IJPSR (2021), Volume 12, Issue 1







Received on 20 January 2020; received in revised form, 14 April 2020; accepted, 22 April 2020; published 01 January 2021

VALIDATING THE PROPERTIES OF *NYMPHAEA PUBESCENS* WILLD. BY PERFORMING UV-VIS, TLC, FTIR AND HPLC ANALYSES

Madhulika ^{*1}, Sanjay Kumar ², K. N. Dwivedi ² and Sangeeta Gehlot ¹

Department of Kriya Sharir¹, IMS, BHU, Varanasi - 221005, Uttar Pradesh, India. Department of Dravyaguna², IMS, BHU, Varanasi - 221005, Uttar Pradesh, India.

Keywords:

Pharmacognostic studies, Water lily, Thin layer chromatography, Organoleptic property, Kumuda

Correspondence to Author: Madhulika

Ph.D. Scholar, Department of Kriya Sharir, Faculty of Ayurveda, Institute of Medical Science, Banaras Hindu University, Varanasi - 221005, Uttar Pradesh, India.

E-mail: madhulikaindia8@gmail.com

ABSTRACT: Introduction: Nymphaea pubescens Willd. is one of the wellknown medicinal plants, being used from ancient times in India and commonly called "Kumuda". It is an aquatic perennial herb. It has leaves having a smooth upper side, a strong root, and a branched rhizome. According to Bentham and Hooker classification "Genera Plantarum", this plant belongs to the division Polypetale of Dicotyledones, which has been divided into 15 order and 84 families. Nymphaea comes under the Araceae family, which itself contains 8 genera and 100 species. Method: Morphological characters have been examined through a transverse section cut by microtome. Physiochemical studies and other screenings were performed accordingly. In addition to this, UV-VIS, TLC, FTIR, and HPLC tests were also carried out for further validation. Result: The morphological and physiochemical examination revealed the presence of saponins, proteins, and various other tests in Nymphaea pubescens Willd. Conclusion: The microscopic, UV-VIS, TLC, FTIR, and HPLC validation can be helpful for using it for medicinal and commercial purposes.

INTRODUCTION: *Nymphaea pubescens* Willd. is also known as water lily; it is grown in ponds, lakes, and water bodies, having simple heartshaped leaves, branched rhizome, striated seed, and globular fruit ¹. The whole plant is used as a medicine in Type 2 diabetes mellitus and urinary tract disease ². Although it is one of the types of lotus species, a very brief illustration is found in different botanical and Ayurvedic treatises. The scarcity of information regarding the characteristics and its uses motivates the author to review it extensively in various research journals and other related literature.



And to standardized the parameter after performing Organoleptic properties, powder microscopy, physiochemical, UV-VIS, TLC, FTIR, and HPLC studies. Such exploration and validation help society in the treatment of various diseases and for medicinal and commercial purposes.

MATERIALS AND METHODS:

Collection: *Nymphaea pubescens* Willd. was collected from its natural habitat near Banaras Hindu University, Varanasi, in July 2018.

Authentication of Plants: Sample (Voucher specimen no. Nymphaea.2018/1) was authenticated by the expert from the Department of Botany, Institute of Science, Banaras Hindu University, Varanasi. Plant specimen was deposited in the museum of the Department of Dravyaguna, Faculty of Ayurveda, for future reference.

Chemicals: All analytical grade chemicals used in the study were purchased through Advanced Quality traders, E. Merk, Germany. **Organoleptic Evaluation:** Organoleptic evaluation of the plant sample was done with the help of sensory organs 3 .

Powder Microscopy: The coarse powder of the sample was boiled with chloral hydrate and collected in a Petri dish, and stained with the different reagent 4 .

Physiochemical Study: Coarse powder of the sample was used for an analytical test which is shown in table number ³.

Loss on Drying: About 10 gm of the sample was placed in a tarred evaporating dish, and the sample was dried at 105 °C for 5 h and weighed. After that, it was cooled in desiccators for 30 min and weighed.

Determination of Total Ash: About 2.0 g of the sample was incinerated in a tarred silica dish at a temperature not exceeding 450 °C until free carbon was left, cooled, and finally weighed.

Acid Insoluble Ash: Ash was dissolved in 25 ml of dilute HCL and boiled for 5 min; then, insoluble matter was collected on an ash-less filter paper, washed with hot water, and ignited to constant weight.

Water Soluble Ash: The sample was dissolved in 25 ml of diluted HCl and made up of 50 ml with water and boiled. Then filtered with Whatmann filter paper and residue was collected.

It was weighed and maintained in a muffle furnace for 6 h at 450-650 °C. The crucible was taken out and cooled at room temperature and weighed.

UV-Vis (Ultraviolet-Visible Spectroscopy): 1 gram of plant extracts added in 10 ml of distill water then filtered with the help of cartilage (0.2 μ m) afterward, it was scanned under ultraviolet, visible spectrophotometer (Perkin Elmer) at a range of 200-900 nm to measuring the size of biomolecules and uncertainty source that may arise from nature of the compound of plant extract.

TLC (**Thin Layer Chromatography**): Thin layer chromatography was detected by observation of spots for identical R_f values and to determine the purity of a sample.

Stationary Phase: Pre-coated plate with silica gel G (Merck) was used as the stationary phase.

Solvent System: Toluene: Ethyl acetate: Formic acid: Methanol was used as a solvent system in the respective ratio of 4.3: 4.3: 1.2: 0.3.

Spraying Reagent: Methanol: sulphuric acid (9:1) was used for drying plates in a hot air oven.

Procedure: Extract was applied 2 cm on the lower edge of the plate with the help of a microcapillary tube. And then extracts were loaded in a small volume spot on each plate, the plate was taken out, the solvent front was marked, and the plate was dried at room temperature.

FTIR (Fourier- Transform Infrared Spectroscopy): A pinch of powder drug was taken and placed over the crystal present on stage. The IR spectrum (Perkin Elmer, Spectrum-2) was scanned between 4000 to 400-1, and transmittance was recorded.

Before scanning the sample, the background signal was also recorded. The peaks thus obtained were matched against the IR interpretation chart, and the functional groups were noted.

HPLC (High-Performance Liquid Chromatography): 1 g of plant extract is added in 10 ml of methanol then sonicator in the sonicator machine (Labman), afterward filtered with the help of cartilage ($0.2 \mu m$) and inject with micro syringe ($20 \mu l$) and finally scanned with HPLC machine to detect the flavonoids and Phenol.

Standard: (Flavonoids and Phenol) Catechin hydrate, Myricetin, Rutin, Quercetin, Caffeic acid, Kaempferol and Gallic acid, all solution is prepared in methanol (1 mg/ml).

Mobil Phase A: methanol: acetonitrile: water: acetic acid (50 ml : 25 ml : 425 ml : 5 ml) For 0-20 min.

Mobil Phase B: methanol: acetonitrile: acetic acid (300 ml : 200 ml : 5 ml) for 20- 25 min.

RESULTS:

Organoleptic Properties: Organoleptic properties of *N. pubescens* Willd. show is shown in **Table 1**.

TABLE 1: ORGANOLEPTIC PROPERTIES OF Ν. PUBESCENS WILLD.

Nature	Coarse powder
Size	Pass through 80 mm of mesh size sieve
Colour	Brownish black
Odour	Aromatic
Taste	Bitter
Texture	Rough

Powder Microscopy: Behaviour of *N. pubescens* Willd. powder with different reagents.

Physiochemical Study: Result of physiochemical analysis of N. pubescens Willd. is shown in Table

TABLE 2: POWDER MICROSCOPY OF N. PUBESCENS

	Reagent	Observation	Characteristic
	Powder + Ruthenium red	Black colour	Mucilaginous cells are
	Powder + Sudan red	Pink colour	Cuticle is present
	Powder + Dil. HCL	Soluble	Calcium oxalate crystals a
	Powder + Safranin	Red colour	Vesicle present
	Powder + Phloroglucinol + Conc. HCl	Pink colour	Lignified cells are pro
	Powder + Dilute iodine solution + Conc. Sulphuric acid	Black colour	Hemicellulose abse
	Powder + Sulphuric acid	Brown colour	Stone cell absent
_	Powder + Dilute iodine solution	No blue colour	Endodermis without s

TABLE 3: PHYSIOCHEMICAL STUDY OF N. PUBESCENS WILLD.

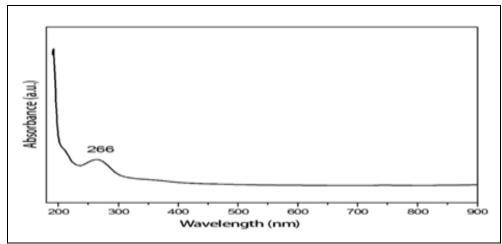
Name of study	Result (%)
Loss on Drying	11.98
Determination of total ash	6.15
Acid Insoluble Ash	11.8
Water-soluble Ash	6.3

3. The method of loss on drying has actually calculated the presence of moister of N. pubescence (11.98 %) Less number of moister prevent from bacteria and fungus⁵.

The total Ash value of N. pubescence (6.15%), indicates the presence of minerals in seed ⁶. Acid insoluble ash of N. pubescence (11.8%) indicates the presence of the earth mineral and silica. 6.3% Water-soluble Ash presence in N. pubescence Table 3.

Black colour	Mucilaginous cells are absent		
Pink colour	Cuticle is present		
Soluble	Calcium oxalate crystals are present		
Red colour	Vesicle present		
Pink colour	Lignified cells are present		
Black colour	Hemicellulose absent		
Brown colour	Stone cell absent		
No blue colour	Endodermis without starch		
The Ultraviolet, visible spectroscopy profile of graph 1 of the extract was observed at 200-900 nm wavelength range and 266 nm recorded band.			

Phenolic and flavonoid components generally absorb at 230-290 nm⁷. Hence, it conform phenolic and flavonoids are present in the extract of N. pubescens Willd.



GRAPH 1: ULTRAVIOLET VISIBLE SPECTROSCOPY OF DISTIL WATER EXTRACT OF N. PUBESCENS WILLD

Chromatography: Thin Layer Thin layer chromatography was detected by the observation of spots for identical R_f values and to determine the purity of a sample. In petroleum ether extract showed six spot, Chloroform indicate five spot,

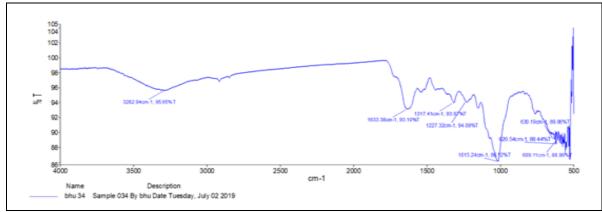
Acetone and Benzene identified seven spots, Ethanol has three spots and lastly Methanol and distill water separate two spot Table 4. Hence, the TLC profile shows compounds are smoothly separated with the help of mobile phase 8 9 10 11.

Extract	<u>N. PUBESCENS WILLD</u> Solvent Front (cm)	TLC plates	Peaks Obtained (cm)	R _f Value (cm)
Petroleum Ether	7.3	The second	S ₁ 1.4	0.30
			S ₂ 2.7	0.36
			S ₃ 3.1	0.42
		-25-0	S ₄ 3.9	0.53
			S ₅ 5.6	0.76
			S ₆ 6.7	0.91
Chloroform	7.5		$S_1 = 2.9$	0.38
			S ₂ 3.6	0.48
			S ₃ 4.6	0.61
			$\begin{array}{c} \mathbf{S}_{2} \\ \mathbf{S}_{3} \\ \mathbf{S}_{4} \\ \mathbf{S}_{4} \\ \mathbf{S}_{4} \end{array}$	0.66
			S ₅ 7.1	0.94
		- 1		
Acetone	7.5	A-1 7	$S_1 = 2$	0.26
			$S_2 = 2.8$	0.37
			$\mathbf{S}_{2}^{I} 2.8$ $\mathbf{S}_{3} 4$	0.53
		24 - C	S_4 4.3	0.55
			S ₅ 4.6	0.61
			S ₆ 6.4	0.85
			S ₇ 7.4	0.98
Benzene	6.7	.e-)	S ₁ 3.5	0.52
		T	$S_2 4.4$	0.65
			$S_3 4.6$	0.68
		E E	$S_4 4.8$	0.71
			S ₅ 5.5	0.82
			S ₆ 5.8	0.86
		L	S ₇ 6.5	0.97
E4h 1	5 (E-1-1		
Ethanol	5.6	T	$S_1 1.4$	0.25
			$S_2 2.3$	0.41 0.64
		(H	S ₃ 3.6	0.04
Methanol	7.5	me i	S ₁ 2.2	0.29
112001001	1.0	1	$S_{2} = 3.3$	0.44
		1-		
Distil Water	7	DLI	S_1 1.2	0.17
			\mathbf{S}_2 2	0.28
		10		
		0 T		
		0		
		•		

TABLE 4: TLC OF N. PUBESCENS WILLD

FTIR (Fourier- Transform Infrared Spectroscopy): FTIR is a characterization method who give the vibration energy based on peak value, compressing act of the functional group that is available on the extract of *N. pubescence*. The major bands were observed at 3282, 1633, 1317, 1227, 1015 and 630 to 609, cm⁻¹. The peak indicates **Table 5** OH stretching might be alcohol, carboxylic acid. CH stretching is alkyne. NH bending is an amine. C=C Stretching is Con. And conjugate alkene. OH, bending is Phenol. S=O Stretching is Sulfone. C-F Stretching is Fluro compound. C-N Stretching is Amine and Aromatic Amine. C-O Stretching is Alkyle arile ether, and C-X (X=Cl or Br) is halo compound $^{12, 13}$.

International Journal of Pharmaceutical Sciences and Research



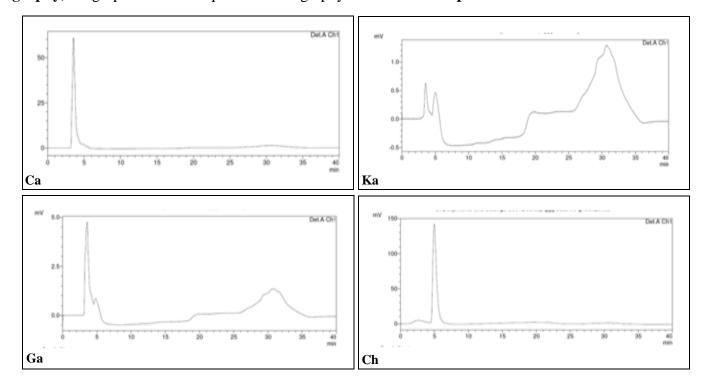
GRAPH 2: FTIR OF CRUDE EXTRACT OF *N. PUBESCENS* **WILLD.** Fourier transform infrared spectroscopy is a technique to measure the wavelength of the sample (*N. pubescens* Willd.) and find out the probable functional group

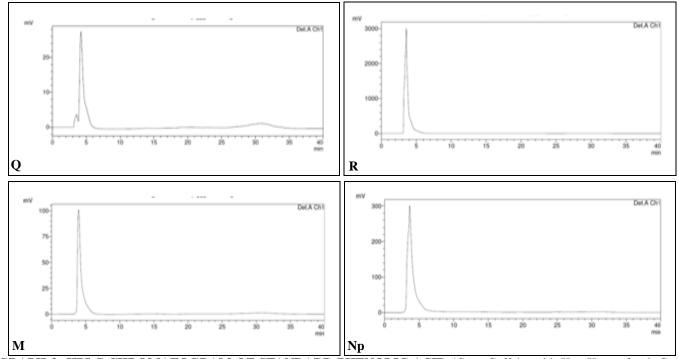
FTIR Profile				
Peak Number	X(cm-1)	Group	Compound Class	
1	3282.94	OH Stretching	Carboxylic acid, Alcohol	
		CH Stretching	Alkyne	
2	1633.38	C=C Stretching	Con. alkene	
		N-H bending	Amine	
		C=C Stretching	Conjugated alkene	
3	1317.41	O-H bending	Phenol	
		S=O Stretching	Sulfone	
		C-N Stretching	Aromatic Amine	
4	1227.32	C-F Stretching	Fluoro compound	
		C-o Stretching	Alkyl aryl ether	
		C-N Stretching	Amine	
5	1015.24	C-F Stretching	Fluoro compound	
6	630.19-609.11	C-X Stretching (X=Cl or Br)	Fluoro compound, Halo compound	

TABLE 5: FTIR OF N. PUBESCENS WILLD

HPLC (High Performance Liquid Chromatography): High-performance liquid chromatography

of methanol extract of *N. Pubescens* Willd is shown in **Graph 3**.





GRAPH 3: HPLC CHROMATOGRAM OF STANDARD PHENOLIC ACID (Ca = Caffeic acid, K = Kaempferol, Ga = Gallic acid), Flavonoids (Ch = Catechin hydrate, Q = Quercetin, R = Rutin and M= Myricetin) and Np = methanol extract of Nymphaea pubescence

TABLE 6: ANALYTICAL CONDITIONS

Column	Shim-pack GIST/GISS C 18		
Mobile Phase	Phase A- Methanol 10: Acetonitrile 5: Water		
	85: Acetic acid 1. Phase B- Methanol 60:		
	Acetonitrile 40: Acetic acid 1.		
Time Program	40 min		
Flow rate	1 ml/min		
Column	32 °C		
Temp.			
Injection Vol.	20 µL		

TABLE 7: RETENTION TIME (RT), WAVELENGTH(NM), AREA AND HEIGHT OF METHANOLEXTRACT OF N. PUBESCENCE AND STANDARD OFPHENOLIC ACID AND FLAVONOIDS FOR HPLCMETHOD VALIDATION

HPLC Profile					
Name of Extract and	λ_{max} (nm)	RT (min)	Area %	Height %	
Standard					
Methanol	254	3.624	99.189	98.564	
extract					
Caffeic acid	254	3.536	85.449	86.704	
Kaempferol	254	4.971	34.333	22.269	
Gallic acid	254	3.575	34.899	35.153	
Catechin	254	4.944	88.978	91.905	
hydrate					
Quercetin	254	4.218	90.996	86.989	
Rutin	254	3.567	99.536	99.837	
Myricetin	254	3.946	99.024	99.333	

The HPLC machine details shown in **Table 6** and in **Table 7**. The plant extract was evaluated with seven standards (gallic acid, quercotin, catechin, rutin, caffic acid, myricetin) of phenolic and flavonoids phytoconstituents to detect their capability. Caffeic acid (3.536 RT), Gallic acid (3.575), and Rutin (3.567) are present in both extracts of methanol and distill water of N. pubescence. Rutin and Caffeic acid are both possessing in previous studies on another plant of *R. arvensis*¹⁴.

CONCLUSION: The present study evaluates macroscopic, microscopic, powder microscopy, physiochemical, UV-VIS, FTIR, TLC, and HPLC of the whole plant of *N. Pubescens* Willd. for correct identification and standardization. Powder microscopy shows the presence of calcium oxalate crystal, lignified Cells, and cuticle. Physiochemical test showing 11.98% loss of the crude drug on drying, 6.15% of total ash, 11.8% of acid in soluble, 6.3% of water-insoluble ash.

The TLC result indicates that the mobile phase successfully separates the compound, and the R_f value shows the medicinal value of *N. Pubescens* Willd. FTIR shows the presence of different functional groups such as Carboxylic acid, Alcohol, Alkene, Amine, Conjugated alkene, Phenol, Sulfone, Aromatic Amine, Alkyl aryl ether, Fluoro compound, and Halo compound.

All of these functional groups have an anti-diabetic effect, especially, Tannin, flavonoids, and phenol are showing antioxidant properties as an explanation for anti-diabetic effects of the Nymphaea pubescence.

ACKNOWLEDGEMENT: The corresponding author expresses her sincere gratitude to her supervisor and co-supervisor for their guidance and providing basic facilities during this study. Grateful thanks Head department of zoology, Dr. Sonam Agrawal, Dr. Jasmeet Singh, Sanjay Kumar, Shalvi Agrawal, Veena, Imran Khan, Suraj and Arun Srivastava to providing support. Also, thankful to Dr. Anurag Mishra for the valuable advice and Dr. Anam Aftab for the required editing in the manuscript. I acknowledge sincere thanks to DST for my studies done in the Department of Kriya Sharir and Dravyaguna Laboratory.

CONFLICTS OF INTEREST: The corresponding author declares no conflicts of interest.

REFERENCES:

- 1. Swapna MM, Prakashkumar R, Anoop KP, Manju CN and Rajith NP: A review on the medicinal and edible aspects of aquatic and wetland plants of India, Journal of Medi Plant Research 2011; 5; 7163-76.
- Angadi KK, Gundampati RK, Jagannadham MV and Kandru A: Journal of Applied Pharmaceutical Science 2013; 3: 127-31.
- 3. Ministry of health & Family Welfare Government of India, The Ayurvedic Pharmacopoeia of India, The controller of

How to cite this article:

Madhulika, kumar S, Dwivedi KN and Gehlot S: Validating the properties of *Nymphaea pubescens* willd. by performing UV-Vis, TLC, FTIR and HPLC analyses. Int J Pharm Sci & Res 2021; 12(1): 615-21. doi: 10.13040/IJPSR.0975-8232.12(1).615-21.

All © 2013 are reserved by International Journal of Pharmaceutical Sciences and Research. This Journal licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported License.

This article can be downloaded to Android OS based mobile. Scan QR Code using Code/Bar Scanner from your mobile. (Scanners are available on Google Playstore)

publications civil lines, Delhi 2001; 70, 71 &102 ISBN-819011512.

- 4. Akbar S, Hanif U, Ali J and Ishtiaq S: Pharmacognostic studies of stem, root and leaf of *Malva parviflora* L. Asian Pacific Journal of Tropical Biomedicine 2014; 4; 410-15.
- 5. African Pharmacopoeia, General methods for analysis I edition 1986; (OAU/STRC) Lagos. 123.
- 6. Wagner H and Bladt S: Plant drug analysis: A thin layer chromatography atlas. Berlin; Springer 1996.
- Mishra A, Mishra VK, Dwivedi D and Dwivedi KN: UV-VIS Spectroscopic study on phytoconstituents of *Asparagus racemosus* Willd root tuber, World Journal of Pharmaceutical research 2015; 4; 10.
- 8. Kumar A, Sanjay K, Rai A and Ram B: Pharmacognostical and phytochemical evaluation of Haritaki (*Terminalia chebula* Retz.) Fruit pulp. International Journal of Pharmaceutical, Chemical and Biological Science2017; 7: 381-87.
- 9. Porika R, Poojari S, Lunavath V and Mamidala E: Preliminary phytochemical investigation and TLC Analysis of *P. angulata* fruit Extract. IOSR-Journal of Pharmacy and Biological Science 2014; 9: 11-14.
- 10. Kanoujiya SK, Chaudhary SP and Kumar N: Physiochemical study of shilajit with arjuna kwath bhvita & khadir kwath bhavita, World Journal of Pharmaceutical Research 2016; 5: 1271-80.
- Verma VK, Kumar A and Dwivedi KN: Phytochemical and Pharmacognostic study on Heartwood of Chirabilva (*Holoptelea integrifolia* planch.). Journal of Pharmacognosy and Phyto-chemistry 2018; 7: 3450-56.
- Ashok kumar R and Ramaswamy M: Phytochemical screening by FTIR spectroscopic analysis of leaf extracts of selected Indian Medical plants. International Journal of Current Microbiology and Applied Science 2014; 3: 395-06.
- 13. Mishra A, Mishra VK, Dwivedi D and Dwivedi KN: A FT-IR spectroscopic study of Phytoconstituents of *Asparagus racemosus* Willd root tuber, World Journal of Pharmaceutical Research 2015; 4: 2754-63.
- 14. Bhatti MZ, Ali A, Ahmad A, Saeed A and Malik SA: Antioxidant and phytochemical analysis of *Ranunculus arvensis* Extract, BMC Research notes 2015; 8: 297.