



Received on 11 February 2021; received in revised form, 12 May 2021; accepted, 27 May 2021; published 01 January 2022

ANTI-DIABETIC AND OXIDATIVE STRESS MARKERS IN MEN WITH TYPE 2 DIABETES MELLITUS

Hasnia Rabehi ¹, Baya Guermouche ^{*1}, Hafida Merzouk ¹, Majda Dali-Sahi ² and Sid Ahmed Merzouk ¹

Laboratory of Physiology ¹, Physiopathology and Biochemistry of Nutrition, Department of Biology, Faculty of Natural and Life Sciences, Earth and Universe, University Abou-BekrBelkaïd University, Algeria.

Department of Technical Sciences ², Faculty of Engineering, University Abou-BekrBelkaïd University, Algeria.

Keywords:

Type 2 diabetes, Metformin, Insulin, Oxidative stress, Free radicals

Correspondence to Author:

Baya Guermouche

Department of Biology,
Physiology, Physiopathology and
Nutrition Biochemistry Laboratory,
Abou Bekr Belkaïd University,
Tlemcen, 13000, Algeria.

E-mail: bguermouche@hotmail.com

ABSTRACT: Background: The aim of this study is to evaluate metformin and insulin effects on metabolic disorders and oxydatif stress markers, in Algerian men with T2D, in order to recommend the best treatment, which can minimize diabetes complications and recommend/or not a combination between metformin and insulin. Patients and **Methods:** We made this study on 120 subjects men divided into four groups (30 healthy control, 30 T2D without treatment, 30 T2D with metformin, and 30 T2D with insulin). Blood samples are collected for the determination of biochemical parameters (glucose, triglycerides, and cholesterol, high and low-density lipoprotein cholesterol) and oxidative markers (superoxide anion, nitric oxide, malondialdehyde, carbonyl proteins, oxygen radical absorbance capacity, vitamin C, catalase, glutathione). **Results:** Compared with healthy subjects, diabetic patients had altered lipid levels (cholesterol, triglycerides, LDL cholesterol) and high levels of pro-oxidant intra-cellular markers (O₂^{•-}, CP and MDA) associated to low concentrations of anti-oxidants (Vitamin C, catalase, and GSH). Our results show that insulin reduces more lipid parameters than metformin, moreover, the oxidant/antioxidant status became normal in patients treated with metformin **Conclusion:** Insulin treatment is more efficient than metformin treatment in improving the lipid profile. In addition, metformin, which reversed redox changes associated with diabetes and insulin, which improve all lipid profiles, should be prescribed in combination, especially in type 2 diabetes patients with hypertriglyceridemia and with severe oxidative stress.

INTRODUCTION: Several lipid disorders have been observed in diabetics and play a key role in the incidence of cardiovascular morbidity and mortality ¹⁻⁵.

The main lipid anomalies are quantitative and qualitative. The quantitative abnormalities, are hypertriglyceridemia and low high-density lipoprotein (HDL) cholesterol, and the qualitative abnormalities mainly include large sizes very-low-density lipoprotein (VLDL), enrichment of low-density lipoprotein (LDL) and HDL in triglycerides, LDL oxidation, and glycation of apolipoproteins ⁶. Additionally, protein glycation (PG) or non-enzymatic glycosylation is one of the consequences of hyperglycemia ^{7, 8, 9}.

	<p style="text-align: center;">DOI: 10.13040/IJPSR.0975-8232.13(1).164-72</p>
	<p style="text-align: center;">This article can be accessed online on www.ijpsr.com</p>
<p>DOI link: http://dx.doi.org/10.13040/IJPSR.0975-8232.13(1).164-72</p>	

The glycation process affects circulating proteins¹⁰. PG can be accompanied by an oxidation process, defined as an unfavorable balance, between oxygen free radicals (FR) and antioxidant systems profit first^{11, 12, 13, 14}. FR are chemical species atoms derived of oxygen (reactive oxygen species (ROS)). The presence of a single electron in a FR gives them reactivity, which can damage several molecules^{15, 16, 17, 18}. However, in T2D, the oxydat if stress (OS) acts as a mediator of IR and its progression to glucose intolerance and the pathology installation¹⁰. In conditions of severe OS, cell damage occurs with decreased pancreatic β -cell function, which is due to low expression of antioxidant enzymes¹⁹. The oxygen radical absorbance capacity (ORAC) has been found to be a good index of oxidative stress in diabetes mellitus²⁰. Clinical studies have shown that specific antioxidant concentrations in plasma and erythrocytes of diabetes patients are reduced²¹. Indeed, superoxide dismutase (SOD), glutathione peroxidase (GPx), catalase (CAT) activities scavenging, and ROS, were decreased in patients with T2D²².

Otherwise, the principal markers of lipid peroxidation (LP) are substances, which react with malondialdehyde / thiobarbituric acid, conjugated dienes, lipid hydroperoxides, and isoprostanes^{23, 24}. However, the two main biological markers of protein oxidation are the formation of protein carbonyls and nitrotyrosines groups²⁵. Several studies have demonstrated increased values of LP products²⁶, and high levels of carbonyls and AGEs in T2D plasma²⁷. In T2D, metformin is often the first glucose-lowering treatment by its effect on IR²⁸. Metformin decreases hepatic glucose output, lowers fasting glycemia, increases glucose uptake in peripheral tissues, reduces insulin resistance (IR)^{29, 30}, and decreases high blood sugar, by suppressing liver glucose production³¹. Moreover, a number of studies have established the favorable effect of metformin on body mass index (BMI) and body mass composition through the reduction of fat mass. In addition, it reduces hyperinsulinemia, lipid parameters, arterial hypertension, and endothelial dysfunction³². Metformin has been demonstrated to have a role in preventing the conversion of impaired glucose tolerance (IGT) to type 2 diabetes mellitus (T2DM) and has a beneficial effect on the blood lipid profile^{33, 34}. In addition, intensified

treatment with insulin eventually becomes necessary to maintain acceptable glycemic control in most patients with T2D³⁵, although this intervention has not been proven to reduce the risk of cardiovascular disease^{36, 37, 38}. Other studies showed a reduction in cardiovascular risk during the first 18 months of treatment with insulin alone, despite a significant increase in total and LDL cholesterol levels and body weight³⁹. In Patients treated with insulin, fasting plasma glucose and glycated hemoglobin (HbA1c) levels were reduced but weight increased³⁹. The aim of this study is to compare metformin to insulin effects on metabolic disorders and OS markers in Algerian men with T2D, in order to select the best treatment which can minimize diabetes complications and recommend/or not a combination between metformin and insulin.

MATERIALS AND METHODS:

Participants: Between January 2018 and June 2019, 120 men were recruited from the diabetic clinic of Tlemcen (Algeria), with primary criteria including non-obese (BMI < 30 kg/m²), age range between 50 and 60 years, not taking any medication, and not having a chronic disease. The subjects were divided into four groups: group I consisted of control health, group II consisted of control diabetic men without treatment, group III consisted of type II diabetic men treated with metformin, group IV consisted of type II diabetic treated with insulin. All men were non-smokers. None of subject's history of HTA, liver or renal diseases, or a history of cardiovascular diseases. Information concerning age, BMI, duration of treatment, and blood pressure were collected by questionnaire. Participation in this study was voluntary, and all subjects gave their written, informed consent. The ethical committee of the Tlemcen-University Hospital (number 01 MDU 531) approved the study.

Blood Collection: Fasting venous blood samples were collected in two tubes; EDTA tubes and dry tubes were centrifuged. Serum was separated for glucose, lipid parameters. Plasma was separated for oxidant/ antioxidant determinations. Superoxide anion and vitamin C were measured in fresh plasma samples. The remaining erythrocytes were washed, hemolyzed by the addition of cold distilled water (1/4), and the cell debris was removed by

centrifugation (2000g for 15 min). The hemolysates were assayed for antioxidant enzyme activities and GSH contents.

Biochemical Analysis: Serum glucose, triglycerides, and cholesterol contents were determined by enzymatic methods (Kits Sigma Chemical Company, St Louis, MO, USA).

Lipoprotein Isolation: Total lipoproteins were isolated from plasma by precipitation according to the method of Burstein *et al.*⁴⁰. Lipoprotein (LDL and HDL) triglyceride and total cholesterol contents were determined by enzymatic methods (Kits from Sigma).

Oxidant/antioxidant Marker Determination: Scavenging capacity of plasma: The oxygen radical absorbance capacity of plasma (ORAC) employs the oxidative loss of the intrinsic fluorescence of allophycocyanin (APC)²⁰.

Determination of Plasma Level Vitamin C: Plasma vitamin C levels were determined in plasma by using the method of Roe and Kuether⁴¹.

Determinations of Erythrocyte Antioxidant Enzyme Activity: Determination of Erythrocyte catalase (EC 1.11.1.6) activity was measured by spectrophotometric analysis of the rate of hydrogen peroxide decomposition, at 240 nm (Sigma Aldrich kit). Erythrocyte-reduced glutathione (GSH) levels were assayed by a colorimetric method, according to a Sigma Aldrich kit (Saint Louis, USA).

Determination of Superoxide Anion: The spectrophotometric determination of the $O_2^{\bullet-}$ was based on the reduction of nitrobluetetrazolium (NBT) in the presence of superoxide anion ($O_2^{\bullet-}$), a chromophor that absorbs at 550 nm⁴².

Determination of Nitric Oxide NO: Plasma and erythrocyte NO was determined by the method of Guevara *et al.*,⁴³ after deproteinization, using the colorimetric method of Griess.

Determination of Malondialdehyde: Plasma-malondialdehyde (MDA) levels were determined by the reaction of MDA with thiobarbituric acid (Sigma Aldrich kit; St. Louis, MO, USA).

Determination of Carbonyl Proteins: Plasma carbonyl proteins were determined by the

derivatization of protein carbonyl groups, with 2, 4-dinitrophenylhydrazine, leading to the formation of stable dinitrophenylhydrazone adducts (Sigma Aldrich kit).

Statistical Analysis: The results are presented as means and Standard deviations, a priori power analysis, was performed to determine the sample size, using power and sample size calculator (Statistical solutions, Sigma). The results were tested for normal distribution using the Shapiro-Wilk test. The comparison of means between the four groups is performed by ANOVA one factor. This analysis is completed by the Tukey's test to locate the source of significant difference. All tests were performed using STATISTICA 4.1 program (StatSoft, Tulsa, OK).

RESULTS:

Clinical and Biochemical Parameters: Table 1 shows that BMI was significantly higher in T2D without treatment and T2D treated with insulin and metformin compared with control subjects. No significant difference was found between T2D with metformin and T2D with insulin for BMI ($P > 0.05$). Blood pressure (systolic, SBP and diastolic, DBP) did not differ significantly among the four groups. Additionally, significant differences were found between T2D with metformin, T2D with insulin compared to control subjects and T2D without treatment for plasma glucose levels **Table 2**. The highest glucose concentrations were apparent in T2D without treatment (ANOVA, $P < 0.001$).

Lipid and Lipoprotein Levels: T2D Without treatment and T2D with metformin patients demonstrated significantly higher plasma levels of total cholesterol, compared with their control and T2D with the insulin treatment group. The highest values were observed in T2D without treatment **Table 2** ($P < 0.001$, $P > 0.05$, $P < 0.01$, respectively). Triglycerides levels were higher in subjects with untreated T2D compared to the control and treated T2D. Metformin and insulin reduce triglyceride concentration in T2D men. A greater reduction was observed with insulin treatment ($P < 0.05$).

Lipoprotein concentrations were markedly different among the four groups studied (Table 2) LDL-C amounts were significantly higher in T2D without treatment compared to T2D treated with metformin or insulin compared to control. LDL-C concen-

trations were significantly higher in T2D with metformin compared to T2D with insulin. HDL-C concentrations were significantly higher in T2D with metformin and T2D with insulin compared to control subjects and T2D without treatment. No significant difference between T2D with metformin and T2D with insulin for HDL-C. The report LDL-

C/HDL-C levels were significantly lower in T2D with metformin and T2D with insulin than controls group and T2D without treatment ($P < 0.001$), but the differences did not reach statistical significance between T2D with metformin and T2D with insulin ($P > 0.05$).

TABLE 1: CHARACTERISTICS OF THE STUDY POPULATION

Characteristic	Control	T2D without treatment	T2D with metformin	T2D with insulin	ANOVA
Number (n)	30	30	30	30	
Age (years)	53 ± 3	52 ± 2	54 ± 3	56 ± 4	
BMI (Kg/m ²)	22,49±1,23 ^c	27,06±1,89 ^a	25,83±1,85 ^b	25,94±1,60 ^b	0.0001
Duration of treatment	-	-	7±3	5±1	
SBP (mm Hg)	124±5.25	130±8.25	127±7.11	120.43±5.26	0.020
DBP (mm Hg)	76.53±4.55	84.35±5.79	80.43±5.38	86.32±5.12	0.020

Values are means ± SD. BMI, body mass index; DBP, diastolic blood pressure; SBP, systolic blood pressure. Statistical comparison between the four groups (Control, T2D without treatment, T2D treated with metformin and T2D treated with insulin) was performed by one-way ANOVA test, followed by Tukey post hoc test. Values for each parameter with different superscripts (a,b,c,d) are significantly different for $P < 0.05$, as determined by one-way ANOVA and the least significance test. P values for ANOVA test.

TABLE 2: BIOCHEMICAL PARAMETERS OF THE STUDY POPULATION

Parameter	Control	T2D without treatment	T2D with metformin	T2D with insulin	ANOVA
Glucose (mmol/L)	4,98±0,27 ^d	7,27±0,49 ^a	6,52±0,54 ^b	5,96±0,24 ^c	0.0001
TG (mmol/L)	1,45±0,09 ^c	1,85±0,22 ^a	1,66±0,15 ^b	1,56±0,17 ^b	0.0400
TC (mmol/L)	4,67±0,31 ^b	5,53±0,22 ^a	5,45±0,50 ^a	4,72±0,46 ^b	0.0001
LDL-C (mmol/L)	2,72±0,44 ^b	3,72±0,20 ^a	2,28±0,34 ^c	1,85±0,45 ^d	0.0001
HDL-C (mmol/L)	1,58±0,25 ^a	1,12±0,10 ^c	2,32±0,26 ^b	2,44±0,24 ^b	0.0001
LDL-C/HDL-C	1,81±0,48 ^b	3,37±0,39 ^a	0,99±0,23 ^c	0,81±0,12 ^c	0.0001

Values are means ± SD. TG, triglycerides; TC, total cholesterol; LDL-C, LDL cholesterol; HDL-C: HDL cholesterol. Statistical comparison between the four groups (Control, T2D without treatment, T2D treated with metformin and T2D treated with insulin) was performed by one-way ANOVA test followed by Tukey post hoc test. Values for each parameter with different superscripts (a, b, c, d) are significantly different for $P < 0.05$, as determined by one-way ANOVA and the least significance test. P values for ANOVA test.

Oxidative Stress Biomarkers:

Oxidant Status: For erythrocyte $O_2^{\bullet-}$, statistical study demonstrated a significant difference among the four groups. The levels of $O_2^{\bullet-}$ were significantly higher in T2D with metformin and T2D with insulin compared to T2D without treatment and controls ($P < 0.001$) **Table 3**. The highest level of $O_2^{\bullet-}$ was observed in T2D with insulin. A significant difference between T2D with metformin and T2D with insulin was found for Erythrocyte NO. The highest NO levels were observed in T2D treated with insulin compared to T2D without treatment and controls. However, no significant difference of NO was observed between

T2D with Metformin and T2D without treatment **Table 3**. Erythrocyte MDA levels were higher in all diabetic patients, treated with metformin or insulin and not treated compared with control men, but T2D without treatment men's values were the highest (ANOVA $P < 0.001$ and $P < 0.001$, respectively) **Table 3**. Similarly, erythrocyte carbonyls protein level was enhanced in T2D without treatment and T2D treated with metformin or with insulin compared to control cases ($P < 0.0006$) and significantly decreased with the two treatments compared to T2D without treatment.

TABLE 3: OXIDANT INTRACELLULAR MARKERS IN THE STUDY POPULATION

Parameter	Control	T2D without treatment	T2D with metformin	T2D with insulin	ANOVA
O ₂ ⁻ (μmol/L)	50,91±2,01 ^d	56,17±2,16 ^c	76,27±5,53 ^b	84,10±3,62 ^a	0.0001
NO (μmol/L)	38,16±3,06 ^b	34,82±4.41 ^c	35,42±2,92 ^c	48,93±6,05 ^a	0.0001
MDA (μmol/L Lyat)	2,11±0,27 ^c	6,17±0,26 ^a	3,35±0,45 ^b	4,02±0,33 ^b	0.0001
CP (nmol/mg protein)	2,7±0,33 ^c	6,15±0,29 ^a	3,98±0,40 ^b	3,82±0,38 ^b	0.0006

Values are means ± SD. O₂⁻, superoxide anion; NO, nitric oxide; MDA, malondialdehyde; CP, carbonyl proteins. Statistical comparison between the four groups (Control, T2D without treatment, T2D treated with metformin and T2D treated with insulin) was performed by one-way ANOVA test followed by Tukey post hoc test. Values for each parameter with different superscripts (a, b, c, d) are significantly different for $P < 0.05$, as determined by one-way ANOVA and the least significance test. P values for ANOVA test.

Antioxidant Status: Oxidative stress biomarkers Plasma total antioxidant status (ORAC) was significantly lower in T2D without treatment compared to controls **Table 4**. Values tended to increase among the two-treated group, but there is no significant difference between the group treated with metformin and insulin. Vit C levels were significantly lower in T2D without treatment compared to the control and treated groups, but its levels were significantly higher in T2D treated with metformin compared to T2D treated with insulin ($P < 0.001$) **Table 4**. Erythrocyte catalase activities were markedly different among the group studied,

it was significantly lower in T2D without treatment and T2D treated with metformin and insulin compared to control subject **Table 4**. Diabetes medication induced a rise in catalase activities but not reaching control values after treatment with insulin and metformin. Erythrocyte GSH values were significantly lower in T2D without treatment compared to control subjects and T2D treated with metformin and T2D with insulin ($P < 0.001$) **Table 4**. However, no significant difference in erythrocyte GSH activities was observed between controls cases and in both T2D treated patients.

TABLE 4: ANTIOXIDANT MARKERS IN THE STUDY POPULATION

Parameter	Control	T2D without treatment	T2D with metformin	T2D with insulin	ANOVA
ORAC (UI)	4,54±0,26 ^a	1.58±0,25 ^c	2,55±0,27 ^b	2,35±0,38 ^b	0.0001
Vit C (µmol/L)	47,48±2,25 ^a	27,92±1,28 ^c	45,88±5,87 ^a	35,36±3,12 ^b	0.0001
Catalase (UI/min/ml)	88,73±2,57 ^a	49,21±1,62 ^c	69,60±5,92 ^b	67,58±4,86 ^b	0.0001
GSH (mmol/L)	1,55±0,06 ^a	0,59±0,14 ^b	1,51±0,08 ^a	1,54±0,06 ^a	0.0010

Values are means ± SD. ORAC, oxygen radical absorbance capacity which represents total plasma antioxidant capacity; Vit C, plasma vitamin C, erythrocyte catalase levels; GSH, erythrocyte reduced glutathione levels. Statistical comparison between the four groups (Control, T2D without treatment, T2D treated with metformin and T2D treated with insulin) was performed by one-way ANOVA test followed by Tukey post hoc test. Values for each parameter with different superscripts (a, b, c, d) are significantly different for $P < 0.05$, as determined by one-way ANOVA and the least significance test. P values for ANOVA test.

DISCUSSION: Diabetes may induce multiple changes in lipids, lipoproteins, and oxidant / antioxidant status. The results obtained from the present study are found to be consistent with previous findings^{44, 45, 46}. Indeed, our results indicate that metformin and insulin are efficacious substances in treating diabetes because there is a significant decrease in plasma glucose levels in T2D patients treated with metformin and insulin as compared to untreated diabetics. The effect of metformin on blood glucose is attributed to decreased hepatic glucose production and increased glucose transport in muscle cells⁴⁷.

The lipid levels are altered, and the content in lipoproteins is significantly modified in diabetic patients compared to control subjects. The diabetic patients presented high levels of plasma triglycerides, cholesterol, and LDL-C but low levels of HDL-C. These findings are consistent with those reported in previous studies⁴⁸⁻⁵¹. In addition, an elevated atherogenicity ratio (LDL-C/HDL-C) was observed in T2D patients. These abnormalities could be attributed to hyperglycemia, insulin resistance, lipase, and CETP activities⁵¹⁻⁵³. Our findings showed that T2D patients presented high levels of pro-oxidant intra-cellular markers

(O2•-, CP and MDA) and low concentrations of anti-oxidants (Vitamin C, catalase, and GSH). These results are consistent with those reported in previous studies⁵⁴. The ORAC values were significantly lower in untreated T2D patients compared with nondiabetic control subjects. In diabetes, several mechanisms seem to be involved in the genesis of oxidative stress. Indeed, glucose auto-oxidation in the presence of iron causes the generation of reactive oxygen species (ROS)^{55, 56}, and leads to protein glycation, overproduction of superoxide radicals at the mitochondrial and NADPH oxidase levels⁵⁷⁻⁶². Reduced (NO) levels were observed in (T2D) patients, which is in agreement with the results reported in a previous study^{57, 63}. Overproduction of (NO) is associated with various inflammatory conditions, including diabetes⁶⁴⁻⁶⁸. The results of this study show a decrease in plasma triglyceride and LDL-C levels and an increase in HDL-C levels. Metformin and insulin therapies allow normalizing the lipid and lipoprotein levels in (T2D). However, the total cholesterol concentrations can be normalized by insulin only. In another study, it was indicated that metformin could play a major role in lowering blood cholesterol^{69, 70}.

In this study, it is shown that patients with T2D who were prescribed metformin presented a reduction in LDL cholesterol, unlike untreated T2D patients, which is in agreement with findings reported in previous studies^{71, 72}. Moreover, metformin and insulin also help to lower the atherogenicity ratio (LDL-C/HDL-C) in (T2D) patients, unlike untreated patients. The greatest reduction in this ratio was observed in patients treated with insulin. This helps to reduce major cardiovascular risk factors. Several studies have shown that elevated triglyceride and blood sugar levels contribute to higher (OS)⁷³. The findings from the present investigation confirm a strong association between glucose plasma concentrations and (OS) parameters. Oxidative stress in diabetes is generally caused by a diminution in the antioxidant defense system and an elevation in ROS production due to hyperglycemia⁷⁴. In addition, chronic hyper-glycemia induces an increase in protein oxidation in T2D patients⁷⁵.

The results from the present study indicate that T2D patients treated with insulin present higher levels of some pro-oxidant markers (NO, O₂•-) as compared to T2D patients treated with metformin. Moreover, significant differences were found between metformin and insulin-treated groups with regard to erythrocytes and NO. Furthermore, low levels of MDA and CP were observed in T2D patients treated with both insulin and metformin, unlike untreated diabetic patients, these findings are in good agreement with those reported in previous studies⁷⁶.

The improved control of glycemia, observed in T2D patients treated with insulin and metformin, can explain the diminution in the production of radicals and the decrease in lipid peroxidation. In our study, markers of protein oxidation, such as carbonyls, showed a significant decrease in protein oxidation in diabetic patients treated with insulin and metformin, unlike untreated patients. This decrease is attributed to low blood glucose levels in diabetic individuals treated with insulin and metformin. This is consistent with the findings of a study, which indicated that appropriate glycemic control lowers plasma levels of carbonyls in T2D patients⁷⁵. Moreover, the results of the present study indicate a significant decrease in erythrocytes, superoxide anions, and nitric oxides

(NO) in metformin-treated T2D patients as compared to those treated with insulin. Therefore, the decrease in (NO) production in diabetic patients, is at the origin of the alteration of the endothelial vasodilatation⁷⁷. On the other hand, (NO) levels are higher in insulin-treated T2D patients as compared to those treated with metformin and those not treated. This is probably due to improved glycemic control, which induces the reduction in the (OS). The present study is an attempt to measure some antioxidant defense markers, vitamin C level, catalase activity, (OS) biomarkers, plasma total antioxidant (ORAC), as well as glutathione concentration. The levels of antioxidants, vitamin C, catalase, and GSH were found higher after treatment. The (ORAC) was significantly lower in untreated T2D patients as compared to control patients, but it was found unchanged in patients treated with metformin and insulin.

The results from this study indicated higher plasma levels of vitamin C in T2D patients treated with metformin as compared to those treated with insulin. The increase in vitamin C levels can be explained by the rise in the concentration of glutathione, which is an essential factor for the enzymatic regeneration of ascorbic acid from dehydroascorbate⁷⁸.

The results obtained revealed a significant increase in erythrocyte catalase activity in treated diabetes patients as compared to untreated subjects. These results are consistent with those of several other authors who reported a decrease in the erythrocyte catalase activity of diabetic patients²⁰.

Glutathione is the main soluble antioxidant in cells^{79, 80}. The results of this study indicated a significant difference in the erythrocyte GSH levels between treated diabetic patients and untreated ones, in favor of improved antioxidant defense. Our results highlight the beneficial effects of metformin and insulin on lipid and lipoprotein profiles in T2D patients. Favorable changes in lipid and lipoprotein parameters were observed in T2D patients treated with insulin, moreover, it was noted a decrease in plasmatotal LDL cholesterol and triglyceride levels and an increase in HDL-C levels. These treatments normalize lipid and lipoprotein levels in diabetic patients.

The present work found that after therapy with metformin and insulin, plasma MDA and carbonyl protein levels significantly decreased, whereas vitamin C, GSH levels, and catalase activities remarkably increased. However, the results obtained suggest that insulin reduces more total and LDL cholesterol than metformin does; it also increases the production of NO and O₂.

In conclusion, the present study suggests that glycemia reduction in T2D subjects leads to a definite reduction in ROS generation. Metformin and insulin present beneficial effects on plasma lipids or oxidative stress markers in diabetic patients, with preexisting plasma lipid and redox abnormalities. In addition, insulin improves the lipid profile, and metformin reverses the redox changes associated with diabetes. It may be concluded that the combination of metformin and insulin may help to correct the oxidative status and lipid disorders observed in diabetic patients.

ACKNOWLEDGEMENT: This work was supported by the Algerian Research project (PNR, 2014). The authors are grateful to the patients and subjects whom volunteered to participate in this study.

CONFLICTS OF INTEREST: The authors have no relevant conflict of interest to enclose.

REFERENCES:

- Ceriello A and Prattichizzo F: Variability of risk factors and diabetes complications. *Cardiovasc Diabetol* 2021; 20(1): 7.
- Dixon ED, Nardo AD, Claudel T and Trauner M: The Role of Lipid Sensing Nuclear Receptors (PPARs and LXR) and Metabolic Lipases in Obesity, Diabetes and NAFLD. *Genes (Basel)* 2021; 12(5): 645.
- Ciccacci F, Majid N, Petrolati S, Agy M, Massango C, Orlando S, Guidotti G, Scarcella P and Marazzi MC: Hypercholesterolemia and related risk factors in a cohort of patients with diabetes and hypertension in Maputo, Mozambique. *Pan Afr Med J* 2021; 38: 102.
- Feingold KR: Role of glucose and lipids in the atherosclerotic cardiovascular disease of patients with diabetes. *Endotext [Internet]*. South Dartmouth (MA): MDText.com, Inc.; 2000-2020.
- Mei-Yueh L, Pi-Jung H, Jiun-Chi H, Wei-Hao H, Szu-Chia C and Shyi-Jang S: Association between Metabolic Syndrome, Microvascular, and Macrovascular Disease in Type 2 Diabetic Mellitus. *Am J Med Sci* 2018; 355(4): 342-49.
- Duvillard L, Florentin E and Lizard G: Cell Surface Expression of LDL Receptor is decreased in type 2 diabetic patients and is Normalized by insulin Therapy. *Diabet Care*. 2003; 26: 1540-4.
- Heidland A, Sebekova K and Schinzel R: Advanced glycation products and the progressive course of renal disease. *Am J Kidney Dis*. 2001; 38: S100-S106.
- Newsholme P, Cruzat VF, Keane KN, Carlessi R and de Bittencourt PIH: Molecular mechanisms of ROS production and oxidative stress in diabetes. *Biochem J* 2016; 473 (24): 4527-50.
- Park CH and Jae WK: Effect of Advanced Glycation End Products on Oxidative Stress and Senescence of Trabecular Meshwork Cells. *Korean J Ophthalmol* 2012; 26: 123-31.
- Byrne NJ, Rajasekaran NS, Abel ED and Bugger H: Therapeutic potential of targeting oxidative stress in diabetic cardiomyopathy. *Free Radic Biol Med* 2021; 169: 317-42.
- Ndrepepa G: Myeloperoxidase - A bridge linking inflammation and oxidative stress with cardiovascular disease. *Clin Chim Acta* 2019; 493: 36-51.
- Jakubczyk K, Dec K, Kalduńska J, Kawczuga D, Kochman J and K Janda: Reactive oxygen species - sources, functions, oxidative damage. *Pol Merkur Lekarski* 2020; 48(284): 124-27.
- Vaidya AR, Wolska N, vara d, mailer rk, schröder k and pula g: diabetes and thrombosis: a central role for vascular oxidative stress. *Antioxidants (Basel)*. 2021; 10(5): 706.
- Rani V, Deep G, Singh RK, Palle K and Yadav UCS: Oxidative stress and metabolic disorders: Pathogenesis and Therapeutic Strategies 2016; 148: 183-93.
- Chainy GBN and Sahoo DK: Hormones and oxidative stress: an overview. *Free Radic Res* 2020; 54(1): 1-26.
- Kumari N, Haider MR, Pathak A and Yar MS: Medicinal prospects of antioxidants. *Eur J Med Chem* 2019; 178: 687-04.
- Pisoschi AM, Pop A, Iordache F, Stanca L, Predoi G and Serban AI: Oxidative stress mitigation by antioxidants - An overview on their chemistry and influences on health status. *Eur J Med Chem* 2021; 209: 112891.
- Kancheva VD, Dettori MA, Fabbri D, Alov P, Angelova SE, Slavova-Kazakova AK, Carta P, Menshov VA, Yablonskaya OI, Trofimov AV, Tsakovska I and Sas L: Natural Chain-Breaking Antioxidants and Their Synthetic Analogs as Modulators of Oxidative Stress. *Antioxidants (Basel)*. 2021; 10(4): 624.
- Pomatto LCD and Davies KJA: Adaptive homeostasis and the free radical theory of ageing. *Free Radic Biol Med* 2018; 124: 420-30.
- Merzouk S, Hichami A, Sari A, Madani S, Merzouk H, Yahia and Berrouiguet A: Impaired oxidant/antioxidant status and LDL-fatty acid composition are associated with increased susceptibility to peroxidation of LDL in diabetic patients. *Gen Physiol Biophys* 2004; 23: 387- 99.
- Alfonso-Muñoz EA, Burggraaf-Sánchez de Las RM, Boronat JM, César J, Martín M and Desco C: Role of Oral Antioxidant Supplementation in the Current Management of Diabetic Retinopathy. *International Journal of Molecular Sciences* 2021; 22(8): 4020.
- He X, Kuang G, Yi Z, Li S, Zhou S and Ou C: The role of non-coding RNAs in diabetic nephropathy-related oxidative stress. *Front Med (Lausanne)*. 2021; 8: 626423.
- Gaschler MM and Stockwell BR: Lipid peroxidation in cell death. *Biochem Biophys Res Commun* 2017; 482(3): 419-25.
- Van der PJ: Effect of lipid peroxidation on membrane permeability of cancer and normal cells subjected to oxidative stress. *Chemical Science* 2016; 7(1): 489-98.
- Wang Z, Wang Y, Liu H, Che Y and Xu Y: Age-related variations of Protein carbonyls in human saliva and

- plasma: Is saliva protein carbonyls an alternative biomarker of aging? *Age Dordr* 2015; 37: 81-97.
26. Kattoor AJ, Pothineni NVK, Palagiri D and Mehta JL: Oxidative Stress in Atherosclerosis. *Curr Atheroscler Rep* 2017; 19(11): 42.
 27. Cakatay U: Protein oxidation parameters in type 2 diabetic patients with good and poor glycaemic control. *Diabetes & metabolism*. ISSN 1262-3636, 2005; 31(6): 551-57.
 28. Madsen KS, Chi Y, Maria-Inti M, Richter B and Hemmingsen B: Metformin for prevention or delay of type 2 diabetes mellitus and its associated complications in persons at increased risk for the development of type 2 diabetes mellitus. *Cochrane Database Syst Rev* 2019; 12.
 29. Mosenzon O, Prato SD, Schechter M, Leiter LA, Ceriello A, DeFronzo RA and Raz I: From glucose lowering agents to disease/diabetes modifying drugs: a "SIMPLE" approach for the treatment of type 2 diabetes. *Cardiovasc Diabetol* 2021; 20(1): 92.
 30. Chun-Yu L, Chun-Hsin W, Chung-Yuan H, Tien-Hsing C, Ming-Shyan L, Yu-Sheng L and Yu-Jih S: Reduced mortality associated with the use of metformin among patients with autoimmune diseases. *Front Endocrinol (Lausanne)*. 2021; 12: 641635
 31. Natali A and Ferrannini E: Effects of metformin and thiazolidinediones on suppression of hepatic glucose production and stimulation of glucose uptake in type 2 diabetes: a systematic review. *Diabetologia* 2006; 49: 434-41.
 32. Madsen KS, Kähler P, Kähler LKA, Madsbad S, Gnesin F, Maria-Inti M, Richter B, Hemmingsen B and Cochrane: Metabolic and endocrine disorders group metformin and second- or third-generation sulphonylurea combination therapy for adults with type 2 diabetes mellitus. *Cochrane Database Syst Rev* 2019; 2019(4): CD012368.
 33. Zilov AV, Abdelaziz SI, AlShammary A, Zahrani AA, Amir A, Khalil SHA, Brand K, Elkafrawy N, Hassoun AAK, Jahed A, Jarrah N, Mrabeti S and Paruk I: Mechanisms of action of metformin with special reference to cardiovascular protection. *Diabetes Metab Res Rev* 2019; 35(7): e3173.
 34. Ursini F, Russo E, Pellino G, D'Angelo S, Chiaravalloti A, De Sarro G, Manfredini R and De Giorgio R: Metformin and Autoimmunity: A "New Deal" of an Old Drug. *Front Immunol*. 2018; 9: 1236.
 35. Inzucchi SE, Bergenstal RM and Buse JB: Management of hyperglycaemia in type 2 diabetes: a patient-centered approach. Position statement of the American Diabetes Association (ADA) and the European Association for the Study of Diabetes (EASD). *Diabetologia* 2012; 55: 1577-96.
 36. Bakh NA, Cortinas AB, Weiss MA, Langer RS, Anderson DG., Gu Z, Dutta S and Strano MS: Glucose-responsive insulin by molecular and physical design. *Nat Chem* 2017; 9: 937-43.
 37. Hemmingsen B, Lund SS and Gluud C: Intensive glycaemic control for patients with type 2 diabetes: systematic review with meta-analysis and trial sequential analysis of randomised clinical trials. 2011; *BMJ* .343:d6898.
 38. Baumgard LH, Hausman GJ and Fernandez MVS: Insulin: pancreatic secretion and adipocyte regulation. *Domest Anim Endocrinol* 2016; 54: 76-84.
 39. St EO: Diabetes: Pharmacotherapy for Type 2 Diabetes. *FP Essent* 2021; 504: 22-27
 40. Burstein M, Fine A, Atger V, Xirbel E and Girard-Globa A: rapid method for isolation of two purified subfraction of high density lipoproteins by differential dextran sulfate-magnesium choride precipitation. *Biochem* 1989; 71: 741-46.
 41. Roe JH and Kuether CA: The determination of ascorbic acid in whole blood and urine through the 2, 4-dinitrophenylhydrazine derivatives of dehydroascorbic acid. *J Biol Chem* 1943; 147: 399-07.
 42. Auclair C and Voisin E: Nitrobluetetrazolium reduction. In: Greenworld R A, editor. *Handbook of Methods for Oxygen Radical Research*. Boca Raton: CRC Press, Inc 1985; 123-32.
 43. Guevara I, Iwanejko J and Dembińska-Kieć A: Determination of nitrite/nitrate in human biological material by the simple Griess reaction. *Clin Chim Acta* 1998; 274: 177-88.
 44. Ting Y, Jia-Xing Z, Fa-Xuan W, Jian-Hua Z, Yu Z, Lan L, Xiu-Ying L, Yu-Hong Z, Yi Z: The Association between Sarcopenic Obesity and Hypertension, Diabetes, and Abnormal Lipid Metabolism in Chinese Adults. *Diabetes Metab Syndr Obes* 2021; 14: 1963-73.
 45. Azarpazhooh MR, Najafi F, Darbandi M, Kiarasi S, Oduyemi T and Spence JD: Triglyceride/high-density lipoprotein cholesterol ratio: a clue to metabolic syndrome, insulin resistance, and severe atherosclerosis. *Lipids* 2021. Doi: 10.1002/lipd.12302.
 46. Hirano T, Kodera R, Hirashima T, Suzuki N, Aoki E, Hosoya M, Oshima T, Hayashi T, Koba S, Ohta M, Satoh N and Ito Y: Metabolic properties of lowdensity lipoprotein (ldl) triglycerides in patients with type 2 Diabetes, Comparison with Small Dense LDL-Cholesterol. *J Atheroscler Thromb* 2021. Doi: 10.5551/jat.62789.
 47. Zhou G, Myers R and Li Y: Role of AMP-activated protein kinase in mechanism of metformin action. *J Clin Invest* 2001; 108: 1167-74.
 48. Shah PK: Inflammation, infection and atherosclerosis. *Trends Cardiovasc Med* 2019; 28: 468-72.
 49. Poznyak A, Andrey V, Grechko, Poggio P, Myasoedova VA, Alfieri V and Orekhov AN; The diabetes mellitus-atherosclerosis connection: the role of lipid and glucose metabolism and chronic inflammation. *Int J Mol Sci* 2020; 21(5): 1835.
 50. Summerhill VI, Grechko AV, Yet SF, Sobenin IA and Orekhov AN: The atherogenic role of circulating modified lipids in atherosclerosis. *Int J Mol Sci* 2019; 20: 3561.
 51. Taleb S: Inflammation in atherosclerosis. *Arch Cardiovasc Dis* 2016; 109: 708-15.
 52. Vergès B: Lipid modification in type 2 diabetes: the role of LDL and HDL. *Fundam Clin Pharmacol* 2009; 23(6): 681-5.
 53. Stankova TR, Delcheva GT, Maneva AI, Vladeva SV: Serum Levels of Carbamylated LDL, nitrotyrosine and soluble lectin-like oxidized low-density lipoprotein receptor-1 in poorly controlled type 2 diabetes mellitus. *Folia Med (Plovdiv)* 2019; 61(3): 419-25.
 54. Roma LP and Jean-Christophe J: Nutrient Metabolism, Subcellular Redox State, and Oxidative Stress in Pancreatic Islets and β -Cells. *J Mol Biol* 2020; 432(5): 1461-93
 55. Gerber PA and Rutter GA: The Role of Oxidative Stress and Hypoxia in Pancreatic Beta-Cell Dysfunction in Diabetes Mellitus. *Antioxid Redox Signal* 2017; 26(10): 501-18.
 56. Yan L: Pathogenesis of chronic hyperglycemia: from reductive stress to oxidative stress. *J Diabetes Res* 2014; 14: 1-11.
 57. Pitocco D, Zaccardi F, Di Stasio E, Romitelli F, Santini S A, Zuppi C and Ghirlanda G: Oxidative stress, nitric oxide, and diabetes. *Rev Diabet Stud* 2010; 7: 15-25.

58. Xiao-Bin H, Ting-Hui L, Zhi-Peng R and Liu Y: Combination of 2-deoxy d-glucose and metformin for synergistic inhibition of non-small cell lung cancer: A reactive oxygen species and P-p38 mediated mechanism. *Biomed Pharmacother* 2016; 84: 1575-58.
59. Ola MS, Berkich DA and Xu Y: Analysis of glucose metabolism in diabetic rat retinas," *The American Journal of Physiology: Endocrinology and Metabolism*. 2006; 290(6): E1057-E1067.
60. Rosca MG, Vazquez E J, Chen Q, Kerner J, Kern TS and Hoppel CL: Oxidation of fatty acids is the source of increased mitochondrial reactive oxygen species production in kidney cortical tubules in early diabetes. *Diabetes* 2012; 61(8): 2074-83.
61. Kulaksızoglu S and Karalezli A: Aqueous humour and serum levels of nitric oxide, malondialdehyde and total antioxidant status in patients with type 2 diabetes with proliferative diabetic retinopathy and nondiabetic senile cataracts. *Can J Diabetes* 2016; 40(2): 115-9.
62. Oguntibeju OO: Type 2 diabetes mellitus, oxidative stress and inflammation: examining the links. *Int J Physiol Pathophysiol Pharmacol* 2019; 11(3): 45-6.
63. Oleson BJ and John A: Corbett. Dual Role of Nitric Oxide in Regulating the Response of β Cells to DNA Damage. *Antioxid Redox Signal* 2018; 29(14): 1432-45.
64. Afonso V, Champy R, Mitrovic D, Collin P and Lomri A: Reactive oxygen species and superoxide dismutases: role in joint diseases, *Jt. Bone Spine*. 2007; 74: 324-29.
65. Velagic A, Qin C, Owen LL, Woodman, Horowitz JD and Rebecca H, Ritchie and Kemp-Harper BK: Nitroxyl: a novel strategy to circumvent diabetes associated impairments in nitric oxide Signaling. *Front Pharmacol* 2020; 11: 727
66. Al-nimer MSM, Al-obaidi SAH and Al-dulaimi KS: Serum nitric oxide and Peroxynitrite levels in adult seropositive rheumatoid arthritis treated with disease modifying antirheumatic drugs: a preliminary report, *Turk. J Med Sci* 2010; 40-191e197.
67. Luvuno M, Khathi A and Mabandla MV: Diet-induced prediabetes: effects of exercise treatment on risk factors for cardiovascular complications. *Nutr Metab (Lond)*. 2021; 18(1): 45.
68. Sanz-Cameno P, Medina J, Garcia-Buey L, Garcia-Sanchez A, Borque MJ, Martin Vilchez S, Gamallo C, Jones EA and Moreno-Otero R: Enhanced intrahepatic inducible nitric oxide synthase expression and nitrotyrosine accumulation in primary biliary cirrhosis and autoimmune hepatitis. *Journal of Hepatology* 2002; 37-723e729.
69. Van SMF, de Graaf AA and Albert K: Groen. Actions of metformin and statins on lipid and glucose metabolism and possible benefit of combination therapy. *Cardiovasc Diabetol* 2018; 17: 94.
70. Vergès B: Insulinosensibilité et lipides. *Diabetes & Metabolism* 2001; 27-2-1262-36-36-01019-Art7.
71. Shokrpour M, Foroozanfard F, Ebrahimi FA, Vahedpoor Z, Aghadavod E, Ghaderi A and Asemi Z: Comparison of myo-inositol and metformin on glycemic control, lipid profiles, and gene expression related to insulin and lipid metabolism in women with polycystic ovary syndrome: a randomized controlled clinical trial. *Gynecol Endocrinol* 2019; 35(5): 406-11.
72. Geerling JJ, Boon MR and van der Zon GC, van den Berg SA and van den Hoek AM: Metformin lowers plasma triglycerides by promoting VLDL-triglyceride clearance by brown adipose tissue in mice. *Diabetes* 2014; 63(3): 880-91.
73. Yarıbeygi H, Sathyapalan T, Atkin SL and Sahebkar A: Molecular mechanisms linking oxidative stress and diabetes mellitus. *Oxid Med Cell Longev* 2020; 2020: 8609213.
74. Zhuang T, Han H and Yang Z: Iron, Oxidative Stress and Gestational Diabetes. *Nutrients* 2014; 6: 3968-80.
75. Alhagh E, Gorgich C, Parsaie H, Yarmand S, Baharv F and Sarbishegi M: Long-Term administration of metformin ameliorates age-dependent oxidative stress and cognitive function in rats. *Behav Brain Res* 2021; 113343
76. Grindel A, Guggenberger B, Eichberger L, Pöppelmeyer C, Gschaider M, Tosevska A, Mare G, Briskey D, Brath H and Karl-Heinz Wa: Oxidative Stress, DNA Damage and DNA Repair in Female Patients with Diabetes Mellitus Type 2. *PLoS One*. 2016; 11(9): e0162082
77. Santilli F, Cipollone F and Mezzetti A: The role of nitric oxide in the development of diabetic angiopathy. *Horm Metab Res* 2004; 36: 319-35.
78. Cammisotto V, Nocella C, Bartimoccia S, Sanguigni V, Francomano D, Sciarretta S, Pastori D, Peruzzi M, Cavarretta E, D'Amico A, Castellani V, Frati G, Carnevale R and SMiLe Group: The Role of Antioxidants Supplementation in Clinical Practice: Focus on Cardiovascular Risk Factors. *Antio (Basel)* 2021; 10(2): 14
79. Orhan H, Onderoglu L, Yücel A and Sahin G: Circulating biomarkers of oxidative stress in complicated pregnancies. *Arch Gynecol Obstet* 2003; 267: 189-195.
80. Xepapadaki E, Zvintzou E, Kalogeropoulou C, Filou S and Kypreos KE: The Antioxidant Function of HDL in Atherosclerosis. *Angiology* 2020; 71: 112-21.

How to cite this article:

Rabehi H, Guermouche B, Merzouk H, Dali-Sahi M and Merzouk SA: Anti-diabetic and oxidative stress markers in men with type 2 diabetes mellitus. *Int J Pharm Sci & Res* 2022; 13(1): 164-72. doi: 10.13040/IJPSR.0975-8232.13(1).164-72.

All © 2022 are reserved by the International Journal of Pharmaceutical Sciences and Research. This Journal licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported License.

This article can be downloaded to **Android OS** based mobile. Scan QR Code using Code/Bar Scanner from your mobile. (Scanners are available on Google Playstore)