



Received on 09 June 2021; received in revised form, 12 July 2021; accepted, 24 July 2021; published 01 March 2022

ANTIMICROBIAL AND ANTIFUNGAL ACTIVITY OF BARK OF *HARDWICKIA BINATA* ROXB (FABACEAE / CAESALPINIACEAE)

Md. Rageeb Md. Usman and Rohini Patil

Department of Pharmacognosy, Smt. Sharadchandrika Suresh Patil College of Pharmacy, Chopda - 425107, Maharashtra, India.

Keywords:

Antimicrobial activity, Antifungal activity, Bark, *Hardwickia binata* Roxb, Microorganisms.

Correspondence to Author:

Dr. Md. Rageeb Md. Usman

Department of Pharmacognosy,
Smt. Sharadchandrika Suresh Patil
College of Pharmacy, Chopda -
425107, Maharashtra, India.

E-mail: rageebshaikh@gmail.com

ABSTRACT: Background and Objectives: This study was carried out with the objective to investigate the antibacterial and antifungal activity of the bark of *Hardwickia binata* Roxb. **Materials and Methods:** Methanolic extract of bark of *Hardwickia binata* Roxb. exhibited an inhibitory effect towards the pathogenic organisms. Antimicrobial activity the zone of inhibition values of methanolic extract of bark of *Hardwickia binata* Roxb. in against two Gram-positive microorganisms viz. *Staphylococcus aureus* and *Bacillus subtilis* and two Gram negative microorganisms viz. *Escherichia coli* and *Pseudomonas aeruginosa* showed significant activity. Antifungal activity of methanolic extract of bark of *Hardwickia binata* Roxb. in against two microorganism, *Candida albicans*, and *Aspergillus niger*, showed significant activity. **Results:** The results showed that the remarkable inhibition of the bacterial growth was shown against the tested organisms. The phytochemical analyses of the plants were carried out. The microbial activity of the *Hardwickia binata* was due to the presence of various secondary metabolites. **Conclusion:** Hence, these plants can be used to discover bioactive natural products that may serve AS leads in the development of new pharmaceuticals research activities.

INTRODUCTION: Antibiotics are one of our most important weapons in fighting bacterial infections and have greatly benefited the health-related quality of human life since their introduction. However, over the past few decades, these health benefits are under threat AS many commonly used antibiotics have become less

effective against certain illnesses not only because many of them produce toxic reactions but also due to the emergence of drug-resistant bacteria. It is essential to investigate newer drugs with lesser resistance. Drugs derived from natural sources play a significant role in the prevention and treatment of human diseases.

In many developing countries, traditional medicine is one of the primary healthcare systems^{1, 4}. Herbs are widely exploited in traditional medicine, and their curative potentials are well documented^{5, 6}. About 61% of new drugs developed between 1981 and 2002 were based on natural products, and they have been very successful, especially in the areas

	QUICK RESPONSE CODE DOI: 10.13040/IJPSR.0975-8232.13(3).1189-93
	This article can be accessed online on www.ijpsr.com
DOI link: http://dx.doi.org/10.13040/IJPSR.0975-8232.13(3).1189-93	

of infectious disease and cancer^{7, 8}. Recent trends, however, show that the discovery rate of active novel chemical entities is declining⁹. Natural products of higher plants may give a new source of antimicrobial agents with possibly novel mechanisms of action^{10, 11}. The effects of plant extracts on bacteria have been studied by a very large number of researchers in different parts of the world¹². Much work has been done on ethnomedicinal plants in India¹³. Plants are rich in a wide variety of secondary metabolites such as tannins, terpenoids, alkaloids, flavonoids, glycosides, etc., which have been found *in-vitro* to have antimicrobial properties^{14, 15}.

The World Health Organization estimates that plant extracts or their active constituents are used as folk medicine in traditional therapies of 80% of the world's population¹⁶. The harmful microorganisms can be controlled with drugs, which results in the emergence of multiple drug-resistant bacteria. It has created alarming clinical situations in the treatment of infections. The pharmacological industries have produced a number of new antibiotics; resistance to these drugs by microorganisms has increased. In general, bacteria have the genetic ability to transmit and acquire resistance to synthetic drugs, which are utilized as therapeutic agents¹⁷. Therefore, there is need to search for new infection-fighting strategies to control microbial infections. *Hardwickia binata* is a deciduous, moderate to large-sized tree. The bark of saplings is almost silvery-white and smooth. Leaves are alternate, bifoliolate resembling Bauhinias. Leaflets are almost kidney-shaped and grayish-green in color, sessile, entire, obliquely ovate, and coriaceous testa^{18, 19}.

Previous researchers worked out tannins from the bark to produce medicines for the treatment of diarrhea, worms, indigestion, and leprosy and produce an appetizer²⁰. The leaves, pods, and bark contain tannins²¹. Bark used for gums and resins²². The leaves are used for fodder and manure²³. The leaves extract showed activity against both gram-positive and gram-negative bacteria and fungi. Bioactive substances showed antimicrobial agents for treating various bacterial and fungal infections, including gonorrhoea, pneumonia, eye infections, mycotic infections, and tannins traditionally used to protect catarrh, wounds, and

hemorrhoids, diarrhea, and antidote in heavy metal poisoning. Flavonoids possess anti-inflammatory, anti-allergic, antithrombotic, antimicrobial and vaso-protective effects. The leaves are used for headache and treatment of constipation²⁴. Therefore, the present study aims to evaluate the antimicrobial activity and antifungal activity of *Hardwickia binata* Roxb and the literature survey reveals that no reports were found on the antimicrobial and antifungal activity of the bark extracts of *Hardwickia binata* Roxb.

MATERIAL AND METHOD:

Plant Material: The plant *Hardwickia binata* Roxb. is widely found throughout India. For my work the plant was collected from in the deep forest of Satpuda hills with the help of forest officers of Chopda Tahsil, Dist. Jalgaon, Maharashtra (India) and authenticated by Prof. (Dr.) Priyanka A. Ingle, scientist, BSI (Botanical Survey of India), Pune (M.S.). The stems of the plant were dried under shade and then coarsely powdered with the help of a mechanical grinder. The powder was passed through sieve no. 40 and stored in an airtight container for further studies. Extraction was carried out by a continuous soxhlet extraction process for 72 h.

Tests Microorganisms for Antimicrobial Activity:

Gram-Positive: Clostridium tetani, Bacillus subtilis.

Gram-Negative: Salmonella typhi, Pseudomonas aeruginosa.

Test Microorganism for Antifungal Activity: Candida albicans, Aspergillus niger.

Anti Microbial and Antifungal Activity:

Preparation and Standardization of Stock Culture: From the cultures maintained on nutrient agar slants, one loop full of the respective organisms was taken and aseptically transferred to 100 ml of sterile nutrient broth in a flask that was shaken thoroughly and incubated at 37°C for 24 h. One ml of this seeded broth was then diluted with 9 ml of sterile water in a culture tube. This was shaken thoroughly, and about 1ml of this suspension was transferred to a second culture tube, which in addition to 9 ml of sterile water. This was

shaken thoroughly and further diluted 10 times with sterile water till 1010 dilution was obtained (up to 10 culture tube). Incubating 0.2 ml of each dilution on solidified nutrient agar medium by spread plate method did standardization of the seeded broth. After incubation at 370 °C for 48 h, the number of wells formed colonies on the plate was counted. The seeded broth was then suitably diluted to contain between 107 to 108 microorganisms c.f.u./ml (colony forming unit per ml). This was designated as the working stock that was used for antimicrobial studies.

Preparation of Test Solution: The test solution of methanol extract of bark of *Hardwickia binata* Roxb. was prepared by dissolving 1 gm of dried methanol extract of bark of *Hardwickia binata* Roxb. in 1 ml of solvent and the filter paper soaked in respective solvent was used as control.

Procedure: Both antibacterial and antifungal activity of methanol extract was screened by filter paper disc method. A previously liquefied Muller Hinton Agar media was inoculated with the requisite quantity of the microorganism's suspension. The suspension was added to the medium at a temperature between 400-500C and the inoculated medium was poured immediately into dried petri dishes to occupy a depth of 3 to 4 mm. The paper disc (No.2 Whatman) was cut down into a small disc (6mm in diameter) and sterilized at 1800C/30' m in a hot air oven and then impregnated with the test solution, standard solution, and paper disc soaked with solvent as control. The dried discs were placed on the surface of the medium.

The dishes were left standing for 1-4 h at room temperature as a period of pre-incubation diffusion to minimize the effects of variation in time between the applications of different solutions. Subsequently incubated for about 18 h at about 370 °C, and the diameter of the circular inhibition zones was measured²⁵.

RESULTS AND DISCUSSION: Methanol extract of the bark of *Hardwickia binata* Roxb exhibited an inhibitory effect on pathogenic organisms. The antimicrobial activity of *Hardwickia binata* extract was evaluated against Gram-negative *Escherichia coli* & *Pseudomonas aeruginosa* and Gram-

positive bacteria namely, *Staphylococcus aureus* & *Bacillus subtilis* by disc method. The zone of inhibition of synthesized HB extracts good antimicrobial activity against gram negative bacteria such as *Escherichia coli* & *Pseudomonas aeruginosa* with zone of inhibition 8.3 mm & 7.9 mm and 9.1 mm & 8.8 mm respectively. *Hardwickia binata* Extract is most effective against both the fungi, i.e., *Candida albicans* and *Aspergillus niger*. The results are depicted in **Table 1, 2,** and **Fig. 1, 2.** Recently, much attention has been directed towards extract and biologically active compounds from popular plant species. The use of medicinal plants plays a vital role in covering the basic health needs in developing countries, and these plants may offer a new source of antibacterial agents with significant activity against infective microorganisms. Our results concerning the methanol extract of bark of *Hardwickia binata* Roxb. showed a significant antibacterial and antifungal activity against both Gram-positive and Gram-negative bacteria.

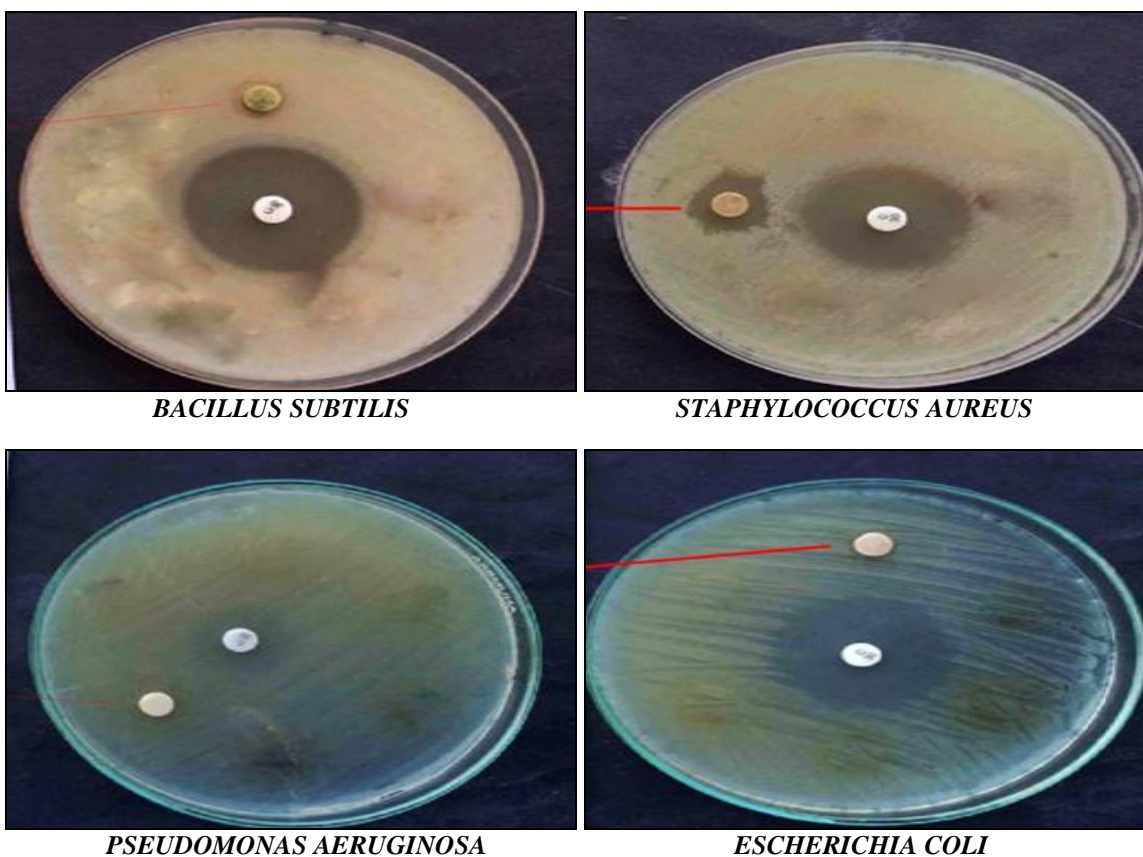
TABLE 1: MICROBIAL CULTURE AND COLLECTION CENTRE

S. no	Name of culture	Culture collection centre
Bacteria		
1	<i>Pseudomonas aeruginosa</i> (NCIM 2036)	NCIM, Pune
2	<i>Escherichia coli</i> (NCIM 2109)	NCIM, Pune
3	<i>Staphylococcus aureus</i>	NCIM, Pune
4	<i>Bacillus subtilis</i>	NCIM, Pune
Fungi		
1	<i>Candida albicans</i>	NCIM, Pune
2	<i>Aspergillusniger</i>	NCIM, Pune

TABLE 2: ANTIMICROBIAL ACTIVITY AND ANTI-FUNGAL ACTIVITY

Microorganisms	Diameter of zone of inhibition(mm)	
	Extract	Standard
Gram-Negative bacteria		
<i>Pseudomonas aeruginosa</i>	8.3	12.46
<i>Escherichia coli</i>	7.9	29.12
Gram-Positive bacteria		
<i>Staphylococcus aureus</i>	9.1	15.32
<i>Bacillus subtilis</i>	8.8	18.02
Fungi		
<i>Candida albicans</i>	4.6	11.59
<i>Aspergillus niger</i>	5.8	12.10

Diameter in mm calculated by Vernier Caliper; '-' means no zone of inhibition; Well diameter - 6 mm; NCIM-National Collection of Industrial Micro-organisms; Standard-Chloramphenicol



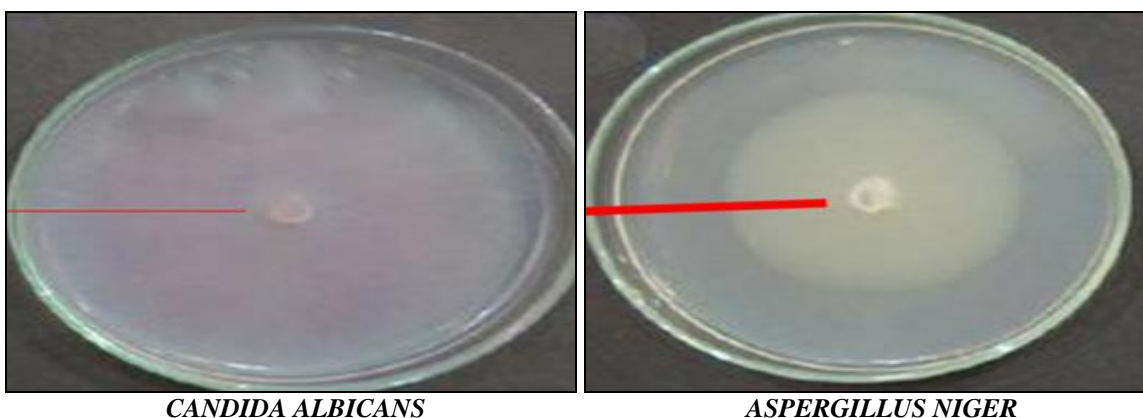
BACILLUS SUBTILIS

STAPHYLOCOCCUS AUREUS

PSEUDOMONAS AERUGINOSA

ESCHERICHIA COLI

FIG. 1: ANTIMICROBIAL ACTIVITY OF METHANOL EXTRACT OF BARK OF *HARDWICKIA BINATA* ROXB



CANDIDA ALBICANS

ASPERGILLUS NIGER

FIG. 2: ANTIFUNGAL ACTIVITY OF METHANOL EXTRACT OF BARK OF *HARDWICKIA BINATA* ROXB

CONCLUSION: The present study justified the claimed uses of barks in the traditional system of medicine to treat various infectious diseases caused by microbes. However, further studies are needed to better evaluate the potential effectiveness of the crude extracts as antimicrobial agents.

The present results will form the basis for the selection of plant species for further investigation in the potential discovery of new natural bioactive compounds. Further studies aimed at the isolation and structure elucidation of antibacterial active constituents from the plant have been initiated.

ACKNOWLEDGMENT: The authors are grateful thanks to the Management and Principal of Smt. Sharad chandrika Suresh Patil College of Pharmacy, Chopda, Maharashtra, for providing timely support for the research work.

CONFLICTS OF INTEREST: The authors declare no conflicts of interest.

REFERENCES:

1. Sparsha Pattnaik, Chandi Charan Rath and Lakshmi Singh: Isolation, identification and screening for bioactive compounds with antimicrobial activities from sub-aerial cyanobacteria of eastern region, odisha. International

- Journal of Pharmaceutical Science and Research 2021; 12(7): 3716-30.
2. Haque M, Sartelli M, McKimm J and Abu Bakar M: Health care-associated infections - an overview. *Infect Drug Resist* 2018; 11: 2321-33.
 3. Negut I, Grumezescu V and Grumezescu AM: Treatment strategies for infected wounds. *Mol* 2018; 23(9): 2392.
 4. Rahim K, Saleha S, Zhu X, Huo L, Basit A and Franco OL: Bacterial contribution in chronicity of wounds. *Microb Ecology* 2017; 73: 710-21.
 5. Rai S, Yadav UN, Pant ND, Yakha JK, Tripathi PP, Poudel A and Lekhak B: Bacteriological profile and antimicrobial susceptibility patterns of bacteria isolated from pus/wound swab samples from children attending a tertiary care hospital in Kathmandu, Nepal. *International Journal of Microbiology* 2017; 5.
 6. Landén NX, Li D and Ståhle M: Transition from inflammation to proliferation: a critical step during wound healing. *Cell Mol Life Sci* 2016; 73(20): 3861-85.
 7. Cherkupally R, Kota SR, Amballa H and Reddy BN: *In-vitro* antifungal potential of plant extracts against *Fusarium oxysporum*, *Rhizoctonia solani* and *Macrophomina phaseolina*. *Annals of Plant Sciences* 2017; 6(9): 1676-80.
 8. Sarker SD and Nahar L: Chemistry for pharmacy students: general, organic and natural product chemistry. John Wiley Sons Inc 2013; 1-383.
 9. Prasannabalaji N, Muraltharan G, Sivanandan RN, Kumaran S and Pugazhvendan SR: Antibacterial activity of some Indian traditional plant extract. *Asian Pacific Journal of Tropical Diseases* 2012; 5293.
 10. Hugo WB and Russel AD: *Pharmaceutical microbiology*, 6th edition Blackwell Science, Ltd. Osney Mead Oxford OX2 25 John Street London WC1N6AJ 2015.
 11. Akinpelu DA, Alayande KA, Aiyegoro OA, Akinpelu OF and Okoh AI: Probable mechanisms of biocidal action of *Cocos nucifera* Husk extract and fractions on bacterial isolates. *Journal of Bio Med Central Complementary and Alternative Medicine* 2015; 15: 116.
 12. Reddy PS, Jamil K and Madhusudhan P: Antibacterial activity of isolates from *Piper longum* and *Taxus baccata*. *Pharmaceutical Biol* 2001; 39: 236-8.
 13. Mugilan V and Sivakami R: Antimicrobial activity of microalgae isolated from fresh water pond, Tamil Nadu, India. *International Journal of Current Microbiology and Applied Sciences* 2016; 5(6): 588-95.
 14. Babu RM, Malathi T and Rao BD: Antifungal activity of selected Cyanobacteria against fungal pathogens. *Int J of Pharma and Biosciences* 2017; 7(4): 207-13.
 15. Safari M, Nowruzi B, Estalaki S and Shokri M: Biological activity of methanol extract from *Nostoc* sp. N42 and *Fischerella* sp. S29 isolated from aquatic and terrestrial ecosystems. *International Journal on Algae* 2019; 21(4): 373-91.
 16. Shaik D, Malika FA, Rafi SM, Naqui B: Studies of antibacterial activity of ethanolic extract from *Nericum indicum* and *Hibiscus rosasinensis*. *J Islamic Acad Sci* 1994; 7: 167-8.
 17. Towers GH, Lopez A, Hudson JB: Antiviral and antimicrobial activities of medicinal plants. *J Ethnopharmacol* 2001; 77: 189-96.
 18. Gunaselvi G, Kulasekaren V, Magalakshmi G and Gopal V: Preliminary phytochemical studies on the leaves of *Hardwickia binata* Roxb. (Fabaceae). *International Journal of Pharmacy and Biomedical Sciences* 2010; 1(3): 37-40.
 19. Maitreyee K: *Hardwickia binata* Roxb. Seed leaflet 2011; 152: 1-2.
 20. Ranganathan R, Vijayalakshmi R and Parameswari P: Ethnomedicinal survey of jawadhu hills in Tamil Nadu. *Asian Journal of Pharmaceutical and Clinical Research* 2012; 5(2): 45-49.
 21. Masilamani P, Singh BG, Chinnusamy C and Annadurai K: Influence of seed orientation and depth of sowing on germination and vigour of Anjan (*Hardwickia binata* Roxb). *Tropical Agricultural Research and Extension* 1999; 2(1): 76-78.
 22. Basha SKM, Umamaheswari P, Rambabu M and Savitramma N: Ethnobotanical study of mamandur forest (kadapa-nallamali range) in Eastern Ghats. *Journal of Phytology* 2011; 3(10): 44-47.
 23. Chand S and Singh AK: Direct somatic embryogenesis from zygotic embryos of a timber-yielding leguminous tree, *Hardwickia binata* Roxb. *Current Science* 2001; 80(7): 882-88.
 24. Gunaselvi G, Kulasekaren V and Gopal V: Anti bacterial and antifungal activity of various leaves extracts of *Hardwickia binata* Roxb. (Caesalpiniaceae). *International Journal of Pharm Tech Research* 2010; 2(4): 2183-87.
 25. Lorenzi H and Matos F A: Plantas medicinais nativas no Brasil: Nativase Exóticas. Instituto Plantarum Nova Odessa São Paulo Brasil 2008; 544.

How to cite this article:

Rageeb M Usman M and Patil: R Antimicrobial and antifungal activity of bark of *hardwickia binata* roxb (fabaceae / caesalpiniaceae). *Int J Pharm Sci & Res* 2022; 13(3): 1189-93. doi: 10.13040/IJPSR.0975-8232.13(3).1189-93.

All © 2022 are reserved by International Journal of Pharmaceutical Sciences and Research. This Journal licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported License.

This article can be downloaded to **Android OS** based mobile. Scan QR Code using Code/Bar Scanner from your mobile. (Scanners are available on Google Playstore)