IJPSR (2020), Volume 11, Issue 1



INTERNATIONAL JOURNAL

(Research Article)

Received on 11 April 2019; received in revised form, 26 July 2019; accepted, 02 September 2019; published 01 January 2020

ESTIMATION OF WITHAFERIN – A FROM ASHWAGANDHADI LEHYA USING HPLC

A. Raval Manan^{*}, G. Gamit Kanan, Y. Vyas Niraj and D. Patel Nishit

Ramanbhai Patel College of Pharmacy, Charusat Campus, Changa, Petlad, Anan - 388421, Gujarat, India.

ABSTRACT: Ashwagandhadi lehya is an Ayurveda formulation, official in **Keywords:** Ayurvedic Formulary of India and used as aphrodisiac, tonic, rejuvenating Ashwagandhadi lehya, agents etc. It is one of the major components of Ashwagandhadi lehya is Withaferin-A, HPLC, Validation Withania somnifera which contain Withaferin -A. The study was undertaken to **Correspondence to Author:** develop an analytical method using HPLC to estimate Withaferin - A, being a A. Raval Manan marker from, methanolic extract of Ashwagandhadi lehya as one of the quality Ramanbhai Patel College of parameters. Method development involved simple sample preparation without Pharmacy, Charusat Campus, any purification, as well as isocratic separation of Withaferin-A. The method Changa, Petlad, Anan - 388421, was subjected to validation involving establishing linearity, precision, accuracy, Gujarat, India. specificity, and robustness.Optimized mobile phase was acetonitrile: water (45: 55 v/v) contained 1% v/v glacial acetic acid. The column used was Phenomenex E-mail: mananraval.ph@charusat.ac.in $(250 \times 4.6 \text{ mm}, 5 \text{ }\mu\text{m})$ with flow rate set at 1 ml/min and chromatogram was recorded at 216 nm. The studies showed that peak corresponding to Withaferin – A within concentration range of 1-75 µg/ml was found to be correlated with respective concentration with correlation coefficient (r^2) 0.99802 \pm 0.0004. The developed analytical method was found precise, while accuracy was established using standard addition method. The average recovery was 95.59%, when determined at three different spiking levels (50%, 100% & 150%), in the preanalyzed sample solution. LOD was found to be 0.3 µg/ml and LOQ was 0.1 µg/ml. Robustness study suggested that increase in acetonitrile proportion, increased R_T as well as increase in flow rate negatively impacted up on R_T . The content of Withaferin-A was estimated as 0.03 ± 0.001 mg % w/w. The results obtained from the studies showed that method was found to be accurate, precise and validated for estimation of Withaferin-A from Ashwagandhadi lehya.

INTRODUCTION: Ashwagandhadi lehya (AL) is a polyherbal formulation, official in Ayurvedic formulary of India. AL is indicated in the treatment of piles, psychosis, as tonic, rejuvenating agents and in treatment of disorders related to male reproductive system. One of the major ingredients of AL is dried powdered roots of Withania sominfera (WS).

QUICK RESPONSE CODE	DOI: 10.13040/IJPSR.0975-8232.11(1).334-42	
	This article can be accessed online on www.ijpsr.com	
DOI link: http://dx.doi.org/10.13040/IJPSR.0975-8232.11(1).334-42		

It is reported to contain equal proportion, each of powdered roots of *Smilax glabra*, *Hemidesmus indicus*, dried powder of fruits of *Cuminum cyminum* and paste of *Vitis vinifera*. The standards to ensure the quality of AL is mentioned in Ayurvedic Pharmacopoeia of India¹. Amount of Withaferin -A (WA) was estimated from AL using HPTLC.

Physical and chemical standards for AL were also evolved ². Apart from other users, WS is used as aphrodisiac, traditionally, and in treatment of spermatorrhoea as well as to treat seminal debilities ³. WS root powder showed spermatogenic potential when evaluated in albino rats. Aqueous extract of the roots of WS showed marked impairment in libido, sexual performance, sexual vigor and penile erectile dysfunction in rats. The spermatogenic potential of WS was thought to be attributed to increased interstitial cells stimulating hormone and testosterone-like effects ⁴. Controlled clinical studies in oligospermia patients showed that WL root extract significantly improved deteriorated semen parameters, serum testosterone level and inhibited lipid peroxidation as compared to placebo-treated patients ⁵⁻⁷.

The roots of WS were found to contain Withanolides. Withanolides are ergostane type of steroids with atoms C-22 and C-26, bridged by a δ -lactone and an oxidized C-1 position. Withanolides include withanone, WA, withanolides, withasomidienone⁸. IUPAC name of WA is 4 β , 27-dihyroxy-5 β -6-epoxy-1-oxowitha-2, 24-dienolide. Chemical structure of WA is shown in **Fig. 1**. WA isolated from WS was found to possess anti-inflammatory, anti-convulsive, anti-tumor and antioxidant activity ⁹⁻¹¹. WA showed immunosuppressive action on blymphocyte proliferation too ¹².



FIG. 1: CHEMICAL STRUCTURE OF WA

These compounds are specific for Solanaceae family, in particular, for the genus Withania, and thus they were used as marker compounds ¹³. WA was estimated from WS and different extracts there of as well as from herbal formulations contained WS, as one of the ingredients, using TLC and HPLC¹⁴⁻³¹, HPLC-MS/MS³², UHPLC–MS/MS³³ Analytical methods involved HPTLC suffered from poor reproducibility. WA was also estimated using HPLC, from variety of matrices. The quantitative estimation procedure from plant drug or from other herbal formulations often found to possess disadvantages, including, either the acetylation of WA was required prior to analysis ³⁴, or the separation time was long ³⁵ or the compounds were not baseline separated and eluted ³⁶. Our efforts were concentrated particularly on developing an

analytical method for quantitative estimation of WA from a complex matrix of AL using HPLC, which would include relatively simple sample preparation alongside, the developed method would robust, enough sensitive and reproducible.

MATERIALS AND METHODS: AL was manufactured by Sunder Pharmacy (GMP certified manufacturing unit associated with from J.S Ayurveda College, Nadiad, Gujarat, India) on our request (batch number -131016). WA was obtained from Pharmanza Pvt., Ltd., Khambhat, Gujarat as gratis sample (purity 95% w/w) and it was used without any further processing.

Reagents and Chemicals: All HPLC grade solvents and chemicals (acetonitrile, glacial acetic acid) were purchased from Merck India Ltd. Water used for HPLC instrument was obtained from a Millipore water treatment system. Solvents utilized for HPLC were filtered through a 0.45 μ m filter and degassed at room temperature for 20 min using laboratory sonicator (Remi Instruments, Mumbai, India), prior to use. The solvents used for extraction purpose were of analytical grade and procured from Ms. Loba Chemie, India.

Preparation of Standard Stock Solution: 100 mg WA was dissolved in 20 ml methanol using sonication. The solution was diluted up to 100 ml using methanol, to the produced standard stock solution (1000 μ g/ml).

Preparation of Serial Dilutions: Different aliquots of standard stock solution of WA were diluted using methanol to produce different dilutions in the range of $1 \mu g/ml$ to 75 $\mu g/ml$.

Preparation of Sample Solution: 50 g AL was loaded on 100 g silica (Silica for Column Chromatography, 60-120). The mixture was placed in a thimble, prepared from filter paper, and extracted for 2 h at 60 °C, using 200 ml hexane in a Soxhlet extractor. The hexane extract was discarded. Dried marc was extracted using Soxhlet extractor, with 200 ml methanol at 60 °C, for 4 h. The methanol extract was collected, filtered and concentrated using rotary vacuum evaporator at 40 °C. Residual solvent was evaporated from concentrated mass on a water bath. The semisolid, black-brown extract obtained was preserved in desiccator at room temperature.

Optimization of Chromatographic Parameters: Chromatographic separation was achieved using a Waters quadra gradient chromatographic system equipped with PDA detector. Optimized mobile phase was acetonitrile: water [contained 1% v/v glacial acetic acid] (45:55 v/v). The column used was Phenomenex (250 × 4.6 mm, 5 μ m) with flow rate set at 1 ml/min. Injection volume was 20 μ l and chromatogram was recorded at 216 nm. Data was collected, processed and integrated using Waters Empower software. The chromatogram of methanolic extract of AL with peak corresponded to WA is shown in **Fig. 2b**. The peak of WA from standard solution is shown in **Fig. 2a**.

System Suitability Parameters: Repeatability (from an area of peak corresponding to WA standard solution contained 30 μ g/ml WA), capacity factor, number of plates, tailing factor and resolution factor were selected as system suitability parameters (SSP). These values were determined from chromatogram and compared with those recommended ³⁷. The parameters were continuously monitored for each experiment. The value determined for selected parameters is shown in **Table 1**.

Validation of Developed Analytical Method: The developed analytical method was subjected to evolve various parameters by adopting the methodology proposed in ICH guideline ³⁸.

Linearity: Linearity was established by injecting six different dilutions of WA solution covering the concentration range of 1-75 µg/ml (1, 15, 30, 45, 60, 75 μ g/ml) individually, each 20 μ l. The optimized chromatographic parameters were adopted to resolve the peak. Area and R_T of the peak corresponding to WA was noted. The experiment was repeated six times. Area and corresponded concentration of WA were processed mathematically to determine line equation using the least square regression analysis. The correlation coefficient (r^2) was also determined. Result of the studies is shown in Table 2, showing average value of slope, y-intercept, and r^2 with SD.

Precision: Precision of the developed analytical method was determined by performing interday and intraday variation in the peak area of WA for three

selected concentration (15 μ g/ml, 30 μ g/ml, 45 μ g/ml) from linearity experiment. Intraday variation was determined by recording peak area of WA in standard solutions of WA, six times in a day while those were determined for six days, once in a day to determine interday precision. The data set obtained for each concentration was subjected to statistical treatment to determine mean and % RSD. The results of the studies are shown in **Table 3**.

Accuracy: Accuracy studies were performed by spiking WA standard, at three different levels (~ 50%, 100% & 150%), in pre-analyzed sample solution. The spiked sample solution was injected in HPLC system, and amount of WA in spiked sample was determined. Amount of WA recovered was calculated and recorded as % recovery in Table 4.

Selectivity: Selectivity of the analytical method was assessed by comparing the UV spectra of WA received for peak of WA from standard solution with that for WA from sample solution. UV spectra were recorded by scanning the eluents continuously in range of 400 -200 nm. Contour plots were developed for peaks, individually, from UV data and were compared. The contour plots for peak of WA in standard solution as well as in sample solution are shown in **Fig. 3a** and **Fig. 3b**, respectively.

LOD and LOQ: LOD and LOQ of the developed method were determined initially by visual observation by noting signal to noise ratio (S/N), while selecting concentrations for linearity experiments. S/N for LOD was considered as 3, while that for LOQ was considered as 10. Later, LOD and LOQ were determined from line equation, by adopting formula give below:

Limit of Detection (LOD): $3.3 \times \sigma/S$

Limit of Quantification (LOQ): $10 \times \sigma /S$

Where, σ = Standard deviation of the y-intercept, S = Slope of the calibration curve (m). The results of the studies are shown in **Table 5**.

Robustness: Robustness studies were performed by introducing deliberate alteration in selected important chromatographic parameters. The selected parameters were, the proportion of concentration of GAA in water (% GAA) [Factor A], % of acetonitrile in mobile phase [Factor B], mobile phase flow rate [Factor C] and wavelength to record chromatogram [Factor D]. The studies were performed by incorporation of alteration in one of selected chromatographic parameters. The recorded chromatogram was using altered chromatographic conditions. R_T for WA peak (Y1) and area of WA peak (Y2) were recorded for each experiment. Each experiment, for a set of condition, was performed thrice and mean was considered. The datasets were then compared, for statistical significance, with respective datasets obtained, following optimized chromatographic conditions.

Studies Effect on of Chromatographic Parameters on Area and R_T: Box-Behnken design was adopted as experiment design. The coding of the factors is shown in Table 6. The chromatograms recorded for each set of chromatographic conditions and response of dependent variables for a set of selected independent variables was noted. The mean value along with SD for both the dependent variables was determined. The data sets were processed using Design ExpertTM software to generate regression equation and regression coefficient for each dependent parameter. The effect of independent variables on dependent variables was determined by calculating p-value for model as well as for each parameter. $p \leq 0.05$ was selected as level of significance.

Estimation of WA from AL: Calibration range selected for the set of experiments was 1µg/ml to 75µg/ml. Each concentration was injected six times (n=6). The calibration curve was constructed by plotting corresponded peak area versus concentration. Optimized chromatographic parameters were adopted to resolve the peak of WA from other peaks in sample solution. The line equation was constructed using the least square regression analysis. Amount of WA present in AL was calculated from line equation using area of peak corresponded to WA.

RESULTS AND DISCUSSION: AL was intended to be evaluated for action on male reproductive system functions. This analytical method was developed as a part of evolving standards to ensure the quality of AL. AL was reported as a polyherbal formulation contained plants other than WS too. Amongst the plants incorporated in AL, WS was the only plant subjected to biological evaluation for action on male reproductive functions. WA was one of the major constituents of WS, thus selected as a chemical marker to standardize AL concerning the presence of WS. An analytical method based on HPLC was developed which could resolve peak of WA from peaks corresponding to nearby constituents in complex matrix of AL.

AL was semisolid material, which contained fatty substances, interfered in the extraction of AL from matrix. AL loaded on silica and was defatted using hexane, to remove fatty constituents present in formulation. Silica was used to avoid clumping and to increase the surface area. Dried marc was then extracted using methanol, being the solvent capable of dissolving WA. Cold continuous extraction using Soxhlet apparatus was opted to maximize the vield with utilization of comparatively less amount of solvent. The extraction procedure excluded the heating of sample mass; instead distilled solvent was passed through the bed of the marc. Time and temperature for extraction processes were set to get maximum yield of WA, which was ensured by performing co-TLC of AL extract and WA standard solution.

The analytical method was modified from the reported analytical method for estimation of WA from WS root powder using HPLC ²². The reported method could not resolve the peak of WA from nearby peaks in sample solution. Incorporation of GAA in mobile phase made the mobile phase acidic, which could resolve WA from sample solution. The concentration of GAA was set using trial and error.

The final concentration of GAA could resolve the peak of WA from nearby peaks, without altering the gaussian shape of WA peak appeared in chromatogram at R_T less than 10 min. UV spectra of the WA peak were recorded. Maximum absorbance was observed at 216 nm, which was selected as wavelength to record chromatogram using PDA detector. As shown in **Fig. 2a** and **Fig. 2b**, optimized mobile phase yielded well resolved, sharp, guassian peak corresponding to WA, at 7.12 min.



FIG. 2A: CHROMATOGRAM SHOWING PEAK OF WA IN STANDARD SOLUTION

SSP was determined for the peak of WA, obtained after following optimized chromatographic conditions, as shown in **Table 1**.

TABLE 1: SYSTEM SUITABILITY PARAMETERSFOR PEAK CORRESPONDING TO WA (n=6)

S.	Parameter	Value	Recommended
no.		determined	value
1	Reproducibility	0.08%	RSD less than 1 %
	(n=10)		
2	Capacity Factor	7.12	Greater than 2.0
3	Number of Plates	5427.85	Greater than 2000
4	Resolution	3.71	Greater than 2.0
5	Tailing Factor	0.916	Less than 1.5

Repeatability value was found less than 2.0. It ensured precision on the same operating conditions over a short interval of time. Theoretical plates were higher than 2000, ensured the sharp and Gaussian peak. Capacity factor value sufficiently high, to provide enough duration for analyte to interact with stationary phase. Resolution factor greater than 2.0 confirmed that peak of WA was separated properly from nearby peak, and could be used for quantitative purpose.

TABLE 2: LINEARITY DATA FOR WA



FIG. 2B: CHROMATOGRAM SHOWING PEAK OF WA IN METHANOLIC EXTRACT OF AL

The developed analytical method was validated to ensure that the method would perform as per the standards set for the purpose. This could be achieved by showing observational and numerical data gathered and analyzed further, during a series of experiments, suggested for the parameters, as given in ICH guidelines ³⁶.

The linearity of an analytical procedure is the ability of method, within the selected ranges, to obtain test results which are directly proportional to the concentration of the analyte. The linearity of analytical method was established by determining the correlation coefficient for the regression curve, prepared by plotting area of peak corresponded to different concentration of WA. The regression line was constructed by fitting the data in the least square regression model. The mean value of the regression coefficient was found to be 0.9980 ± 0.0004 . Approaching the correlation coefficient value to 1.0, confirmed the linearity of the response in the selected concentration range.

TABLE 2. LINEARITT DATA FOR WA					
Analyte	Slope (m)	Confidence	y-intercept (C)	Confidence	Co-relation Co-efficient
	Mean ± SD	interval*	Mean ± SD	interval*	(\mathbf{r}^2) Mean \pm SD
WA	28053 ± 159.30	27925.53 - 28180.46	6356.70 ± 1392.76	5242.25 - 7471.14	0.99802 ± 0.0004
n=6 *Confidence interval at $n=95$ %					

n=6, *Confidence interval at p = 95 %

Precision studies were performed to expresses the degree of scattering between a series of measurements obtained from multiple sampling of the same homogeneous sample under optimized conditions. Intraday and interday analysis was performed on three different concentrations. The mean of % RSD for intraday was 0.76% and interday was 1.17%.

THE STALL ST
--

WA Concentration	Intra-day variation in peak area of WA	Inter day variation in peak area of WA
	(Intraday) (% RSD)	(% RSD)
15 µg/ml	0.71	0.96
30 µg/ml	0.81	0.97
45 µg/ml	0.73	1.58

Accuracy studies were performed by adopting the standard addition method. WA was spiked at three different levels in pre analyzed sample solution. The amount of WA recovered from spiked sample solution was determined. The result of the studies, as reported in **Table 3**, showed average recovery

TABLE 4: ACCURACY STUDIES (n=3)

was 95.59%. The closeness of recoveries nearer to
100% assured the accuracy of developed analytical
method for the purpose. As, the amount of WA was
low and matrix was complex, recoveries in range of
90 % -110% was acceptable to ensure accuracy.

S. no.	% WA Spiked	Conc. of WA (µg/ml) determined in	% Amount Recovered	% Average Recovery
		Solution Average ± SD	\pm SD	
1	0	1.86 ± 0.01		
2	50	2.78 ± 0.09	96.03 ± 2.89	95.59%
3	100	3.82 ± 0.13	97.05 ± 3.12	
4	150	4.66 ± 0.06	93.70 ± 1.15	

The concentration of WA equivalent to reported value LOD and LOQ was injected in HPLC and chromatogram was recorded. S/N ratio was determined manually, to ensure the results. Finally, 0.3 μ g/ml and 1.0 μ g/ml were accepted as LOD and LOQ for developed method from manual observations and calculating S/N of the recorded chromatogram.

The selectivity of an analytical method is ability of the analytical method to measure accurately analyte in presence of different interfering compounds. It was to be proved for the purpose, that nothing except WA was co-eluted with WA.



FIG. 3A: CONTOUR OF WA IN STANDARD SOLUTION

Selected		led Va	lue
	-1	0	1
Factor A:	0.8	1	1.2
% v/v glacial acetic acid in water			
Factor B:	43	45	47
% v/v of ACN in Mobile Phase			
Factor C:	0.8	1	1.2
Flow Rate of Mobile Phase (ml/min)			
Factor D:	213	216	219
Wavelength selected for recording			
chromatogram			

UV spectra of peak WA were compared and found comparable. Contour was received (generated in range of 400 nm to 200 nm) for the peak of WA obtained in sample solutions as well as in standard solution.

The contours were found to be matched in shapes, as shown in **Fig. 3a** and **Fig. 3b**. These two observations suggested that nothing was co-eluted with WA, confirmed specificity.

 TABLE 5: LOD AND LOQ

LOD	0.3 µg/ml
LOQ	1.0 µg/ml



FIG. 3B: CONTOUR OF WA IN METHANOLIC EXTRACT OF AL

Robustness of the analytical method was ascertained by incorporating deliberate alterations in the optimized chromatographic conditions, and chromatogram was recorded for a selected concentration of WA. The studies showed that, peak area and R_T for WA peak, was not affected statistically as compared to standard, when flow rate was altered by \pm 0.1 ml, % of GAA in water was altered by \pm 0.05% v/v and wavelength was altered by \pm 1 nm, while composition of

acetonitrile was altered by $\pm 1\%$ in mobile phase. It must be noted that only one parameter was altered at a time while performing the studies.

Though, the robustness studies showed the developed analytical method was robust within the limits proposed for the selected factors, an extended set of studies were performed to evaluate relative impact of each selected factor on area and R_T of WA peak. Box - Behnken design, including four independent variables set at three different levels (-1, 0, +1) for their effect on two dependent variables, as shown in Table 6, was evolved and implemented. regression Multiple analysis equations was constituted for both the selected variables.

The regression equation is shown as equation a,

 $R_T (Y1) = (1.52) X_1 + (-0.83) X_2 + (-1.72) X_3 + (-0.012) X_4 + 7.02$ (Analogy to $AX_1 + BX_2 + CX_3 + DX_4$,

Where, A, B, C, D are the numbers associated with factors).

The model was reported to be significant, with regression coefficient 0.0028. The studies also showed that alteration was not significant statistically, in case of dependent variable Y1, for factor D as well as combination of selected factors-AB, BC,CD AD,BC, BD, CD (p>0.05).

The reduced regression equation, as shown in b,

$$R_T(Y2) = (1.52) X_1 + (-0.83) X_2 + (-1.72) X_3 + 7.02.$$

Thus, it was suggested from studies that, wavelength selected to record peak of WA, did not affect R_T while and increase in acetonitrile proportion and flow rate decreased R_T as well as increase in % glacial acetic acid increased R_T of WA peak.

Similarly, the regression equation for second dependant variable, area of WA peak was constructed, shown as equation B.

Area (Y₂) = $(7441.83)X_1 + (-3683.67)X_2 + (-3805.50)X_3 + (-14334.83)X_4$ (Analogy to A X₁ + BX₂ + CX₃ +DX₄

Where, A, B, C, D are the numbers associated with factors). The model was found significant with regression coefficient 0.7770. The results showed that effect of factor D, combined effect of factor

AC and BC was statistically significant on dependant variable Y_2 . The reduced regression equation, as shown in c, was

 $\begin{array}{l} Y_2 = (\text{-}14334.83) \ X_4 + (\text{-} \ 1.86 \times 10^5) \ X_1 X_3 + \ 1.801 \times 10^5 \\ X_2 X_4. \end{array}$

The mathematical treatment, thus, showed that effect of alteration in % GAA, mobile phase ratio or flow rate did not significantly affect the area of WA; though wavelength affected area only. The combined effect of % GAA and flow rate as well as of acetonitrile proportion and wavelength might affect the area of WA peak, in a statistically significant manner. It was observed in robustness studies that, the range for selected chromatographic parameters, were narrow; to get statistically unaltered mean value for area of WA peak, as compared to optimized chromatographic system. The above mathematical treatment yielded the relative impact of alterations in selected chromatographic parameters on area and R_T of WA.

Methanolic extract of AL was used to estimate WA in AL. This sample solution was chromatographed using optimized chromatographic conditions. Area of the guassian peak (y) obtained at R_T value in the chromatogram of sample solution, corresponded to WA peak received for standard solution, was noted. The concentration of WA in sample solution (x) was determined using regression equation obtained for linearity. The experiment performed thrice. The amount of WA present in AL was found to be 0.037 ± 0.001 mg % W/W.

CONCLUSION: An analytical method using HPLC was developed for estimation of WA from AL. The amount of WA was estimated to be $0.03 \pm 0.001 \text{ mg } \%$ w/w in sample of AL taken. The optimized chromatographic parameters might serve as a tool to record chromatographic fingerprint for AL. The developed analytical method included simple sample preparation. The resolution was achieved without gradient set in proportions of mobile phase as well as without any pre chromatographic treatment using PDA detector.

Results from validation studies confirmed that, developed method had linear response in selected concentration ranges. The method was found to be precise and accurate too. The robustness studies suggested that, an increase in acetonitrile proportion, increased R_T and an increase in flow rate negatively impacted up on R_T . Area of WA peak was not statistically affected by alteration in wavelength selected or by altering concentration of glacial acetic acid in mobile phase.

ACKNOWLEDGEMENT: The authors are thankful to CHARUSAT for providing instrument facility, J. S. Ayurveda College, Nadiad, Gujarat for manufacturing AL on our request, WA was obtained from Pharmanza Pvt., Ltd., Khambhat, for providing WA as a gratis sample.

CONFLICTS OF INTEREST: Nil

REFERENCES:

- Anonymous: The Ayurvedic pharmacopeia of India, Part-II, vol-III, first edition, Government of India, Ministry of Health and Family Welfare, Dept of AYUSH 2010; 27-28.
- Gupta M, Bisht D, Pandey MM, Ojha SK, Khatoon S, RastogiSubha and Rawat AKS: Standardization of *Ashwagandhadi lehya* - an important Ayurvedic Formulation of Withania somnifera. Ind J of TradKnowl 2011; 10(4): 594-98.
- 3. Nadakrni KM: Indian material medica, with Ayurveidc, Unani-Tibbi, Siddha, Allopatathic, Homeopathic, Naturopathic and home remedies, Appendix and Indexes, vol-I, Bombay Popular Prakashan, reprint 2009.
- Ilayperuma I, Ratnasooriya WD and Weerasooriya TR: Effect of Withania somnifera root extract on the sexual behaviour of male rats. Asian J Androl 2002; 4(4): 295-98.
- Ambiye VR, Langade D, Dongre S, Aptikar P, Kulkarni M and Dongre A: Clinical evaluation of the spermatogenic activity of the root extract of Ashwagandha (*Withania somnifera*) in oligospermic males: a pilot study. Evid Based Complement Alternat Med 2013; 2013: 571420.
- 6. Ahmad MK, Mahdi AA,Shukla KK, Islam N, Rajender S and Madhukar D: *Withania somnifera* improves semen quality by regulating reproductive hormone levels and oxidative stress in seminal plasma of infertile males. FertilSteril 2010; 94(3): 989-96.
- Mahdi AA, Shukla KK, Ahmad MK, Rajender S, Shankhwar SN, Singh V and Dalela D: Withania somnifera improves semen quality in stress-related male fertility. Evid Based Complement Alt Med 2009 Sep 29.
- Anonymous: Indian Herbal Pharmacopoeia, A joint publication of Regional Research Laboratory, Jammu Tawi and Indian Drug Manufacturers Association, Mumbai, Vol- I, 1998; 165.
- Al-Hindwi MK, Khafagi A and Nabi A: Anti-granuloma activity of Iraqi *Withania somnifera*. J Ethnopharmacol 1992; 37(2): 113-16.
- Kulkarni SK, George B and Mathur R: Protective effect of Withania somnifera root extract on electrographic activity in a lithium-pilocarpine model of status epilepticus. Phytother Res 1998; 12: 451-53.
- 11. Bhattacharya SK, Satyam KS and Ghosal S: Antioxidant activity of glycowithanolides from *Withania somnifera*. Indian J Exp Bio 1997; 35: 236-39.
- 12. Jayaprakasam B and Nair MG: Cyclooxygenase-2 enzyme inhibitory withanolides from *Withania somnifera* leaves. Tetrahedron 2003; 59: 841-49.

- Bruneton J: InPharmacognosy Phytochemistry Medicinal plants, Lavoisier, Paris 1999; 1062.
- 14. Gupta AP, Verma RK, Misra HO and Gupta MM: Quantitative determination of withaferin A in different plant parts of *Withania somnifera* by TLC densitometry. J Med Arom Plant Sci 1996; 18: 788-90.
- Dalavayi S, Kulkarni SM, Itikala RL and Itikala S: Determination of Withaferin-A in two Withania Species by RP-HPLC method. Indian J Pharm Sci 2006; 68(2): 253-56.
- 16. Srivastava P, Tiwari N, Yadav AK, Kumar V, Shanker K, Verma RK, Gupta MM, Gupta AK and Khanuja SP: Simultaneous quantification of withanolides in *Withania somnifera* by a validated HPTLC method. J AOAC Int 2008; 91(5): 1154-61.
- 17. Siriwardane A, Dharmadasa R and Samarasinghe K: Distribution of Withaferin A in different parts of two varieties of *Withania sominfera*. Journal of Biological Sciences 2013; 141-44.
- 18. Rajasekar S and Elango R: Estimation of alkaloid content of Ashwagandha (*Withania somnifera*) with HPLC methods. J of Exp Sci 2011; 2(5): 39-41.
- Agrawal P, Vegda R and Laddha K: Simultaneous Estimation of Withaferin A and Z-Guggulsterone in Marketed Formulation by RP-HPLC. J of chromsci 2015; 53(6): 940-44.
- 20. Sharma V, Gupta AP, Bhandari P, Gupta RC and Singh B: Validated and densitometric HPTLC method for the quantification of Withaferin-A and Withanolide-A in different plant parts of two morphotypes of *Withania somnifera*. Chromatographia 2007; 66(9-10): 801-4.
- Sharma V: HPLC-ELSD method for quantification of Withaferin-A and Withanolide- A in diploid (n=12) and tetraploid (n=24) cytotypes of 'Indian Ginseng' Withania somnifera (L.) Dunal from North India. Int J Indigenous Med Plants. 2013; 205 (1): 1245-50.
- 22. Jirge SS, Tatke PA and Gabhe SY: Development and validation of a novel HPTLC method for simultaneous estimation of beta sitosterol d glucoside and withaferin A. Int J Pharm Pharm Sci 2011; 2(3): 227-30.
- 23. Bhondave PD, Potawale SE, Gabhe SY and Harsulkar AM: A rapid and simple high-performance thin layer chromatographic method for simultaneous analysis of β -sitosterol-D-glucoside, gallic acid, withaferin A and withanolide- A in *Ashvagandha rishta*. J of Appl Pharma Sci 2011; 4(7): 82-87.
- 24. Patel JB, Lahiri SK and Shah MB: Development of a new method for identification and estimation of *Withania somnifera* root, and a method for quantitative analysis of Withaferin A in young and old roots. J of Planar Chrom 2009; 22(4): 283-86.
- 25. Patel BH, Patel JB and Patel RK: Novel validated HPTLC method for estimation of Withaferin A in polyherbal formulations. Anall Methods 2014; 6(9): 3009.
- 26. Gautam V and Kalia AN: High-performance thin layer chromatography method for simultaneous estimation of vicine, trigonelline and withaferin-A in a polyherbal antidiabetic formulation. Int J Pharm Pharm Sci 2013; 5(2): 367-71.
- 27. Rathod B and Kilambi P: Development of validated RP-HPLC method for the estimation of Withaferin –A in *Withania somnifera*, its extract and polyherbal formulation. Int J of Pharmasci 2013; 4(3): 1-9.
- 28. Manwar JV, Mahadik KR, Paradkar AR, Takle SP, Sathiyanarayanan L and Patil SV: Determination of withanolides from the roots and herbal formulation of

Withania somnifera by HPLC using DAD and ELSD detector. Der Pharmacia Sinica 2012; 3(1): 41-46.

- 29. Mirjalili1 MH, Navarro A, Hernández L, Jáuregui O and Bonfill M: LC–MS/MS method for the quantification of Withaferin-A in plant extracts of Withania spp. Acta Chromatographica 2013; 25(4): 745-54.
- Anonymous: Quality standards of Indian Medicinal Plant, Vol 9, Indian Council For Medical Research, New Delhi 2011a; 356-67.
- Anonymous: The Ayurvedic Pharmacopoeia of India, Part-I, Vol-III, First edition (Formulations), Government of India, Ministry of Health and Family Welfare, Department of AYUSH 2011b; 28-32.
- 32. Patil D, Gautam M, Mishra S, Karupothula S, Jadhav S, Pawar S, Patwardhan B and Gairola S: Determination of Withaferin A and Withanolide A in mice plasma using high-performance liquid chromatography-tandem mass spectrometry: Application to pharmacokinetics after oral administration of *Withania somnifera* aqueous extract. J of Pharma and Biomed Anal 2013; 80: 203-12.
- 33. Chandra P, Kannujia R, Saxena A, Srivastava M, Pal M, Singh B, Ojha S, Kumar B and Bahadur L: Quantitative determination of multi markers in five varieties of *Withania somnifera* using ultra-high-performance liquid

chromatography with hybrid triple quadrupole linear ion trap mass spectrometer combined with multivariate analysis: Application to pharmaceutical dosage forms. J of Pharma and Biomed Anal 2016; 129: 419-26.

- 34. Bessalle R and Lavie D: Semi-quantitative reversed-phase high-performance liquid chromatographic analysis of the ecotypes of *Withania somnifera* chemotype III. J of Chrom 1987; 389: 195-10.
- Hunter IR, Walden MK, Heftmann E, Glotter E and Kirson I: Separation of Withanolides by high-pressure liquid chromatography with coiled columns. J Chromatogr 1979; 170(2): 437-42
- Banerjee S, Naqvi AA, Mandal S and Ahuja PS: Transformation of *Withania somnifera* (L) Dunal by *Agrobacterium rhizogenes*: infectivity and phytochemical studies. Phytother Res 1994; 8(8): 452-55.
- 37. Anonymous: CDER, Guidance for Industry Analytical Procedures and Methods Validation for Drugs and Biologics Guidance for Industry Analytical Procedures and Methods Validation for Drugs and Biologics. U.S. Department of Health and Human Services and Food and Drug Administration 2014.
- 38. Anonymous: Guideline ICH, Validation of analytical procedures: text and methodology, Q2 (R1), Nov 2005.

How to cite this article:

Manan AR, Kanan GG, Niraj YV and Nishit DP: Estimation of Withaferin – a from *Ashwagandhadi lehya* using HPLC. Int J Pharm Sci & Res 2020; 11(1): 334-42. doi: 10.13040/JJPSR.0975-8232.11(1).334-42.

All © 2013 are reserved by International Journal of Pharmaceutical Sciences and Research. This Journal licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported License.

This article can be downloaded to Android OS based mobile. Scan QR Code using Code/Bar Scanner from your mobile. (Scanners are available on Google Playstore)