATP Binding Cassette Transporter A1: ABCA1 is expressed in liver, lungs, adrenal glands, fetal tissues, and placenta. ABCA1 acts as a rate-limiting step in the biogenesis of HDL^{30, 31}. ATP generates the energy needed to transport cholesterol, phospholipids, vitamins, and cytotoxins between different cellular compartments. ABCA1 also facilitates cholesterol efflux to the monomolecular pre-beta migrating, lipid-poor apo A-1.

Targeted disruption of the ABCA1 gene in mice results in almost undetectable levels of HDL-C and apo A-1^{32, 33}. This study suggests that ABCA1 plays an important role in modulating apo A-I binding and cell association. Suppression of ABCA1 in bovine aortic endothelial cells through RNA interference decreases apo A-I binding and cell association³⁴.

Tangier's disease, a mutation in the functional gene which encodes ABCA1 impairs lipidation causing hypercatabolism of lipid of lipid-free apo A-I and consequently resulting in substantially low HDL-C levels^{20, 35, 36}. For example, in patients with Tangier's disease, fibroblasts and macrophages rich in cholesterol do not release free cholesterol and phospholipids to the lipid-free apo A-I^{37, 38}.

Cholesteryl Ester Transfer Protein: CETP is a hydrophobic glycoprotein produced in the liver, spleen, and adipose tissue. It mediates the mass transfer of cholesteryl esters from HDL to apo B

containing lipoproteins such as LDL, VLDL, and IDL in exchange for triglycerides³⁹. The exchanged triglycerides form the core of the HDL particle, thus, act as a suitable substrate for hepatic lipase⁴⁰. Moreover, triglycerides have a greater molecular weight than CEs; therefore, HDL particles formed are increased size. In addition, hepatic lipase along with CETP convert larger HDL₂ into smaller and denser HDL₃ particles releasing lipid-free apo A-I and fatty acids that can be taken up by tissues.

Transgenic expression of CETP in mice that naturally lack this protein results in increased HDL levels⁴¹. CETP has a net effect of reducing HDL-C levels in hypertriglyceridemic individuals because it is thought that the high levels of triglycerides activate CETP which in turn reduces the levels of HDL⁴².

Phospholipid Transfer Protein: PLTP catalyzes the transfer of phospholipids from the surfaces of triglyceride-rich lipoproteins to HDL during lipolysis which is a necessary step during remodeling of pre- β HDL particles⁴³. PLTP can be found in kidneys, lungs, heart, skeletal muscles, and the brain. It regulates the sizes of HDL particles by converting them into smaller or larger particles⁴⁴. What's more, ABCA1 interacts with PLTP to enhance cholesterol efflux in the body. Therefore, a deficiency in PLTP results in decreased levels of HDL-C and phospholipids⁴⁵.

Hepatic Lipase: HL is synthesized in the liver and is bound to the sinusoid capillaries of the liver⁴⁶. It is involved in remodeling HDL and other lipoproteins such as LDL, VLDL, and IDL⁴⁷. Furthermore, HL has a high specificity for phospholipids and triglycerides⁴⁸ and consequently, hydrolyzes triglycerides and phospholipids present in larger lipid-rich HDL₂ particles to form smaller HDL₃ particles^{49, 50}.

Endothelial Lipase: EL is synthesized in the endothelial cells and liver and acts locally at the site of synthesis⁵¹. It is also expressed in other tissues such as lungs, kidneys, testes, thyroid gland, ovaries, hepatocytes, and placenta⁴⁹. Deleting functional endothelial lipase in mice results in attenuated atherosclerotic lesions⁵². Inhibition of EL in mice increases HDL-C *in-vivo*.

On the contrary, over-expression of EL decreases the levels of HDL-C^{53, 54}. EL concentrations are inversely associated with HDL-C levels in people with metabolic syndrome⁵⁵.

Scavenger Receptor Class B Type 1: SR-BI is a glycoprotein expressed in the liver, endothelial cells, adrenal glands, ovary, and testes. It mediates selective uptake of HDL cholesteryl esters by the liver⁵⁶. Through its action, the liver takes up lipids too. Over-expression of SR-BI receptors in the liver of mice increases uptake of HDL cholesteryl esters resulting in increased catabolism of the cholesteryl esters and an eventual decrease in HDL-CEs in circulation⁵⁷⁻⁵⁹. SR-BI over-expression reduces atherosclerosis⁵⁷.

Reverse Cholesterol Transport Pathways: The cholesterol delivered to the liver is broken down into neutral sterols and bile acids. The main pathways of RCT include:

- Direct delivery of HDL-C to the liver *via* SR-BI
- CETP-mediated transfer of HDL-C to the liver
- Esterification of cholesterol by LCAT
- Cholesterol efflux from extrahepatic tissues to the plasma *via* ABCA1

Direct Delivery of HDL-C to the Liver *via* SR-BI: SR-BI receptors are present in the liver. These receptors influence the uptake of HDL-C esters by the liver, with HDL₂ having a better binding affinity than HDL₃⁶⁰. SR-BI plays a crucial role in cholesterol efflux.

CETP-Mediated Transfer of HDL-C to the Liver: This is an indirect pathway of delivery of cholesterol to the liver. CETP mediates the exchange of cholesteryl ester present on HDL for triglycerides from VLDL and LDL lipoproteins, enriching the apo B containing lipoproteins with cholesterol. The triglycerides present in the HDL particle are then hydrolyzed by hepatic lipase, thus converting the larger HDL₂ particle into the smaller HDL₃ particle. SR-BI receptors then mediate the delivery of HDL₂ to the liver. The cholesterol-rich apo-B lipoproteins then deliver cholesterol to the liver *via* LDL receptors for excretion (See **Fig. 1**).

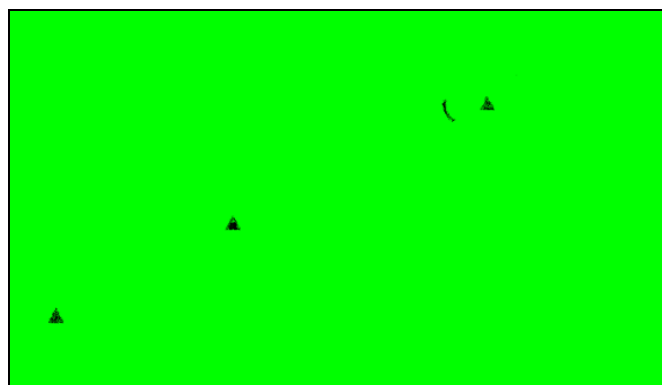


FIG. 1: A SCHEMATIC REPRESENTATION OF HDL-MEDIATED REVERSE CHOLESTEROL TRANSPORT

LCAT-Mediated Esterification of Cholesterol: LCAT esterifies free cholesterol on discoidal HDL, and as a result converts it into spherical HDL particle. Esterification of the FC creates space on the spherical HDL particle which can then take up cholesterol through other pathways such as diffusion, SR-BI, and ABCG1.

HDL can also obtain cholesterol from caveolae and lipid rafts present in the cell membrane, thus the HDL removes cholesterol from macrophages as well. All HDL particles can take up cholesterol.

ABCA1-Mediated Cholesterol Efflux from Peripheral Tissues to Plasma: Apo A-I is synthesized in the liver and secreted in plasma. In plasma, the apo A-I takes up phospholipids to form nascent pre- β -HDL. ABCA1 takes up cholesterol present in the cell and delivers it to the cell membrane. The lipid-poor apo A-I then interacts with cellular cholesterol to form discoidal HDL. ABCA1 also mediates the transfer of cholesterol from macrophages to HDL for excretion.

Therapeutic Strategies to Increase HDL Levels: Therapeutic interventions such as the use of CETP inhibitors and niacin have shown a mild increase in HDL-C levels in the blood.

Niacin: Nicotinic acid is the oldest known lipid-lowering agent that acts by reducing the synthesis of VLDL in the liver by its action on Diacylglycerol O-acyltransferase-2 (DGAT-2). Niacin, when used in combination with statin or bile acid-binding resin, increases the cholesterol levels in HDL While it decreases cholesterol in LDL and VLDL⁶¹. Patients with established coronary artery disease, and were on treatment with high-dose statins, experienced significant

improvements in their lipid profiles when extended-release niacin was added to the statin therapy. However, it is worth noting that these changes in lipid profile were not associated with improved endothelial function⁶².

Niacin use has been limited because of its perceived side effects. Laropiprant, a selective prostaglandin D2 receptor-1 antagonist, decreases flushing associated with extended-release niacin. A study in dyslipidemia patients that compared extended-release niacin/laropiprant in doses of 1g-2g showed significantly less flushing than in patients receiving gradually titrated doses of niacin extended-release⁶³. Flushing has been reported in more than 90% of the patients who used niacin as part of their lipid-lowering strategy. Other side effects include itching, hyperpigmentation, urticaria and ichthyosis⁶⁴.

Low-dose niacin strategy to minimize adverse effects has also been explored in patients with chronic kidney disease. The enrolled patients who had been on a fixed dose of 500 mg/day of niacin for six months were compared to another group of patients who had been taking a statin for 9 months. The niacin group of patients experienced a low frequency of adverse effects with the benefit of significantly raised HDL-C levels and decreased levels of LDL-C at 12 and 24 weeks compared to the baseline level⁶⁵.

CETP Inhibitors: Inhibition of CETP increases HDL-C cholesterol, and decreases potentially atherogenic and non-HDL particles, by retention of cholesterol in the HDL fraction. With this concept, CETP inhibitors have been used in clinical trials with the hope that they would increase the levels of HDL-C which in turn would confer protection against atherosclerosis. However, there is lack of direct clinical evidence that raising HDL-C levels translates to protection against cardiovascular events.

In the ILLUMINATE trial, patients received either torcetrapib plus atorvastatin or atorvastatin alone over 12 months. Torcetrapib increased HDL-C levels by 72.1% and decreased LDL-C by 24.9%, but the trial was terminated prematurely because of increased risk of mortality and morbidity by an unknown mechanism⁶⁶.

A study evaluated evacetrapib monotherapy or in combination with statins, against statin monotherapy or placebo in 338 dyslipidemia patients. Evacetrapib combination with statins, when compared to placebo or statin monotherapy, increased HDL-C levels and decreased LDL-C levels. Interestingly, a combination of statin and evacetrapib resulted in greater reductions in LDL-C, but not a greater increase in HDL-C when compared to evacetrapib monotherapy⁶⁷. These results are similar to a study conducted in Japanese patients with dyslipidemia, where evacetrapib monotherapy or combination with atorvastatin decreased LDL-C levels and increased HDL-C levels after 12 weeks⁶⁸. Studies evaluating the efficacy of evacetrapib were terminated prematurely because it did not lower the rate of cardiovascular events when compared to placebo⁶⁹.

Dalcetrapib, a less potent CETP inhibitor than evacetrapib, was given to patients with low target levels of LDL-C to investigate its effects on endothelial function, lipid levels, and blood pressure. CETP activity after 36 weeks of treatment decreased by 56% when compared to placebo, HDL-C increased by 31% when compared to placebo. On the contrary, LDL-C levels did not change⁷⁰.

CONCLUSION: Removal of cholesterol from the peripheral tissues is an important physiological step in the body. Excess cholesterol can accumulate in blood vessels resulting in atherosclerosis. These atherosclerotic plaques pose cardiovascular risks, which could ultimately lead to coronary artery disease, stroke, and myocardial infarction. To prevent or decrease the likelihood of these cardiovascular events, raising the levels of 'good cholesterol' with the help of HDL has been a topic of focus in recent research. Many observational studies have shown that raising HDL-C could prevent these events. However, the clinical trials conducted till date has not prevented CVD events by raising HDL-C levels. CETP inhibitors such as evacetrapib, anacetrapib, torcetrapib, and dalcetrapib have been used extensively in clinical trials to raise HDL-C levels and lower LDL-C and triglycerides but have failed to prevent or lower the risk of cardiovascular events. On the other hand, niacin raises HDL-C levels, but its use has been

limited because of flushing. Therefore, further research is needed to understand how HDL and therapies to raise HDL could be used to prevent these events.

ACKNOWLEDGEMENT: Nil

DECLARATION OF INTEREST: The authors declare that they have no competing interests.

SUBMISSION DECLARATION AND VERIFICATION: This review article is not under consideration for publication elsewhere and is read and approved by all authors.

REFERENCES:

1. Glomset JA: The plasma lecithin: cholesterol acyl-transferase reaction. *The Journal of Lipid Research* 1968; 9(2): 155-167.
2. Ansell BJ, Watson KE, Fogelman AM, Navab M and Fonarow GC: High-Density lipoprotein function recent advances. *Journal of American College of Cardiology* 2005; 46(10): 1792-1798.
3. Gordon DJ, Probstfield JL, Garrison RJ, Neaton JD, Castelli WP, Knoke JD, Jacobs DR Jr, Bangdiwala S and Tyroler HA: High-density lipoprotein cholesterol and cardiovascular disease. Four prospective American studies. *Circulation* 1989; 79(1): 8-15.
4. Khera AV, Cuchel M, de la Llera-Moya M, Rodrigues A, Burke MF, Jafri K, French BC, Phillips JA, Mucksavage ML, Wilensky RL, Mohler ER, Rothblat GH and Rader DJ: Cholesterol efflux capacity, high-density lipoprotein function, and atherosclerosis. *The New England Journal of Medicine* 2011; 364(2): 127-135.
5. Santos-Gallego CG, Badimon JJ and Rosenson RS: Beginning to understand high-density lipoproteins. *Endocrinology Metabolism Clinics of North America* 2014; 43(4): 913-47.
6. Yetukuri L, Söderlund S, Koivuniemi A, Seppänen-Laakso T, Niemelä PS, Hyvönen M, Taskinen MR, Vattulainen I, Jauhiainen M and Oresic M: Composition and lipid spatial distribution of HDL particles in subjects with low and high HDL-cholesterol. *The Journal of Lipid Research* 2010; 51(8): 2341-2351.
7. Brown WV: High-density lipoprotein and transport of cholesterol and triglyceride in the blood. *Journal of Clinical Lipidology* 2007; 1(1): 7-19.
8. McGrowder D, Riley C, Morrison EYSA and Gordon L: The role of high-density lipoproteins in reducing the risk of vascular diseases, neurogenerative disorders, and cancer. *Cholesterol* 2011; Article ID 496925, 9 pages.
9. Rosenson RS, Brewer HB Jr, Ansell B, Barter P, Chapman MJ, Heinecke JW, Kontush A, Tall AR and Webb NR: Translation of high-density lipoprotein function into clinical practice current prospects and future challenges. *Circulation* 2013; 128(11): 1256-1267.
10. Acton SL, Kozarsky KF and Rigotti A: The HDL receptor SR-BI: A new therapeutic target for atherosclerosis? *Molecular Medicine Today* 1999; 5(12): 518-524.
11. Soran H, Schofield JD and Durrington PN: Antioxidant properties of HDL. *Frontiers in Pharmacology* 2015; 6: 222.
12. Tran-Dinh A, Diallo D, Delbosc S, Varela-Perez LM, Dang QB, Lapergue B, Burillo E, Michel JB, Levoye A, Martin-Ventura JL and Meilhac O: HDL and endothelial protection. *British Journal of Pharmacology* 2013; 169(3): 493-511.
13. Perségol L, Darabi M, Dauteuille C, Lhomme M, Chantepie S, Rye KA, Therond P, Chapman MJ, Salvayre R, Nègre-Salvayre A, Lesnik P, Monier S and Kontush A: Small dense HDLs display potent vasorelaxing activity, reflecting their elevated content of sphingosine-1-phosphate. *The Journal of Lipid Research* 2018; 59(1): 25-34.
14. Rosenson RS, Brewer HB Jr, Ansell BJ, Barter P, Chapman MJ, Heinecke JW, Kontush A, Tall AR and Webb NR: Dysfunctional HDL and atherosclerotic cardiovascular disease. *Nature Reviews Cardiology* 2016; 13(1): 48-60.
15. Barter PJ: Antiinflammatory Properties of HDL. *Circulation Research* 2004; 95(8): 764-772.
16. Van-Lenten BJ, Wagner AC, Nayak DP, Hama S, Navab M and Fogelman M: High-density lipoprotein loses its anti-inflammatory properties during acute influenza infection. *Circulation* 2001; 103(18): 2283-2288.
17. Green PHR, Tall AR and Glickman RM: Rat intestine secretes discoid high-density lipoprotein. *Journal of Clinical Investigation* 1978; 61(2): 528.
18. Miyazaki O, Fukamachi I, Mori A, Hashimoto H, Kawashiri MA, Nohara A, Noguchi T, Inazu A, Yamagishi M, Mabuchi H and Kobayashi J: Formation of pre- β 1-HDL during lipolysis of triglyceride-rich lipoprotein. *Biochemical and Biophysical Research Communications* 2009; 379(1): 55-59.
19. Von-Eckardstein A, Jauhiainen M, Huang Y, Metso J, Langer C, Pussinen P, Wu S, Ehnholm C and Assmann G: Phospholipid transfer protein-mediated conversion of high-density lipoproteins generates pre-beta 1-HDL. *Biochimica et Biophysica Acta* 1996; 1301(3): 255-262.
20. Gelissen IC: ABCA1 and ABCG1 Synergize to Mediate Cholesterol Export to ApoA-I. *Arteriosclerosis, Thrombosis, and Vascular Biology* 2005; 26(3): 534-540.
21. Cavelier LB: Regulation and activity of the human abca1 gene in transgenic mice. *The Journal of Biological Chemistry* 2001; 276(21): 18046-18051.
22. Vedhachalam C, Ghering AB, Davidson WS, Lund-Katz S, Rothblat GH and Phillips MC: ABCA1-induced cell surface binding sites for ApoA-I. *Arteriosclerosis, Thrombosis, and Vascular Biology* 2007; 27(7): 1603-1609.
23. Wang N, Silver DL, Costet P and Tall AR: Specific binding of ApoA-I, enhanced cholesterol efflux, and altered plasma membrane morphology in cells expressing ABC1. *The Journal of Biological Chemistry* 2000; 275(42): 33053-33058.
24. Chambenoit O, Hamon Y, Marguet D, Rigneault H, Rosseneu M and Chimini G: Specific Docking of Apolipoprotein A-I at the Cell Surface Requires a Functional ABCA1 Transporter. *The Journal of Biological Chemistry* 2001; 276(13): 9955-9960.
25. Röhr C and Stangl H: HDL endocytosis and resecretion. *Biochimica et Biophysica Acta* 2013; 1831(11): 1626-1633.
26. Rye KA and Barter PJ: Formation and metabolism of pre-beta-migrating, lipid-poor apolipoprotein A-I. *Arteriosclerosis, Thrombosis, and Vascular Biology* 2004; 24(3): 421-428.

27. Jonas A: Lecithin-cholesterol acyltransferase in the metabolism of high-density lipoproteins. *The Journal Lipid Research* 1966; 7(5): 638-648.
28. Rader DJ, Alexander ET, Weibel GL, Billheimer J and Rothblat GH: The role of reverse cholesterol transport in animals and humans and relationship to atherosclerosis. *The Journal of Lipid Research* 2008; 50: 189-194.
29. Osei-Hwedieh DO, Amar M, Sviridov D and Remaley AT: Apolipoprotein mimetic peptides: Mechanisms of action as anti-atherogenic agents. *Pharmacology and Therapeutics* 2011; 130(1): 83-91.
30. Yvan-Charvet L, Wang N and Tall AR: Role of HDL, ABCA1, and ABCG1 transporters in cholesterol efflux and immune responses. *Arteriosclerosis, Thrombosis and Vascular Biology* 2010; 30(2): 139-143.
31. Umemoto T, Han CY, Mitra P, Averill MM, Tang C, Goodspeed L, Omer M, Subramanian S, Wang S, Den Hartigh LJ, Wei H, Kim EJ, Kim J, O'Brien KD and Chait A: Apolipoprotein AI and high-density lipoprotein have anti-inflammatory effects on adipocytes *via* cholesterol transporters ATP-binding cassette A-1, ATP-binding cassette G-1, and scavenger receptor B-1. *Circulation Research* 2013; 112(10): 1345-1354.
32. McNeish J, Aiello RJ, Guyot D, Turi T, Gabel C, Aldinger C, Hoppe KL, Roach ML, Royer LJ, de Wet J, Broccardo C, Chimini G and Francone OL: High-density lipoprotein deficiency and foam cell accumulation in mice with targeted disruption of ATP-binding cassette transporter-1. *Proceedings of the National Academy of Sciences of the United States of America* 2000; 97(8): 4245-4250.
33. Chung S, Timmins JM, Duong M, Degirolamo C, Rong S, Sawyer JK, Singaraja RR, Hayden MR, Maeda N, Rudel LL, Shelness GS and Parks JS: Targeted deletion of hepatocyte ABCA1 leads to very low-density lipoprotein triglyceride overproduction and low-density lipoprotein hypercatabolism. *The Journal of Biological Chemistry* 2010; 285(16): 12197-12209.
34. Rohrer L, Ohnsorg PM, Lehner M, Landolt F, Rinninger F and Von-Eckardstein A: High-density lipoprotein transport through aortic endothelial cells involves scavenger receptor BI and ATP-binding cassette transporter G1. *Circulation Research* 2009; 104(10): 1142-1150.
35. Aiello RJ: Increased atherosclerosis in hyperlipidemic mice with inactivation of ABCA1 in macrophages. *Arteriosclerosis, Thrombosis, and Vascular Biology* 2002; 22(4): 630-637.
36. Oram JF: HDL Apolipoproteins and ABCA1: Partners in the removal of excess cellular cholesterol. *Arteriosclerosis, Thrombosis, and Vascular Biology* 2003; 23(5):720-727.
37. Yancey PG, Bielicki JK, Johnson WJ, Lund-Katz S, Palgunachari MN, Anantharamaiah GM, Segrest JP, Phillips MC and Rothblat GH: Efflux of cellular cholesterol and phospholipid to lipid-free apolipoproteins and class A amphipathic peptides. *Biochemistry* 1995; 34(24): 7955-7965.
38. Yancey PG: Importance of Different Pathways of Cellular Cholesterol Efflux. *Arteriosclerosis, Thrombosis and Vascular Biology* 2003; 23(5): 712-719.
39. Attie AD, Kastelein JP and Hayden MR: Pivotal role of ABCA1 in reverse cholesterol transport influencing HDL levels and susceptibility to atherosclerosis. *The Journal of Lipid Research* 2001; 42(11): 1717-1726.
40. Brunham LR and Hayden MR: Human genetics of HDL: Insight into particle metabolism and function. *Progress in Lipid Research* 2015; 58: 14-25.
41. Francone OL, Royer L and Haghpassand M: Increased pre-beta-HDL levels, cholesterol efflux, and LCAT-mediated esterification in mice expressing the human cholesteryl ester transfer protein (CETP) and human apolipoprotein A-I (apoA-I) transgenes. *The Journal of Lipid Research* 1996; 37(6): 1268-1277.
42. Yassine HN, Belopolskaya A, Schall C, Stump CS, Lau SS and Reaven PD: Enhanced cholesterol efflux to HDL through the ABCA1 transporter in hypertriglyceridemia of type 2 diabetes. *Metabolism* 2014; 63(5): 727-734.
43. Mulya A, Lee JY, Gebre AK, Boudyguina EY, Chung SK, Smith TL, Colvin PL, Jiang XC and Parks JS: Initial interaction of apoA-I with ABCA1 impacts *in-vivo* metabolic fate of nascent HDL. *The Journal Lipid Research* 2008; 49(11): 2390-2401.
44. Huuskonen J, Olkkonen VM, Jauhiainen M and Ehnholm C: The impact of phospholipid transfer protein (PLTP) on HDL metabolism. *Atherosclerosis* 2001; 155(2): 269-281.
45. Ehnholm S, van Dijk KW, van't Hof B, van der Zee A, Olkkonen VM, Jauhiainen M, Hofker M, Havekes L and Ehnholm C: Adenovirus-mediated overexpression of human phospholipid transfer protein alters plasma HDL levels in mice. *The Journal of Lipid Research* 1998; 39(6): 1248-1253.
46. Zambon A, Austin MA, Brown BG, Hokanson JE and Brunzell JD: Effect of hepatic lipase on LDL in normal men and those with coronary artery disease. *Arteriosclerosis, Thrombosis, and Vascular Biology* 1993; 13(2): 147-153.
47. Zambon A, Bertocco S, Vitturi N, Polentarutti V, Vianello D and Crepaldi G: Relevance of hepatic lipase to the metabolism of triacylglycerol-rich lipoproteins. *Biochemical Society Transactions* 2003; 31(5): 1070-1074.
48. Jin W, Marchadier D and Rader DJ: Lipases and HDL metabolism. *Trends in Endocrinology and Metabolism* 2002; 13(4): 174-178.
49. Annema W and Tietge UJF: Role of hepatic lipase and endothelial lipase in high-density lipoprotein-mediated reverse cholesterol transport. *Current Atherosclerosis Reports* 2011; 13(3): 257-265.
50. Connelly PW: The role of hepatic lipase in lipoprotein metabolism. *Clinica Chimica Acta* 1999; 286(1-2): 243-255.
51. Choi SY, Hirata K, Ishida T, Quertermous T and Cooper AD: Endothelial lipase: a new lipase on the block. *The Journal of Lipid Research* 2002; 43: 1763-1769.
52. Ishida T, Choi SY, Kundu RK, Spin J, Yamashita T, Hirata K, Kojima Y, Yokoyama M, Cooper AD and Quertermous T: Endothelial lipase modulates susceptibility to atherosclerosis in apolipoprotein-E-deficient mice. *The Journal of Biological Chemistry* 2004; 279(43): 45085-45092.
53. Jin W, Millar JS, Broedl U, Glick JM and Rader DJ: Inhibition of endothelial lipase causes increased HDL cholesterol levels *in-vivo*. *Journal of Clinical Investigation* 2003; 111(3): 357-362.
54. deLemos AS, Wolfe ML, Long CL, Sivapackianathan R and Rader DJ: Identification of Genetic Variants in Endothelial Lipase in Persons With Elevated High-Density Lipoprotein Cholesterol. *Circulation* 2002; 106(11): 1321-1326.
55. Badellino KO, Wolfe ML, Reilly MP and Rader DJ: Endothelial lipase concentrations are increased in metabolic syndrome and associated with coronary atherosclerosis. *PLoS Medicine* 2006; 3(2): 22.
56. Azhar S and Reaven E: Scavenger receptor class BI and selective cholesteryl ester uptake: Partners in the regulation of steroidogenesis. *Molecular and Cellular Endocrinology* 2002; 195(1-2): 1-26.

57. Kozarsky KF, Donahee MH, Rigotti A, Iqbal SN, Edelman ER and Krieger M: Overexpression of the HDL receptor SR-BI alters plasma HDL and bile cholesterol levels. *Nature* 1997; 387(6631): 414-417.
58. Wang N, Arai T, Ji Y, Rinninger F and Tall AR: Liver-specific overexpression of scavenger receptor BI decreases levels of very low-density lipoprotein ApoB, low-density lipoprotein ApoB, and high-density lipoprotein in transgenic mice. *The Journal of Biological Chemistry* 1998; 273(49): 32920-32926.
59. Ueda Y, Royer L, Gong E, Zhang J, Cooper PN, Francone O and Rubin EM: Lower plasma levels and accelerated clearance of high-density lipoprotein (HDL) and non-HDL cholesterol in scavenger receptor class B type I transgenic mice. *The Journal of Biological Chemistry* 1999; 274(11): 7165-7171.
60. de Beer MC, Durbin DM, Cai L, Jonas A, de Beer FC and van der Westhuyzen DR: Apolipoprotein A-I conformation markedly influences HDL interaction with scavenger receptor BI. *The Journal of Lipid Research* 2001; 42: 309-313.
61. Zambon A, Zhao XQ, Brown BG and Brunzell JD: Effects of niacin combination therapy with a statin or bile acid resin on lipoproteins and cardiovascular disease. *American Journal Cardiology* 2014; 113(9): 1494-1498.
62. Philpott AC, Hubacek J, Sun YC, Hillard D and Anderson TJ: Niacin improves lipid profile but not endothelial function in patients with coronary artery disease on high dose statin therapy. *Atherosclerosis* 2013; 226(2): 453-458.
63. Maccubbin D, Koren MJ, Davidson M, Gavish D, Pasternak RC, Macdonell G, Mallick M, Sisk CM, Paolini JF and Mitchel Y: Flushing profile of extended-release niacin/laropiprant versus gradually titrated niacin extended-release in patients with dyslipidemia with and without ischemic cardiovascular disease. *American Journal of Cardiology* 2009; 104(1): 74-81.
64. Guyton JR: Effect of niacin on atherosclerotic cardiovascular disease. *American Journal of Cardiology* 1998; 82(12): 18-23
65. Jin Kang H, Kim DK, Mi Lee S, Han Kim K, Hee Han S, Hyun Kim K, Eun Kim S, Ki Son Y and An WS: Effects of low-dose niacin on dyslipidemia and serum phosphorus in patients with chronic kidney disease. *Kidney Research and Clinical Practise* 2013; 32(1): 21-26.
66. Barter PJ, Caulfield, M and Eriksson M: Effects of Torcetrapib in Patients at High Risk for Coronary Events. *The New England Journal of Medicine* 2007; 357: 2109-2122.
67. Nicholls SJ, Brewer HB, Kastelein JJ, Krueger KA, Wang MD, Shao M, Hu B, McErlean E and Nissen SE: Effects of the cholesteryl ester transfer protein inhibitor, evacetrapib, administered as monotherapy or in combination with statins on cholesterol efflux and HDL particles in patients with dyslipidemia. *Circulation* 2014; 130(19): 2099-2109.
68. Teramoto T, Takeuchi M, Morisaki Y, Ruotolo G and Krueger KA: Efficacy, safety, tolerability, and pharmacokinetic profile of evacetrapib administered as monotherapy or in combination with atorvastatin in Japanese patients with dyslipidemia. *American Journal of Cardiology* 2014; 113(12): 2021-2029.
69. Lincoff AM, Nicholls SJ, Riesmeyer JS, Barter PJ, Brewer HB, Fox KAA, Gibson CM, Granger C, Menon V, Montalescot G, Rader D, Tall AR, McErlean E, Wolski K, Ruotolo G, Vangerow B, Weerakkody G, Goodman SG, Conde D, McGuire DK, Nicolau JC, Leiva-Pons JL, Pesant Y, Li W, Kandath D, Kouz S, Tahirkheli N, Mason D and Nissen SE: ACCELERATE Investigators: Evacetrapib and Cardiovascular Outcomes in High-Risk Vascular Disease. *The New England Journal of Medicine* 2017; 376(20): 1933-1942.
70. Lüscher TF, Taddei S, Kaski JC, Jukema JW, Kallend D, Münzel T, Kastelein JJ and Deanfield JE: dal-VESSEL Investigators: Vascular effects and safety of dalcetrapib in patients with or at risk of coronary heart disease: the dal-VESSEL randomized clinical trial. *European Heart Journal* 2012; 33(7): 857-865.
71. Wierzbicki AS, Hardman TC and Viljoen A: New lipid-lowering drugs: An update. *International Journal of Clinical Practice* 2012; 66(3): 270-280.
72. Min KL, Park MS, Jung J, Chang MJ and Kim CO: Comparison of pharmacokinetics and safety of a fixed-dose combination of rosuvastatin and ezetimibe versus separate tablets in healthy subjects. *Clinical Therapeutics* 2017; 39(9): 1799-1810.

How to cite this article:

Menge DM, Nair NK, Varghese TP and Vijayakumar PRA: High-density lipoprotein: role in reverse cholesterol transport. *Int J Pharm Sci & Res* 2019; 10(2): 481-88. doi: 10.13040/IJPSR.0975-8232.10(2).481-88.

All © 2013 are reserved by International Journal of Pharmaceutical Sciences and Research. This Journal licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported License.

This article can be downloaded to **ANDROID OS** based mobile. Scan QR Code using Code/Bar Scanner from your mobile. (Scanners are available on Google Playstore)