IJPSR (2020), Volume 11, Issue 4



INTERNATIONAL JOURNAL



Received on 31 January 2020; received in revised form, 18 March 2020; accepted, 20 March 2020; published 01 April 2020

EVALUATION OF ANTIGENOTOXIC POTENTIAL OF BEET ROOT EXTRACT AGAINST HAIR DYE INDUCED GENOTOXICITY

Sunil Kumar¹, Veena Vishwakarma¹, Bharti Yadav¹, Ranjan Gupta², Neeraj K. Aggarwal³ and Anita Yadav^{*1}

Department of Biotechnology¹, Department of Biochemistry², Department of Microbiology³, Kurukshetra University, Kurukshetra - 136119, Haryana, India.

Keywords:

p-phenylenediamine (PPD), Hair dye, Genotoxicity, Beet root extract (BRE), Peripheral blood lymphocytes (PBLs)

Correspondence to Author: Dr. Anita Yadav

Professor, Department of Biotechnology, Kurukshetra University, Kurukshetra - 136119, Haryana, India.

E-mail: ayadav@kuk.ac.in

ABSTRACT: *In-vitro* investigations toward DNA damage reported that the biological effect of genotoxic exposures could influence the life cycles of different human cells. Sister chromatid exchange (SCE) assay is broadly used, sensitive cytogenetic biomarker of exposure to genotoxic agents. Naturally available products may play a significant role against the cytogenetic damage but available research data is not consistent. The purpose of this study was to evaluate the anti-genotoxic potential of beet root extract (BRE) against hair dye induced genotoxicity in cultured human peripheral blood lymphocytes (PBLs) using SCE assay. For this, PBLs were treated with different doses of hair dye ingredients along with BRE. Significant (p<0.05) rise in SCE frequency indicates genotoxicity of hair dye ingredients (p-Phenylenediamine-200 µg/ml, resorcinol-200 µg/ml, and hydrogen peroxide-6.168 µg/ml). Treatment of BRE along with hair dye showed significant (p<0.05) reduction in SCE frequency in a dose-dependent manner as compared to hair dye treated samples.

INTRODUCTION: P-phenylenediamine (PPD) is an essential ingredient of oxidative hair dyes. PPD and its metabolites are allergenic, mutagenic, and highly toxic. Some body organs like kidneys and bladder are particularly vulnerable to toxic effects of PPD¹. Various epidemiological investigations have indicated increase incidences of malignancy like bladder cancer due to the dermal application of PPD containing oxidative hair dyes^{2, 3}. Many researches also show carcinogenic, mutagenic, and genotoxic effects of PPD using different assay systems^{4, 5}.

QUICK RESPONSE CODE	
	DOI: 10.13040/IJPSR.0975-8232.11(4).1874-78
	The article can be accessed online on www.ijpsr.com
DOI link: http://dx.doi.org/10.13040/IJPSR.0975-8232.11(4).1874-78	

Another study in human uroepithelial cells indicates the genotoxic potential of PPD due to induced mutation in p53 and upregulation of COX-2 gene ⁶. Beet root extract is a good source of bioactive components like flavonoids, polyphenols, betalains, and others, which decrease oxidative stress and improve antioxidant status in humans ⁷. They have significant anti-mutagenic or anti-genotoxic effects against Methylnitro-nitrosoguanidine (MNNG) induced mutations ⁸.

Sister chromatid exchange (SCE) involve breakage and exchange of genetic material between two sister chromatid at the homologous position during metaphase. These can be induced by different types of DNA-damaging agents. SCE has been accepted as a good biomarker for DNA damage ⁹. In present study, potential protective effects of beet root extract against hair dye induced genotoxicity have been investigated in cultured human peripheral blood lymphocytes.

MATERIALS AND METHODS:

Preparation of Solution: P-Phenylenediamine (Sigma-Aldrich, CAS: 106-50-3) and resorcinol (Sigma-Aldrich, CAS: 108-46-3) and 3% H₂O₂ were prepared in autoclaved double-distilled water. Beet root extract (BRE) (Sigma-Aldrich, CDS000584) was also prepared in double-distilled water. Sterilization of all stock solutions (1mg/ml) was done through 0.22-micron syringe filters. Before storage at -20 °C, all stock solutions were appropriately covered to protect from light. Hair dye was prepared by using an equal volume ratio of PPD, resorcinol and 3% H₂O₂.

Blood Sampling: For lymphocytes culture, about 5 ml of venous blood was collected in heparin (sodium) coated sterile vacutainers from healthy individuals. Consent was taken from each individual along with a questionnaire regarding their health history. Ethical approval for this study has been taken from the Institutional Human Ethical Committee of Kurukshetra University (IHEC/17/422).

Culture Setup: The cultures of human peripheral blood lymphocytes (PBLs) were set by slightly modifying the method of Moorhead et al. 10 Briefly, 500 µl of peripheral blood was added in 5 ml of RPMI-1640 medium containing L-glutamine (1%), fetal bovine serum (20%), penicillin (100 U/ml), streptomycin (100)µg/ml) and phytohemagglutinin (2%) (HiMedia). Different concentrations of hair dye ingredients PPD + RE + H_2O_2 (33.33 µg/ml + 33.33 µg/ml + 1.028 µg/ml - $200 \ \mu g/ml + 200 \ \mu g/ml + 6.17 \ \mu g/ml)$ were added to check their genotoxicity by SCE assay. The antigenotoxic potential of beet root extract was assessed by pre-treating the above culture containing different ingredients of hair dye with different doses of beet root extract (2-12 µg/ml). All cultures were incubated in a CO₂ incubator (5%) at 37 °C. Beet root extract was also checked for its possible genotoxic effect if any. Ethyl methanesulfonate was used as the positive control.

Sister Chromatid Exchange (SCE) Assay: For SCE analysis, the culture of PBLs was set up as stated above for 72 h at 37 °C using the

methodology of Perry & Wolff¹¹. After 24 h of incubation, 10 µg/ml of 5-bromo-2-deoxyuridine (Sigma-Aldrich) was added in culture and again transferred to the incubator for another 48 h at the same conditions. Two-three drops of colchicines were added in a final concentration of 0.2 µg/ml about 45 min before harvesting the cells. After centrifugation harvesting, hypotonic solution (0.075 M KCl) treatment was given and then fixed in methanol: acetic acid (3:1). Using these fixed cell suspension slides were prepared by putting cells form a height followed by air drying. Subsequently, slides were stained with Hoechst 33258 and 4% Giemsa stain. SCE frequency per cell was calculated by analyzing fifty metaphases.

Statistical Analysis: SPSS software was used for carrying out statistical analysis. The values correspond to $P \le 0.05$ were considered to be statistically significant. All experiments were performed in duplicates. ANOVA (one way) was used for multiple samples, and Student's t-test was used for comparing paired sample tests, and final outcomes were described as Mean \pm SD.

RESULTS: In this current study, the antigenotoxic effects of beet root extract against hair dye induced genotoxicity were studied using sister chromatid exchange (SCE), a classic cytomolecular method which has been considered as an easy and accurate method to monitor genetic damage in DNA.



FIG. 1: EFFECT OF DIFFERENT TREATMENTS OF HAIR DYE ON DNA DAMAGE IN PBLS AS EVALUATED BY USING SISTER CHROMATID EXCHANGE (SCE) ASSAY. *p<0.05 (significant as compared to untreated sample). c-control; ems-ethyl methanesulfonate; hd- hair dye (PPD + RE + H₂O₂)

A significant increase in the SCE frequency was observed in dose-dependent manner when cultures were treated with different concentration of hair dye ranging from PPD $(33.33 \ \mu g/ml) + RE (33.33 \ \mu s/ml)$ $\mu g/ml$) + H₂O₂ (1.028 $\mu g/ml$) to PPD (200 $\mu g/ml$) + RE $(200 \ \mu g/ml) + H_2O_2$ (6.17 $\mu g/ml)$. Significant higher SCE frequency (8.66 \pm 0.76) was observed at a concentration of PPD (200 µg/ml) + RE (200 $\mu g/ml$) + H₂O₂ (6.17 $\mu g/ml$) treated culture as compared to control (2 ± 0.11) indicating increased genotoxicity in the presence of hair dye Fig. 1 and **3**. Beet root extract showed anti-genotoxic potential against hair dye induced genotoxicity in a dosedependent manner as evident by a decrease in SCE frequency. The maximum decrease in SCE frequency (P ≤ 0.05) (3.676 \pm 0.37) was observed at a concentration of 12 µg/ml of beet root extract Fig. 2 and 3. A separate culture along with beet root extract only (12 µg/ml) showed to genotoxic effects in PBLs as evident by no significant difference in SCE frequency (2.332 ± 0.27) as compared to control sample.



FIG. 2: EFFECT OF DIFFERENT TREATMENTS OF BRE ON DNA DAMAGE IN PBLS WAS EVALUATED BY USING SISTER CHROMATID EXCHANGE (SCE) ASSAY. *p<0.05 (significant as compared to hair dye alone treated sample). c-control; hd- hair dye (PPD + RE + H₂O₂); bre-beet root extract



FIG. 3: SISTER CHROMATID EXCHANGE (SCES) IN PBLS (A) CONTROL; (B) HAIR DYE TREATED (C) HAIR DYE ALONG WITH BRE TREATED (ARROW SHOWS SISTER CHROMATID EXCHANGES)

DISCUSSION: In our study, it has been observed that an increase in BRE concentration leads to a significant decrease in SCE frequency. Due to some novel findings, beet root extract catches the attention of researchers towards its health benefits.

Beet root extract exhibit powerful free radical scavenging property and act as an antioxidant that helps in oxidative stress-related disease, including cancer ^{12, 13}. In this regard, the anti-tumour activity of beet root extract was demonstrated in laboratory

animal ^{14, 15}. Short and prolonged treatment of beet root extract using a mouse model has shown a delay in the NDEA-induced tumor ¹⁶. This delay in tumor onset could involve the protection against DNA damage caused by NDEA-derived electrophiles or ROS.

Beet roots are a good source of phytonutrients called betalains. Two best-studied betalains are betanin and vulgaxanthin, which exhibit free radical scavenging antioxidant activity, antiinflammatory, and detoxification potential ¹⁷. These natural compounds present in the beet root extract may interact directly with the genotoxicant and inhibit them chemically. Possibly, these compounds protect DNA from damage by altering the binding of mutagen by competitively interacting with the nucleophilic sites in DNA. But, mutagenesis inhibition can act through different complex molecular mechanisms ¹⁸.

Esatbeyoglu *et al.*, found that beet root extract reduced DNA damage in cultured human liver cells (hepatoma) induced by H_2O_2 ¹⁹. It also considerably inactivated pathway for the production of ROS in cultured human neutrophils, and reduced DNA damage ^{20, 21, 22}.

Chyau *et al.*, evaluated the ability of betanin (a component of beet root extract) to cause any cytotoxicity effect in cell survival and found no cytotoxicity in the range from 10-100 μ M²³.

Lee *et al.*, reported that pre-treatment of beet root extract significantly decreased MDA level and increased GSH and SOD level against ethanol mediated toxicity in rats ²⁴. Kapadia *et al.*, studied the effect of beet root extract on multiorgan tumors in experimental animals and found it significantly effective in suppression of tumors ¹².

CONCLUSION: Our results concluded that PPD, a component of oxidative hair dyes, can cause genetic damage in peripheral blood lymphocytes. Beet root extract may be used as natural dietary supplement against genotoxicity of hair dye.

ACKNOWLEDGEMENT: We are grateful to all the blood donors for their contribution to this study.

CONFLICTS OF INTEREST: The authors declare that there is no conflict of interest.

REFERENCES:

- Soni S, Nagarik A, Dinaker M, Adikey G and Raman A: Systemic toxicity of paraphenylenediamine. Indian Journal of Medical Sciences 2009; 63(4): 164-6.
- 2. Rauscher GH, Shore D and Sandler DP: Hair dye use and risk of adult acute leukemia. American journal of epidemiology 2004; 160(1): 19-25.
- 3. Malvestio A, Bovenzi M, Hoteit M, Belloni FA, Peserico A, Teresa CM and Filon LF: p-Phenylenediamine sensitization and occupation. Contact Dermatitis. 2011; 64(1): 37-42.
- 4. Van BD: Carcinogenicity of hair dye components. Journal of environmental pathology and toxicology 1980; 3(4 Spec No): 237-51.
- 5. Shahin MM: Evaluation of the mutagenicity of azo dyes in Salmonella typhimurium: a study of structure-activity relationships. Mutagenesis 1989; 4(2): 115-25.
- Huang YC, Hung WC, Kang WY, Chen WT and Chai CY: p-Phenylenediamine induced DNA damage in SV-40 immortalized human uroepithelial cells and expression of mutant p53 and COX-2 proteins. Toxicology letters 2007; 170(2): 116-23.
- Mekki LL, Mansour H, Eldean EG and Nasser AA: Evaluation of anti-genotoxicity of the ethanolic plant extract of Beta vulgaris Maritima using *Allium cepa* root assay. Egypt J Exp Biol 2015; 11: 147-53.
- Lechner JF and Stoner GD: Red beetroot and betalains as cancer chemopreventative agents. Molecules 2019; 24(8): 1602.
- 9. Perry PE: Chemical mutagens and sister-chromatid exchange. In Chemical mutagens. Springer, Boston, MA 1980; 1-39.
- 10. Moorhead PS, Nowell PC, Mellman WJ, Battips DT and Hungerford DA: Chromosome preparations of leukocytes cultured from human peripheral blood. Experimental cell research 1960; 20(3): 613-6.
- Perry P and Wolff S: New Giemsa method for the differential staining of sister chromatids. Nature. 1974; 251(5471): 156-8.
- 12. Kapadia GJ and Rao GS: Anticancer effects of red beet pigments. In Red Beet Biotechnology. Springer, Boston, MA 2013; 125-54.
- 13. Kanner J, Harel S and Granit R: Betalains a new class of dietary cationized antioxidants. Journal of Agricultural and Food Chemistry 2001; 49(11): 5178-85.
- 14. Kapadia GI, Tokuda H, Konoshima T and Nishjnod H: Chemoprevention of lung and skin cancer by *Beta vulgaris* (beet) root extract. Cancer Letters 1996; 100: 211-14.
- Lechner JF, Wang LS, Rocha CM, Larue B, Henry C, McIntyre CM, Riedl KM, Schwartz SJ and Stoner GD: Drinking water with red beetroot food color antagonizes esophageal carcinogenesis in N-nitroso methylbenzylamine-treated rats. Journal of Medicinal Food 2010; 13(3): 733-9.
- 16. Kapadia GJ, Azuine MA, Sridhar R, Okuda Y, Tsuruta A, Ichiishi E, Mukainake T, Takasaki M, Konoshima T, Nishino H and Tokuda H: Chemoprevention of DMBAinduced UV-B promoted, NOR-1-induced TPA promoted skin carcinogenesis and DEN-induced phenobarbital promoted liver tumors in mice by extract of beetroot. Pharmacological Research 2003; 47(2): 141-8.
- Čanadanović-Brunet JM, Savatović SS, Ćetković GS, Vulić JJ, Djilas SM, Markov SL and Cvetković DD: Antioxidant and antimicrobial activities of beet root pomace extracts. Czech Journal of Food Sciences 2011; 29(6): 575-85.

- Aqil F, Zahin M and Ahmad I: Antimutagenic activity of methanolic extracts of four ayurvedic medicinal plants. Indian Journal of Experimental Biology 2008; 46(9): 668-72.
- 19. Esatbeyoglu T, Wagner AE, Schini- Kerth VB and Betanin RG: A food colorant with biological activity. Mole Nutrition and Food Research 2015; 59(1): 36-47.
- 20. Clifford T, Constantinou CM, Keane KM, West DJ, Howatson G and Stevenson EJ: The plasma bioavailability of nitrate and betanin from Beta vulgaris rubra in humans. European Journal of Nutrition 2017; 56(3): 1245-54.
- 21. Esatbeyoglu T, Wagner AE, Motafakkerazad R, Nakajima Y, Matsugo S and Rimbach G: Free radical scavenging and antioxidant activity of betanin: electron spin resonance spectroscopy studies and studies in cultured cells. Food and Chemical Toxicology 2014; 73: 119-26.
- 22. Zielińska-Przyjemska M, Olejnik A, Kostrzewa A, Łuczak M, Jagodziński PP and Baer-Dubowska W: The beetroot component betanin modulates ROS production, DNA damage and apoptosis in human polymorphonuclear neutrophils. Phytotherapy Research 2012; 26(6): 845-52.
- Chyau CC, Chu CC, Chen SY and Duh PD: The inhibitory effects of djulis (*Chenopodium formosanum*) and its bioactive compounds on adipogenesis in 3T3-L1 adipocytes. Molecules 2018; 23(7): 1780.
- 24. Lee JH, Son CW, Kim MY, Kim MH, Kim HR, Kwak ES, Kim S and Kim MR: Red beet (*Beta vulgaris* L.) leaf supplementation improves antioxidant status in C57BL/6J mice fed high fat high cholesterol diet. Nutrition Research and Practice 2009; 3(2): 114-21.

How to cite this article:

Kumar S, Vishwakarma V, Yadav B, Gupta R, Aggarwal NK and Yadav A: Evaluation of antigenotoxic potential of beet root extract against hair dye induced genotoxicity. Int J Pharm Sci & Res 2020; 11(4): 1874-78. doi: 10.13040/JJPSR.0975-8232.11(4).1874-78.

All © 2013 are reserved by the International Journal of Pharmaceutical Sciences and Research. This Journal licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported License.

This article can be downloaded to Android OS based mobile. Scan QR Code using Code/Bar Scanner from your mobile. (Scanners are available on Google Playstore)