E-ISSN: 0975-8232; P-ISSN: 2320-5148



PHARMACEUTICAL SCIENCES



Received on 12 April 2021; received in revised form, 15 June 2021; accepted, 21 June 2021; published 01 February 2022

VALIDATION STUDY FOR PARACETAMOL DETERMINATION IN LIQUID PRODUCTS (PARACEROL) USING RAMAN SPECTROSCOPY

T. Kocaağa ¹, F. Yıldırım ¹, U. Kumrulu ¹, E. Kumrulu ¹, N. Mutlu ¹, E. Sopacı ¹, B. Demirdoğan ¹ and D. Giray Dilgin ^{* 2}

Polifarmaİlaç San. ve Tic. AŞ ¹, Ergene/Tekirdağ, Turkey.

Secondary Science and Mathematics Education Department ², Faculty of Education, University of Canakkale Onsekiz Mart, 17100, Canakkale, Turkey.

Keywords:

Raman spectroscopy, Validation, Paracetamol, Quantification

Correspondence to Author: D. Giray Dilgin

Associated Professor in Analytical Chemistry, University of Çanakkale Onsekiz Mart, Faculty of Education, Secondary Science and Mathematics Education Department, 17100, Çanakkale, Turkey.

E-mail: didemgiray79@hotmail.com

ABSTRACT: In this study, Raman spectroscopy was used as an analytical quality control methodology to evaluate the paracetamol (PCT) quantity in a commercially available formulation in the pharmaceutical industry. The Raman Spectroscopy has been established for the validation and the quantification of PCT in a commercial product called Paracerol (Prc) Solution for I.V. Infusion containing 10.0 mg.mL⁻¹ PCT. The validation of the proposed method was performed by evaluating the specificity, linearity range, accuracy, precision, detection limit, quantification limit, and robustness parameters. The acceptable results for all parameters showed that the proposed method is valid. The Raman spectroscopic method was also compared with the HPLC method in terms of PCT determination and statistical tests such as t and F tests and the obtained results indicated that there is no any difference in the averages and precision results between the proposed Raman spectroscopy and HPLC methods. It has been reported that Raman spectroscopy can be applied in the pharmaceutical industry for the quality control analysis of the commercial formulation of PCT.

INTRODUCTION: (PCT) Paracetamol or (IUPAC acetaminophen name is N-(4hydroxyphenyl) acetamide) has been widely used as an active pharmaceutical ingredient in analgesic and antipyretic drugs. It has been first prepared by Morse in 1878 1 and has been used as a drug since 1956². This important compound is often found as the main ingredient in numerous cold and flu medicines, and it is widely used to relieve many aches and pains, especially headache.



DOI: 10.13040/IJPSR.0975-8232.13(2).755-62

This article can be accessed online on www.ijpsr.com

DOI link: http://dx.doi.org/10.13040/IJPSR.0975-8232.13(2).755-62

The combination of PCT with other drugs such as opioid analgesics increases the effectiveness of its pain reliever and thus it can also be used in the treatment of more severe pain after surgery and for relieving of the pain of cancer patients ³.

In the market, PCT is available as an over-the-counter drug with different brand names and can be found in different forms such as tablets, liquids (as suspensions and solution), capsules, and suppositories. PCT can be taken with food or on an empty stomach. It is not listed as a nonsteroidal anti-inflammatory drug (NSAID) like other pain killers like aspirin and ibuprofen ³ for that reason; it is the most widely accepted drug. However, taking too much PCT than recommended doses (1.0 g/single dose and 4.0 g/day for adults) can lead to fatal liver damage, kidney problems, inflammation

in the pancreas, skin rashes, low blood sugar and even death ⁴. Therefore, a lot of analytical methods such as chromatography ⁵⁻⁶, spectrophotometry ^{4, 7-10} and electrochemical methods ^{5, 11} have been developed for the determination and validation of PCT due to its great importance in human health.

The validation of the drug products is a very important process in the manufacture pharmaceuticals. The compliance of the products includes the identity, purity, stability, potency of the drug etc. The quantitative analytical methods based on the structure and the properties of the active component of the drug formulation are utilized in drug industries. In the method development and validation for determination of a drug in the manufacture of pharmaceuticals, several analytical techniques such as chromatography (GC, HPLC), capillary electrophoresis (CE), spectroscopy (Uv-Vis, Fluorescence, NIR, NMR, AAS, ICP) and especially hyphenated methods (GC-MS, LC-MS, ICP-MS) have been widely used ¹². Among them the chromatographic techniques (especially liquid chromatography) have attracted a great attention in the validation of the drug formulation since these methods offer rapidity in process, better peak shape, higher resolution, accuracy, and precision and no isolation process ¹³. Although the chromatographic methods exhibit these advantages, they are destructive methods as well as they require time consuming sample pretreatment steps and use large volume of organic or inorganic solvents during sample preparation and analysis ¹⁴⁻¹⁶. Therefore, there is a need to develop rapid and reliable alternative analytical methods containing green chemistry for the validation of the drug products.

Recently, vibrational spectroscopic methods (IR, NIR and Raman) have been taken a place as an alternative green analytical method in the pharmaceutical industry ¹⁴⁻²⁰. These methods have several advantages over the chromatographic methods such as label free, fast acquisition, solvent free, non-invasive, non-destructive, low-cost, minimization of the sample preparation step, direct measurement inside packaging material (glass, plastic *etc.*) and possibility to use probes ¹⁴⁻¹⁷. Although IR and NIR are very useful techniques for the identification and validation of drug samples in the pharmaceutical industry, Raman

spectroscopy has some superiorities over IR and For example, aqueous pharmaceutical formulations can be directly studied due to being Raman-inactive of water molecules and Raman spectra have higher chemically specificity and analysis speed than NIR ^{15, 17}. Moreover, Raman spectroscopy can give information at the molecular level to identify unknown solids or liquids of both organic and inorganic materials. It is also a practical method for identification and verification of known components and the determination of the impurities in pharmaceuticals. In this context, many studies based on Raman and FT spectroscopic techniques have previously been carried out for the quantitative determination and validation of many drugs in liquid or solid pharmaceutical formulations ^{9, 10, 14-16, 19-36}.

The Raman spectrophotometric determination of PCT in the pharmaceutical solution (Paracerol, Prc) has been described in this study. Although the PCT determination was studied ^{9, 33-35} in some solid and liquid pharmaceuticals, a detailed validation study has not been reported yet. Therefore, this study reports the validation based on using the parameters of sensitivity, selectivity, accuracy, reproducibility, repeatability, stability, linear range, and robustness and determination of PCT in Prc solution.

MATERIALS AND METHODS:

Apparatus: Thermo Scientific TruScanTM RM Handheld Analyzer was used to perform the Raman Spectroscopic measurements. It has a 785 nm excitation laser (250 mW) covering a range of 250-2875 cm⁻¹. Raman shifts have a spectral resolution of 8–10.5 cm⁻¹ (FWHM) across the range. The principle of the technology is based on the optimization of the signal-to-noise ratio so that the analysis stops when this ratio is considered optimal inducing different acquisition parameters for each sample. Recently, a new upgrade known as TruToolsTM was released, allowing the operator to set acquisition parameters, develop qualitative and quantitative chemometric models, and embed them into the spectrophotometer. As the need to quickly identify and quantify more materials increases, the pharmaceutical and biotechnology manufacturers require instruments that decrease laboratory sample testing and enable more at-line decisions. The Thermo ScientificTM TruScanTM RM analyzer with TruToolsTM embedded chemometrics package provides users with the flexibility to build customized qualitative and quantitative methods for complex material analysis problems. The HPLC experiments as a reference method were performed with the HPLC system UV detector. A column of C18 (300 \times 3.9 mm, 10 μ m) and a mobile phase solution that is a mixture of high purified water and methanol in a ratio of 3:1were used in the HPLC.

Reagent and Solutions: Paracetamol (%99.8) was supplied from Hebei Jiheng Pharmaceutical company and all other used chemicals such as Mannitol, L-cysteine hydrochloride, disodium phosphate were at analytical grade. The PCT serum solution (Prc) containing 10.0 mg.mL⁻¹ studied in the validation was produced by Polifarma company in Turkey. A placebo (Plc) solution was prepared by weighing precisely 3.85 g of mannitol, 25.0 mg of cysteine hydrochloride monohydrate, and 13.0 mg of disodium phosphate dihydrate to a 100 mL volumetric flask and diluting to volume with pure water.

The stock solution of PCT (20.0 mg.mL⁻¹) was prepared by weighing precisely 4.0 g of PCT to a 200 mL volumetric flask and diluting to the volume with pure water. Standard PCT solutions between 7.0 and 13.0 mg.mL⁻¹ PCT were prepared by diluting a known volume of stock PCT solution to 20 mL with pure water.

Validation of the Method: The proposed Raman spectroscopic method was validated according to the ICH guidelines ³⁷ and according to the guidelines described by Ferenczi-Fodor *et al.* ³⁸ for the following parameters: specificity, linearity range, accuracy, precision (system precision, repeatability, and intermediate precision), detection limit, quantitative limit, and robustness.

Specificity: The specificity of the method was checked by recording the spectra of the Plc solution, the standard solution, and the test solution, respectively.

Linearity and Range: The linearity of the proposed method was evaluated by analyzing five standard solutions of PCT (7.0; 9.0; 10.0; 11.0, and 13.0 mg mL⁻¹) prepared by diluting with pure water. The experiments were performed in three different analyses.

Accuracy: The accuracy of the method was evaluated by measuring the recovery after adding PCT to Plc solution such that the linearity range was in the low (8.0 mg.mL⁻¹), medium (10.0 mg mL⁻¹), and high (12.0 mg.mL⁻¹) levels. The experiments were performed in three parallel measurements containing three analyzes of each.

Precision: The system precision was evaluated by six consecutive measurements of the standard PCT solution of 10.0 mg.mL⁻¹. The repeatability was tested by measuring Prc solution (10 mg mL⁻¹ PCT) for six times.

In addition, intermediate precision was tested by measuring six different Prc solution for infusion on different days by a different analyst in the same laboratory.

The precision was evaluated as the standard deviation (SD) or relative standard deviation percentage (RSD%), and the results were compared according to the acceptance criteria.

Limit of Detection (LOD) and Limit of Quantification (LOQ): The signal of Plc solutions that do not contain PCT was measured six times under the studied conditions.

Then the LOD and LOQ were calculated using 3.3 sbl/m, and 10 sbl/m equations, respectively, where sbl is the standard deviation of the response of Plc solution accepted as blank solution and m is the slope of the calibration curve.

Robustness: The robustness of the method was tested by evaluation of the signal of Prc solution for infusion based on the changes of parameters such as temperature and pH.

RESULTS AND DISCUSSION: The Raman spectrum of 10.0 mg mL-1PCT solution was recorded in **Fig. 1** to define the PCT peaks. It can be seen that the characteristic peaks of PCT were observed, such as phenyl-N bending mode at 1176 cm⁻¹, stretching of the hydroxyl group with respect to the phenyl moiety at 1240 cm⁻¹, CC, CN stretching, and CH bending at 1331 cm⁻¹, CH₃ bending at 1382 cm⁻¹, CC stretching and NH deformation at 1517 cm⁻¹, C-C stretching and NH deformation peak at 1620 cm⁻¹. Similar results were also obtained in the previous reported studies ^{9,34}.

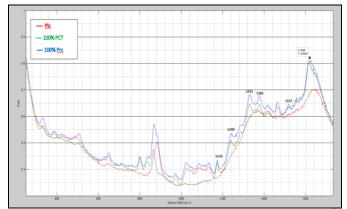


FIG. 1: RAMAN SPECTRA OF PLC, PCT (10 mg mL⁻¹) AND PRC SOLUTIONS FOR I.V INFUSION (CONTAINING 10 mg mL⁻¹ PCT)

Validation of the Raman Spectroscopic Method for Prc Solution:

Specificity: The specificity demonstrates the ability of the analytical method to measure accurately and selectively the analyte in the presence of components that may be expected to be present in the sample matrix. It was already mentioned that Raman spectroscopic and especially chromatographic methods were used and validated for PCT drugs and solutions. However, our literature survey shows that the spectroscopic method has not been used to validate a PCT drug solution, Prc. For this purpose, the Raman spectra of the Plc, standard (100% or 10 mg mL⁻¹ PCT) and test solutions (100%) were recorded under same conditions, and they were shown in Fig. 1. It was observed that the excipients present in the Prc formulations did not interfere with the PCT peak. These results indicate that an acceptable specificity was obtained for the Raman spectroscopic determination of PCT in Prc formulations.

Calibration and Linearity Range: The calibration curve demonstrates that the analytical method is capable to obtain test results which are directly proportional to the analyte concentration in a sample. In this context the Raman spectra of various concentration of PCT in the range from 7.0 to 13.0 mg mL⁻¹ were given in **Fig. 2**. The calibration curve for the real values versus predicted concentration was presented in **Fig. 3**. This figure shows that a linear relationship exists between real concentration and experimentally obtained PCT concentration in the range from 7.0 to 13.0 mg mL⁻¹.

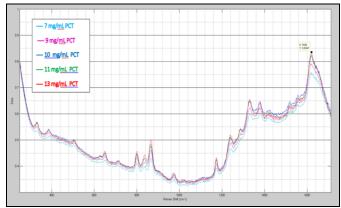


FIG. 2: RAMAN SPECTRA OF PCT SOLUTIONS OF DIFFERENT CONCENTRATIONS IN THE RANGE FROM 7.0 TO 13.0 mg mL⁻¹

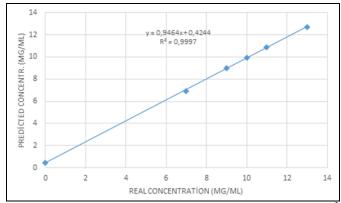


FIG. 3: REAL PCT CONCENTRATIONS AS mg mL⁻¹ VS. CALCULATED CONCENTRATIONS RESULTING FROM PARTIAL LEAST SQUARES ANALYSIS

Accuracy: The accuracy of an analytical procedure expresses the closeness of agreement between the value which is accepted either as a true conventional value or as an accepted reference value and the value found. After the addition of a known amount of PCT to Plc solution, the recovery results were used to evaluate the accuracy of the proposed method. The recovery values given in **Table 1** for three different concentrations of PCT were obtained in the range between 100.4% and 101.9%. The low RSD values (RSD<2%) indicate that the proposed method has a high accuracy for PCT determination in Prc solution.

Precision: The precision of an analytical method is described as the harmonization of the results obtained from the replications of a homogenous sample. This protocol includes system precision, repeatability, and intermediate precision. The system precision is tested at the same conditions and in a short time interval. It is the measurement of the system performance parameter that is

E-ISSN: 0975-8232; P-ISSN: 2320-5148

independent of errors occurred at the sample preparation stage. The results of the standard solution of 10.0 mg mL⁻¹ obtained from six consecutive injections were presented in Table 2. The repeatability is tested under the same conditions over a short interval of time. It is the measurement of the system performance dependent on the errors that occurred at the sample preparation. The results of six different Prc solution for I.V infusion drug product (containing 10 mg mL⁻¹ PCT) were given in **Table 2**. The

intermediate precision determines the effects of the changes on the results of a homogenous sample analyzed on a different day by a different analyst. It is not necessary to make each of the changes at the same time. The results of six different test solutions (Prc 10.0 mg mL⁻¹ Solution for I.V. Infusion) analyzed in the same laboratory on a different day by a different analyst were also presented in **Table** 2. The obtained RSD values are smaller than 1% in all precision studies indicating that the proposed method is highly precise for PCT determination.

TABLE 1: ACCURACY RESULTS OBTAINED FROM DIFFERENT CONCENTRATIONS OF PCT (8.0, 10.0, AND

12.0 mgmL ⁻¹) Theoretical value	Found value	Mean	SD	RSD	Mean	Recovery	SD	RSD%
$(mg mL^{-1})$	(mg mL ⁻¹⁾			%	(n=9)	%	(n=9)	(n=9)
8.0 (1)	8.10	8.08	0.029	0.36	8.15	101.9	0.075	0.92
	8.05							
	8.10							
8.0 (2)	8.28	8.23	0.042	0.51				
	8.20							
	8.22							
	8.18	8.13	0.046	0.57				
8.0(3)	8.09							
	8.12							
10.0(1)	10.10	10.12	0.049	0.48	10.17	101.7	0.073	0.72
	10.18							
	10.09							
10.0(2)	10.29	10.25	0.059	0.58				
	10.18							
	10.27							
10.0(3)	10.14	10.13	0.036	0.36				
	10.09							
	10.16							
12.0(1)	11.97	12.06	0.079	0.66	12.05	100.4	0.071	0.59
	12.12							
	12.09							
12.0(2)	12.02	12.01	0.031	0.26				
	11.98							
	12.04							
12.0(3)	11.97	12.07	0.100	0.82				
	12.08							
	12.17							

TABLE 2: PRECISION RESULTS OBTAINED FROM 10.0 mg mL⁻¹ STANDARD PCT AND PCR TEST SOLUTIONS CONTAINING 10.0 mgmL⁻¹ PCT (N=6)

System Precision				Repeata	bility	Intermediate precision	
Standard solutions	Results mg mL ⁻¹	RSD%	Test solution	Results mg mL ⁻¹	RSD%	Results mg mL ⁻¹	RSD%
St1	10.01	0.58	1	9.77	0.44	9.91	0.42
St2	10.08		2	9.73		9.91	
St3	10.09		3	9.82		9.87	
St4	10.04		4	9.83		9.82	
St5	10.06		5	9.84		9.90	
St6	10.18		6	9.83		9.94	

LOD and LOQ Values: LOD and LOQ were calculated using 3.3 sbl/m, and 10 sbl/m equations, respectively, where sbl is the standard deviation of the response of Plc solution accepted as blank solution and m is the slope of the calibration curve. The LOD and LOQ were found to be 2.2 and 7.3 mg mL⁻¹, respectively. All results obtained in the validation studies are presented in **Table 3**.

TABLE 3: METHOD-VALIDATION DATA FOR PCT DETERMINATION WITH RAMAN SPECTROSCOPY

Validation parameters	Obtained Results
Specificity	Specific
Linearity range	$7.0-13.0 \text{ mg mL}^{-1}$
LOD	$2.2~\mathrm{mg~mL^{-1}}$
LOQ	7.3 mg mL ⁻¹

Robustness: The robustness of an analytical procedure is a measure of its capacity to remain unaffected by small but deliberate variations in

method parameters and provides an indication of its reliability during normal usage. Some variations in the robustness parameters such as temperature, pH and the effects of these variations on the system suitability parameters and test results should be examined. In this study, the temperature of the analytical method was changed from 20 to 30 °C and the pH of the solutions was changed from 5.0 to 6.0. The obtained results were given in **Table 4**.

The difference between the results obtained with the original method and the modified method should be less than 2.0%. These results show that the proposed method is robust.

TABLE 4: RESULTS OBTAINED FROM THE ROBUSTNESS OF THE PROPOSED METHOD

		Temperature (°C	C)		pН	
Number of Samples	20	25	30	5.0	5.5	6.0
1 as mg mL ⁻¹	10.354	10.381	10.216	10.280	10.417	10.121
2 as mg mL ⁻¹	10.153	10.163	10.321	10.462	10.224	10.192
3 as mg mL ⁻¹	10.220	10.164	10.158	10.391	10.219	10.270
Mean as mg mL ⁻¹	10.242	10.236	10.232	10.378	10.287	10.194
SD	0.084	0.103	0.067	0.075	0.092	0.061
RSD%	0.816	1.002	0.659	0.722	0.896	0.597

Comparison of Method with HPLC: The results obtained from the Raman spectroscopic method was compared with a standard HPLC method to verify the proposed method for the PCT determination. The results obtained by the Student's t and F statistical tests of 73 samples are given in **Table 5**. The mean PCT concentration as mg.mL⁻¹ obtained by the proposed Raman spectroscopic and HPLC methods was found to be very close to the Prc solution for I.V. Infusion containing 10 mg mL⁻¹ PCT.

In the evaluation of t-test, the Null Hypothesis (H0, $(\alpha=0.05)$ indicates that there is no significant difference between the mean values, while the Alternative Hypothesis (H1, $(\alpha=0.05)$ says the opposite. If t critical is higher than texp, H0 is accepted, and H1 is rejected. According to the results presented in Table 5, the evaluation of the t critical with texp at 95% confidence level demonstrated that there was no significant difference between the Prc solution (accepted real value) and the experimentally found PCT concentrations. In the evaluation of the F test, the Null Hypothesis (H0, $(\alpha=0.05)$ says that there is no significant difference between the variances, while

the Alternative Hypothesis (H1, $(\alpha=0.05)$ says the opposite. If F critical is higher than Fexp, H0 is accepted, and H1 is rejected. According to the results presented in **Table 5**, there was no significant difference between the precisions (as standard deviations) of the proposed Raman spectroscopic and HPLC methods due to Fcritical > Fexp.

TABLE 5: COMPARISON OF THE STATISTICAL EVALUATION BETWEEN THE PROPOSED RAMAN SPECTROSCOPIC METHOD AND THE HPLC METHOD (N=73)

	HPLC	Raman
	Method	Spectroscopy
Mean as mg mL ⁻¹	10.04	10.08
Variance	0.032	0.026
$t_{\rm exp}$		1.31
t _{crtical}		1.98
F		1.25
F _{critical}		1.48

CONCLUSION: In this study, a simple, sensitive, selective, and fast method for the determination of PCT was developed and validated based on Raman spectroscopy. A pharmaceutical solution containing 10.0 mg mL⁻¹ of PCT was successfully quantified using the PLS models based on the Raman

PCT in Prc solution for I.V. Infusion.

Spectroscopy. The method demonstrated an acceptable accuracy, precision and stability over the applied concentration range with the linear concentration range from 7.0 to 13.0 mg mL⁻¹; the LOD and LOQ values were calculated as 2.2 and 7.3 mg mL⁻¹ respectively. All these results confirmed that the Raman Spectroscopic method was successfully validated for the determination of

ACKNOWLEDGEMENT: The authors thank Professor Yusuf Dilgin, Department of Chemistry, Faculty of Ars and Science, Çanakkale Onsekiz Mart University, Çanakkale, Turkey, for checking the scientific organization of this study. The authors are also thankful to Polifarmaİlaç San ve Tic. AŞ. Ergene/Tekirdağ, Turkey, for providing the facilities necessary to conduct the research.

CONFLICTS OF INTEREST: There are no conflicts of interest among all the authors about the publication of the manuscript.

REFERENCES:

- 1. Morse HN: Uebereineneue darstellungsmethode der acetylamidophenole[On a new method of preparing acetylamidophenol]. Berichte der Deutschen Chemischen Gesellschaft 1878; 11(1): 232–33.
- Thompson C, Davies MC, Roberts CJ, Tendler SJB and Wilkinson MJ: The effects of additives on the growth and morphology of paracetamol (acetaminophen) crystals. International Journal of Pharmaceutics 2004; 280: 137-50.
- Scottish Intercollegiate Guidelines Network (SIGN)
 Control of pain in adults with cancer. Scotland: National
 Health Service (NHS). ISBN 9781905813384. Archived
 (PDF) from the original on 2010-12-20.
- 4. Behera S, Ghanty S, Ahmad F, Santra S and Banarjee S: UV-Visible spectrophotometric method development and validation of determination of paracetamol tablet formulation. International Journal of Pharmaceutical Sciences and Research 2012; 3(12): 4945-53.
- El-Yazbi AF, Guirguis KM, Bedair MM and Belal TS: Validated specific HPLC-DAD method for simultaneous estimation of paracetamol and chlorzoxazone in the presence of five of their degradation products and toxic impurities. Drug Development and Industrial Pharmacy 2020; 46(11): 1853-61.
- Sridevi S, Vijayakumar R and Nalini CN: Method development and validation for the simultaneous estimation of ascorbic acid, phenylephrine HCl, paracetamol and levocetirizine HCl using RP-HPLC. Research Journal of Pharmaceutical Technology 2020; 13(4): 1911-16.
- Chefirat B, Zergui A, Nour Belmessabih M, Rahmani C, and Rezkkallah H: Validation of a spectrophotometric method for the determination of paracetamol in plasma applicable for toxicological emergencies in laboratories with limited resources. Toxicologie Analytique et Clinique 2020; 32(4): 266-77.

- E-ISSN: 0975-8232; P-ISSN: 2320-5148
- 8. Andronie L, Coroian A, OdagiuA and Bebu A: Characterization of pharmaceutical paracetamol tablets using Raman and Surface-Enhanced Raman Spectroscopy. ProEnvironment 2018; 11: 180-6.
- 9. Szostak R and Mazurek S: Quantification of active ingredients in suppositories by FT-Raman spectroscopy. Drug Testing and Analysis 2013; 5: 126–9.
- Tyszczuk-Rotko K, Jaworska I and Jędruchniewicz K: Application of unmodified boron-doped diamond electrode for determination of dopamine and paracetamol. Microchemical Journal 2019; 146: 664-72
- 11. Saleem SJ and Güler M: Electroanalytical determination of paracetamol using Pd nanoparticles deposited on carboxylated graphene oxide modified glassy carbon electrode. Electroanalysis 2019; 31: 2187-98.
- 12. Raposo F andIbelli-Bianco C: Performance parameters for analytical method validation: Controversies and discrepancies among numerous guidelines. TrACTrends in Analytical Chemistry 2020; 129: 1159132.
- Gumustas M, Kurbanoglu S, Uslu B and Ozkan SA: UPLC versus HPLC on Drug Analysis: Advantageous, Applications and Their Validation Parameters. Chromatographia 2013; 76: 1365-27.
- 14. Bakkar MA, Nawaz H, Majeed MI, Naseem A, Ditta A, Rashid N, Ali S, Bajwa J, Bashir S, Ahmad S, Hyat H, Bukhari KS and Bonnier F: Raman spectroscopy for the qualitative and quantitative analysis of solid dosage forms of sitagliptin. Spectrochimica Acta A: Molecular and Biomolecular Spectroscopy 2021;245:118900.
- 15. De Beer TRM, Baeyens WRG, Vermeire A, Broes D, Remon JP and Vervaet V: Raman spectroscopic method for the determination of medroxyprogesterone acetate in a pharmaceutical suspension: validation of quantifying abilities, uncertainty assessment and comparison with the high performance liquid chromatography reference method. Analytica Chimica Acta 2007;589:192–9.
- 16. De Bleye C, Dumont E, Rozet E, Sacre PY, Chavez PF, Netchacovitch L, Piel G, Hubert P and Ziemons E: Determination of 4-aminophenol in a pharmaceutical formulation using surface enhanced Raman scattering: From development to method validation. Talanta 2013; 116: 899–05.
- 17. Corredor CC, Vikstrom C, Persson A, Bu X and Both D: Development and robustness verification of an at-line transmission raman method for pharmaceutical tablets using quality by design (QbD) principles. Journal ofPharmaceutical Innovation 2018; 13: 287-00.
- Roggo Y, Chalus P, Maurer L, Martinez CL, Edmond A and Jent N: A review of near infrared spectroscopy and chemometrics in pharmaceutical technologies. Journal of Pharmaceutical and Biomedical Analysis 2007; 44: 683– 00.
- 19. Makki AA, Bonnier F, Respaud R, Chtara F, Tfayli A, Tauber C, Bertrand D, Byrne HJ, Mohammed E and Chourpa I: Qualitative and quantitative analysis of therapeutic solutions using Raman and infrared spectroscopy. SpectrochimicaActa Part A: Molecular and Biomolecular Spectroscopy 2019; 218: 97–08.
- 20. Casian T, ReznekA, VonicaGligor AL, Ranterghem JV, De Beer T and Tomuta I: Development, validation and comparison of near-infrared and Raman spectroscopic methods for fast characterization of tablets with amlodipine and valsartan. Talanta 2017; 167: 333–43.
- 21. Chellini PR, Mendes TO, Franco PHC, Porto BLS, Tippavajhala VK, César IC, Oliveira MAL and Pianetti GA: Simultaneous determination of rifampicin, isoniazid, pyrazinamide and ethambutol in 4-FDC tablet by Raman

E-ISSN: 0975-8232; P-ISSN: 2320-5148

- spectroscopy associated to chemometric approach. Vibrational Spectroscopy 2017; 90: 14–20.
- 22. Khandasammy SR, Fikiet MA, Mistek E, Ahmed Y, Halámková L, Bueno J and Lednev IK: Bloodstains, paintings, and drugs: Raman spectroscopy applications in forensic science. Forensic Chemistry 2018; 8: 111–33.
- Lia Y, IgnebB, Drennen JK and Anderson CA: Method development and validation for pharmaceutical tablets analysis using Transmission Raman Spectroscopy. International Journal of Pharmaceutics 2016; 498: 318–25.
- 24. Bajwa J, Nawaz H, Majeed MI, Hussain AI, Farooq S, Rashid N, Bakkar MA, Ahmad S, Hyat H, Bashir S, Ali, S and Kashif M: Quantitative analysis of solid dosage forms of cefixime using Raman Spectroscopy. Spectrochimica Acta A: Molecular and Biomolecular Spectroscopy 2020; 238: 118446.
- Jiang HY, Ding CH, Wang Y, Zhang YX, Mohammed A, Pan Y and Han B: Determination of acetaminophen spatial distribution and content in tablets using confocal microraman spectroscopy mapping. Journal of Nanoparticle Research 2020; 22: 265.
- 26. Makraduli L, MakreskiP, Goracinova K, Stefov S, Anevska M and Geskovski N: A Comparative approach to screen the capability of Raman and Infrared (Mid- and Near-) spectroscopy for quantification of low-active pharmaceutical ingredient content solid dosage forms: The case of alprazolam. Applied Spectro 2020; 74(6): 661–73.
- 27. Mazurek S and Szostak R: Quantification of active ingredients in pharmaceutical suspensions by FT Raman spectroscopy. Vibrational Spectroscopy 2017; 93: 57–64.
- 28. Shimamura R, Koide T, Hisada H, Inoue M, Fukami T, Kartori N and Goda Y: Pharmaceutical quantification with univariate analysis using transmission Raman Spectroscopy. Drug Development and Industrial Pharmacy 2019; 45(9): 1430-36.
- 29. Lim YI, Han J, Woo YA, Kim JJ and Kang MJ: Rapid quantitation of atorvastatin in process pharmaceutical powder sample using Raman Spectroscopy and evaluation of parameters related to accuracy of analysis. Spectrochimica Acta A: Molecular and Biomolecular Spectroscopy 2018; 200: 26–32.

- Mai Lêa LM, Berge M, Tfayli A, Zhou J, Prognona P, Baillet-Guffroy A and Caudron E: Rapid discrimination and quantification analysis of five antineoplastic drugs in aqueous solutions using Raman spectroscopy. European Journal of Pharmaceutical Sciences 2018; 111: 158–66
- 31. Da Silva de Jesus JIS, Löbenberg R and Bou-Chacra NA: Raman Spectroscopy for quantitative analysis in the pharmaceutical industry. Journal of Pharmacy and Pharmaceutical Science 2020; 23: 24-46.
- 32. Griffena JA, Owena AW and Matousek P: Quantifying low levels (<0.5% w/w) of warfarin sodium salts in oral solid dose forms using Transmission Raman spectroscopy. Journal of Pharmaceutical and Biomedical Analysis 2018; 155: 276–83.
- Borio VG, Vinha R, Nicolau RA, de Oliveira HPM, de lima CJ and Silveir L: Quantitative evaluation of acetaminophen in oral solutions By Dispersive Raman Spectroscopy for quality control. Spectroscopy: An International Journal 2012; 27(4): 215–28.
- Duraipandian S, Knopp MM, Pollard MR, Kerdoncuff H, Petersen JC and Müllertz A: Fast and novel internal calibration method for quantitative Raman measurements on aqueous solutions. Analytical Methods 2018; 10: 3589.
- 35. Mazurek S and Szostak R: Quantitative determination of acetylsalicylic acid and acetaminophen in tablets by FT-Raman spectroscopy. Analyst 2002; 127: 144–48.
- Kachrimanis K, Braun DE and Griesser UJ: Quantitative analysis of paracetamol polymorphs in powder mixtures by FT-Raman spectroscopy and PLS regression. Journal of Pharmaceutical and Biomedical Analysis 2007; 43: 407– 12
- ICH Harmonised Tripartite Guideline: Validation of Analytical Procedures: Text and Methodology, Q2(R1), ICH, Geneva, Switzerland, 2005, http://www.ich.org. internet
- 38. Ferenczi-Fodor K, Renger B and V'egh Z: The frustrated reviewer-recurrent failures in manuscripts describing validation of quantitative TLC/HPTLC procedures for analysis of pharmaceuticals. Journal of Planar Chromatography-Modern TLC 2010; 23(3): 173–79.

How to cite this article:

Kocaağa T, Yıldırım F, Kumrulu U, Kumrulu E, Mutlu N, Sopacı E, Demirdoğan B and Dilgin DG: Validation study for paracetamol determination in liquid products (paracerol) using raman spectroscopy. Int J Pharm Sci & Res 2022; 13(2): 755-62. doi: 10.13040/IJPSR. 0975-8232.13(2).755-62.

All © 2022 are reserved by the International Journal of Pharmaceutical Sciences and Research. This Journal licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported License.

This article can be downloaded to Android OS based mobile. Scan QR Code using Code/Bar Scanner from your mobile. (Scanners are available on Google Playstore)