IJPSR (2022), Volume 13, Issue 8



(Research Article)

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Received on 30 November 2021; received in revised form, 12 January 2022; accepted, 28 April 2022; published 01 August 2022

EVALUATION OF SEEDS OF AN INDIAN SACRED PLANT *AEGLE MARMELOS*, FOR THEIR ANTIOXIDANT AND CYTOTOXIC POTENTIAL

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Keywords:

Aegle marmelos seeds, Antioxidant activity, Cytotoxic activity, Flavonoids

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ABSTRACT: Introduction: The seeds of *Aegle marmelos* (bael) are less explored for their therapeutic activity; cytotoxic activity of bael seeds was yet to be studied. The present study aims at finding the cytotoxic activity of the successive extracts and the flavonoid fraction from the methanolic extract of the seeds. Methods: The extracts were prepared using solvents (petroleum ether, chloroform, ethyl acetate, methanol, and water) in increasing order of their polarity. Anti-oxidant activity of the extracts was evaluated using DPPH radical scavenging assay, nitric oxide scavenging assay, and hydrogen peroxide scavenging assay. Flavonoid fractions were obtained by thin-layer chromatography of the flavonoid-rich fraction prepared from the total methanolic extract. Human colon cancer cell line (HT 29), human breast cancer cell line (MCF-7), human lung cancer cell line (A-549) were used to evaluate the cytotoxicity of the extracts and the flavonoid fractions. Results: Various phytochemicals like alkaloids, flavonoids, tannins and phenols, steroids, and saponins were detected in phytochemical investigation of the extract. Ethyl acetate fraction showed comparable DPPH radical scavenging activity and hydrogen peroxide scavenging activity with ascorbic acid (standard). Methanol extract showed good nitric oxide scavenging activity of all the extracts. Flavonoid fraction F-01 showed 50% cell growth inhibition at concentration $<10 \ \mu g/ml$ on the human breast cancer cell line (MCF-7), which indicates its potent activity. Conclusion: F-01, one of the flavonoid fractions, showed comparable activity with the standard (Adriamycin) on MCF-7 cell line. Anti-oxidant study was conducted for the extracts, and the extracts showed good activity comparable to the standards used.

INTRODUCTION: Traditional medicine is well known and used for ages due to less side effects after its administration. One such medicinal plant used in India since ancient times is *Aegle marmelos*.

	DOI: 10.13040/IJPSR.0975-8232.13(8).3261-74		
	This article can be accessed online on www.ijpsr.com		
DOI link: http://dx.doi.org/10.13040/IJPSR.0975-8232.13(8).3261-74			

Aegle marmelos, commonly known as Bael belong to the Rutaceae family. The plant is precious to the Hindu god Lord Shiva, and its leaves are offered as offering to Lord Shiva. It is the most ancient documented plant in India, according to Charak $(1500 \text{ BC})^{-1}$.

Charak Samhita, prominent ayurvedic literature, describes bael as an essential ingredient in the ayurvedic system of medicine¹. Although native to Northern India, it is widely distributed throughout the Indian subcontinent, Sri Lanka, Pakistan, Bangladesh, Burma, Thailand and most southeastern Asian countries 2 . It is a mid-sized tree that grows up to 15m tall. Every part of the tree, *i.e.*, leaf stem bark, root, fruit and seeds, have medicinal values.

Bael is used as a carminative, laxative, astringent and stomachic and also treats conditions like fever, diarrhea, vomiting, dysentery, diabetes, asthma, inflammation, *etc*.

Unripe bael fruits are bitter, astringent, and haveanti-laxative properties, improve digestion and treat diarrhea. Ripe fruits are sweet, aromatic, and astringent with laxative properties ^{3, 4}. *Aegle marmelos* are known by various names in different Indian languages, some listed in **Table 2**.

There are marketed products of bael, like jam, chyawanprash, squash, bael powder, capsules, syrup and tablets ⁵. **Table 1** provides the taxonomic classification of *Aegle marmelos*.

TABLE 1: TAXONOMIC CLASS OF AEGLE MARMELOS⁶

Taxonomic Classification				
Kingdom	Plantae			
Subkingdom	Tracheobionta			
Superdivision	Spermatophyta			
Division	Magnoliophyta			
Class	Magnoliopsida			
Subclass	Rosidae			
Order	Sapindales			
Family	Rutaceae			
Genus	Aegle Corr. Serr.			
Species	Aegle marmelos (L.) Corr. Serr.			

TABLE 2: NAMES OF AEGLE MARMELOS INVARIOUS INDIAN LANGUAGES 7

Language	Vernacular name
Hindi, Marathi	Bel, Belgiri and Bili
Gujarati	Bilavaphal
Urdu	Belk ham
Oriya	Belo
Tamil	Vilva marum or Vilvama
Sanskrit	Shivaphala
Malayalam	Marredy

TABLE 3. LITERATURE REVIEW	OF	VARIOUS PARTS OF A MARMELOS
TABLE 5. LITERATORE REVIEW	Or	VARIOUS I ARIS OF A. MARMELOS

Extract/Active Compound	Activity	Animal model /Cell lines	Year	References	
	Le	af			
Leaf juice	Hypoglycemic activity	Patients	2021	8	
Successive leaf extracts	Anticancer activity	MDA-MB-231, Vero cells, HEp-2	2019	9	
Methanolic and aqueous leaf		In-vitro antioxidant tests	2019	10	
extracts	Antioxidant activity				
Methanolic leaf and fruit	Antihepatotoxic activity		2018	11	
extract					
Sequential extraction	Antibacterial, Antifungal,	Antibacterial, antifungal activity-	2017	12	
Chloroform, ethyl acetate,	Antioxidant and Cytotoxic	Various strains of bacteria and			
acetone, methanol; Aqueous	Properties	fungi Cytotoxicity-MTT assay			
extract	-	using T47D cell line			
Ethanolic extract of leaf and	Anti-inflammatory and	Wistar albino rats	2017	13	
fruit pulp	analgesic activity				
Ethanolic leaf extract	Against cognitive and	Albino Wistar rats	2019	14	
	neurological disorder				
Fruit					
Ethanolic and aqueous extract	Anti-microbial		2018	15	
Aqueous Fruit extract	Antioxidant, anticancer,	In vitro anti-oxidant tests,	2018	16	
	antibacterial activity	Bacterial strains,			
		MCF-7 cell line			
	See	ed			
Aqueous seed extract	Hypoglycemic and	Male albino wistar rats	2006	17	
	antihyperglycemic activity				
1-Methyl-2-(3'-methyl-but2'-	Antifungal activity	Aspergillus fumigatus,	2010	18	
enyloxy)-anthraquinone from		Aspergillus flavus, Aspergillus			
methanolic extract of seed		niger and Candida albicans			
Methanolic seed extract	Anti-inflammatory activity	Male Wistar rats	2011	19	

Literature survey **Table 3** reports various activities of different parts of *Aegle marmelos*. Leaf, fruit, bark and seeds are evaluated for their activities using various solvents. Therapeutic activities like antidiabetic, antifungal, antimicrobial, and antiinflammatory activity of seeds are reported in the literature. Preclinical studies have shown that bael leaf extracts effectively inhibit the growth of leukemic K562, T-lymphoid, B-lymphoid, erythroleukemic HEL, melanoma Colo 38 and breast cancer cell lines MCF-7 and MDA-MB-231. Seeds are evaluated for their anti-inflammatory, hypoglycemic and anti-hyperglycemic, antidiabetic and antifungal activities. There are no reports of studies on seeds for cytotoxic potential to date as explored on databases like pubmed, medscape, google scholar, and a gray area. Present research work aims on finding the cytotoxicity of the extracts and the flavonoid fractions.

EXPERIMENTAL:

Collection and Authentication of Plant Material: *Aegle marmelos* (Bael) seeds were procured from Greenfield Agro forestry products, Madhya Pradesh, India. Procured seeds were authenticated from Blatter Herbarium, St. Xavier's College, Mumbai, India. It was found that the specimen matched with Blatter Herbarium specimen number Bole-21 P. V. Bole and hence authenticated and certified that the specimen was of *Aegle marmelos*.

Preparation of Extracts: The seeds were shade dried and ground into a coarse powder. 80 gm of seeds was defatted using petroleum ether. The powder was then successively extracted using the Soxhlet apparatus using solvents to increase polarity: petroleum ether, chloroform, ethyl acetate, methanol, and distilled water.

Qualitative Phytochemical Investigation: The plant extracts were examined for the presence of alkaloids, flavonoids, tannins and phenols, glycosides, saponins, and steroids as per standard protocols 20 .

Antioxidant Studies:

DPPH Radical Scavenging Activity: The DPPH radical scavenging activity of the extract was determined by the method reported by Awah *et al.*²¹ 2ml of different concentrations (20, 40, 60, 80,100 ppm) of the extracts were prepared and added to 1ml of 0.3 mM DPPH prepared in ethanol. The resulted solution was agitated and left to stand for 25 min at room temperature. 2ml of test solutions and 1 ml of ethanol were mixed to prepare blank solutions and 1 ml of 0.3mM DPPH solution and 2 ml of ethanol were mixed to prepare negative control. L-Ascorbic acid was used as the positive control.

The absorbance of the solutions was measured at 518 nm against each blank using a UV Vis Diode Array spectrophotometer. The assay was done in triplicate. The DPPH radical scavenging activity was calculated using the following equation:

Scavenging activity (%) = A _{control}- A _{test} / A _{control} \times 100

Nitric Oxide Radical Scavenging Assay: The activity of the extracts was determined by the method reported by Boora et al. with some modifications²². 1000 ppm stock solutions of the extracts were prepared in ethanol. The extracts were diluted to prepare concentrations ranging from 20-100 ppm (20, 40, 60, 80, 100 ppm). Gallic acid was used as standard. To prepare the Griess reagent, volumes of 0.1% N-(1-naphthyl) equal ethylenediamine dihydrochloride (NEDD) and 1% sulphanilamide in 2.5% phosphoric acid were mixed and used immediately.

10mM sodium nitroprusside solution was prepared in phosphate-buffered saline and 0.5 ml of this solution was mixed with 1ml of different concentrations of the extracts and incubated for 180 mins at 25°C. The incubated solutions were mixed with equal proportions of freshly prepared Griess reagent. Control samples were prepared similarly but without the extracts. The absorbance of the samples was measured at 546nm using a UV Vis Diode Array spectrophotometer. The assay was done in triplicate. Percent inhibition of the extracts and the standard was calculated using following formula:

Nitric oxide Scavenged (%) = A _{control} - A _{test} / A _{control} $\times 100$

Hydrogen Peroxide Scavenging Assay: The scavenging activity of the extracts toward hydrogen peroxide radicals was determined by the method by Ngonda *et al.* ²³ 40mM solutions of hydrogen peroxide was prepared in phosphate buffer (ph 7.4). Extracts of different concentrations (50, 100, 150, 200, 250, 300 ppm) were prepared. 2ml of hydrogen peroxide was added to 1 ml of extracts and incubated for 30min. Absorbance was recorded at 560 nm. Ascorbic acid was used as the standard. The assay was done in triplicate. Percent scavenging activity was calculated using the following formula:

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\begin{array}{l} \mbox{Percent scavenged } (H_2O_2) = 1 \mbox{ - Absorbance (standard)} \, / \\ \mbox{ Absorbance (control)} \times 100 \end{array}
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Total Flavonoid Content of Methanolic Extract: Total flavonoid content of the methanolic extract was determined by the method by Chang *et al.* with slight modifications 24 . Quercetin was used to obtain a calibration curve. A 1000 ppm stock solution of quercetin was prepared in 80% ethanol. Dilutions were made to prepare 6.25, 12.5, 25, 50, 80, 100 ppm solutions from the stock solution. Each 0.5ml of diluted sample was mixed individually with 1.5 ml of 95% methanol, 0.1 ml of AlCl3 (aluminum chloride), 0.1 ml of 1M CH₃CO₂K (potassium acetate), and 2.8 ml of distilled water. The samples were incubated for 30mins at room temperature. The absorbance of the sample solutions was measured at 415 nm using UV spectrophotometer. For the blank solution, aluminium chloride was replaced with the same amount of distilled water. On similar lines, 0.5 ml of methanolic extract was made to react with aluminium chloride to estimate total flavonoid content.

$$\mathbf{C} = (\mathbf{c} \mathbf{x} \mathbf{V}) / \mathbf{m}$$

C = Flavonoid content (QE/g) c = Concentration of quercetin from calibration curve (mg/mL) V= Volume of extract (mL) m = weight of plant extract (g)

Preparation of Flavonoid Rich Fraction (FRF): Total methanolic extract was prepared. The extract was suspended in distilled water. The resulting aqueous solution was partitioned with petroleum ether using a separating funnel.

The petroleum ether fraction was discarded the aqueous fraction was further separated with ethyl acetate. The aqueous fraction was discarded, and the ethyl acetate fraction was concentrated to obtain the flavonoid-rich fraction. The percent extractive yield of the flavonoid-rich fraction was determined ²⁵.

Thin Layer Chromatography of FRF: Thin layer chromatography of FRF was performed using the mobile phase Ethyl acetate: Formic acid: Glacial acetic acid: Water: 10: 0.5: 0.5: 1.5. The obtained

bands (silica) were scraped off from the TLC plate. They were suspended in ethyl acetate.

The suspension was filtered using a Whatman filter paper. The filtered fractions were concentrated in a water bath.

LC-MS Analysis of Isolated Flavonoid Fractions: The isolated flavonoid fractions (Band 1, Band 2, Band 3, Band 4) 10mg(approx.) were subjected to LC-MS analysis at IIT SAIF, Mumbai-400076 (LC-MS; Agilent technologies; ES-API scan).

In-vitro Cytotoxic Studies: The plant extracts were evaporated to dryness under a rotary evaporator. The concentrated extracts, the flavonoid-rich extract, and the isolated flavonoid fractions were subjected to *in-vitro* cytotoxicity studies on MCF-7, HT-29 and A-549 cell lines at ATREC (The Advanced Centre for Treatment, Research and Education in Cancer).

RESULTS:

Preparation of Extracts: The extracts were prepared successively. The excess solvents were recovered using the Soxhlet apparatus. The extract was concentrated using a water bath. **Table 4** shows the percent yield of the extracts obtained.

TABLE 4:	PERCENT	YIELD	OF	THE	EXTRACTS
OBTAINED	USING VAL	RIOUS S	OLV	ENTS	

Solvents	% Yield
Petroleum ether	35.66%
Chloroform	1.25%
Ethyl acetate	0.225%
Methanol	6.36%
Water	0.58%
Total Methanolic extract	2.9%

Qualitative Phytochemical Investigation: Phytochemical investigation of the extracts was performed qualitatively using standard procedures in which chloroform extract shows the presence of the alkaloids, flavonoids, tannins and phenols, saponins. Ethyl acetate extract showed the presence of flavonoids, tannins, phenols and saponins.

TABLE 5: PHYTOCHEMICAL INVESTIGATION OF THE SUCCESSIVE EXTRACTS

IIIDEE CITIIII I O OIIEM		inon or im	вессыры	IT B BHIILIC		
Solvents	Petroleum Ether	Chloroform	Ethyl	Methanol	Water	Total Methanolic
			Acetate			Extract
Alkaloids	-	+	-	-	-	-
Flavonoids	-	+	+	+	-	+
Tannins and phenols	-	+	+	+	-	+
Glycosides	-	-	-	-	-	-

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Steroids	+	-	-	+	-	
Saponins	-	+	+	+	+	+

Present: + Absent: -

Methanol extract showed positive tests for flavonoids, tannins and phenols, saponins and steroids. Total methanolic extract showed the presence of flavonoids, tannins and phenols, and saponins. Petroleum showed the presence of steroids. **Table 5** summarizes the results of phytochemical tests.

Antioxidant Studies:

DPPH Assay: DPPH assay was performed, ethyl acetate extract showed better DPPH radical scavenging activity of all the extracts (IC_{50} = 27.2 µg/ml) comparable with that of the standard ascorbic acid (IC_{50} = 22.5 µg/ml). **Table 6** gives the IC_{50} values of the extracts and the standard ascorbic acid. **Fig. 1** shows a graphical representation of the DPPH radical scavenging activity of the extracts and the standard.

TABLE 6: DPPH RADICAL SCAVENGING ACTIVITYOF STANDARD AND EXTRACTS

Extract / Control	IC_{50} (µg/ml)
Ascorbic acid	22.5
Petroleum ether	49.6
Chloroform	34.2
Ethyl acetate	27.2
Methanol	30.1
Distilled water	45.4



FIG. 1: DPPH RADICAL SCAVENGING ACTIVITY

Nitic Oxide Scavenging Assay: Methanolic extract scavenged more nitric oxide (IC₅₀ =22.6 μ g/ml) as compared to other extracts and showed comparable activity with the standard gallic acid (IC₅₀ = 20.8 μ g/ml). IC₅₀ values of extracts and the standard for nitric oxide scavenging activity is given in Table 7, and a graphical representation of the same is shown in Fig. 2.

TABLE 7: NITRIC OXIDE SCAVENGING ACTIVITYOF STANDARD AND EXTRACTS

Extract	IC ₅₀ (μg/ml)
Gallic acid	20.8
Petroleum ether	45.3
Chloroform	30.4
Ethyl acetate	24.2
Methanol	22.6
Distilled water	40.8



FIG. 2: NITRIC OXIDE SCAVENGING ACTIVITY

Hydrogen Peroxide Scavenging Assay: Hydrogen peroxide scavenging activity was shown best by ethyl acetate (IC_{50} = 180.5 µg/ml) compared to other extracts. The IC_{50} value of the standard ascorbic acid was found to be 150.3µg/ml. Table 8 gives the hydrogen peroxide scavenging assay result, and Fig. 3 gives its graphical representation.

TABLE 8: HYDROGEN PEROXIDE SCAVENGINGACTIVITY OF STANDARD AND EXTRACTS

Extract	IC ₅₀ (μg/ml)
Ascorbic acid	150.3
Petroleum ether	250.5
Chloroform	208.2
Ethyl acetate	180.5
Methanol	190.3
Distilled water	200.2



FIG. 3: HYDROGEN PEROXIDE SCAVENGING ACTIVITY

Total Flavonoid Content: Flavonoid content of methanolic extract by aluminium chloride method was found to be 12.933 mg of QE/g. **Fig. 4** shows the calibration curve of quercetin. The total flavonoid content was calculated to be 12.933 mg of QE/g.



FIG. 4: CALIBRATION CURVE OF QUERCETIN

Preparation of Flavonoid Rich Fraction: The percent yield of the flavonoid-rich fraction was 14% w/w.

Thin Layer Chromatography of FRF: Four distinct fluorescent bands were obtained on the TLC plate under long UV exposure. Rf of these bands were 0.129 (Band 1), 0.17 (Band 2), 0.612 (Band 3), and 0.79 (Band 4). Fig. 5 shows the thin layer chromatography plate under long UV exposure showing 4 distinct flavonoid bands.



LC-MS Analysis of Isolated Flavonoid Fractions:

The LC-MS analysis of the fractions showed the presence of the following compounds:

Fraction 01: Fig. 6-10.

- Annofoline
- ✤ Scopoletin
- ✤ Agyrolobine
- ✤ Luvangetin
- Fencamfamine



FIG. 7: MASS SPECTRUM OF SCOPOLETIN





- ➢ L-Galactose
- ➢ 11-amino-undecanoic acid

- ➢ 6-Hydroxyangelicin
- ➢ Formononetin
- Withaphysacarpin



FIG. 11: MASS SPECTRUM OF L-GALACTOSE













FIG. 15: MASSSPECTRUM OF WITHAPHYSACARPIN

Fraction 03: Figure 16-20.

- Lycoflexine
- Carnegine

- Trihydroxycoprostanoic acid
- Isobergaptol
- D8'-Merulinic acid A



FIG. 19: MASS SPECTRUM OF ISOBERGAPTOL

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Fraction 04: Fig. 21-25.

- ✤ 1-(5-Methyl-3pyridinyl)-1-decanone
- ✤ Alnustone
- Chloropanaxydiol
- ★ (±)-2-Methyl-3-(4methylphenyl)propanal
- Phenanthrene



FIG. 23: MASS SPECTRUM OF ALNUSTONE

255 260 265 270 Counts vs. Mass-to-Charge (m/z)

280

275

285

290

235

240

245

250

0



In-vitro Cytotoxicity Studies: Human Breast Cancer Cell Lines (MCF-7):

In-vitro cytotoxicity study of extracts and flavonoid fractions on MCF-7 human breast cancer cell line revealed that flavonoid fraction F-01 showed comparable with the standard, Adriamycin. F-01

and Adriamycin showed activity at concentration $<10\mu$ g/ml. All the extracts, FRF and fractions F-02, F-03, F-04 showed activity at concentration $>80\mu$ g/ml. **Fig. 26** gives the growth curve of the extracts, FRF and the flavonoid fractions on human breast cancer cell lines(MCF-7).



FIG. 26: GROWTH CURVE: HUMAN BREAST CANCER CELL LINE MCF-7

Human Colon Cancer Cell Line (HT-29): In the *in-vitro* cytotoxicity study on human colon cancer cell line HT-29, all the extracts, FRF and the flavonoid fractions showed activity at concentration > $80 \mu g/ml$.

Adriamycin, the standard, showed activity at concentration $<10\mu$ g/ml. **Fig. 27** gives the growth curve of the extracts, FRF and the flavonoid fractions on human colon cancer cell line (HT-29).

Human Colon Cancer Cell Line (HT-29):

In the *in-vitro* cytotoxicity study on human colon cancer cell line HT-29, all the extracts, FRF, and the flavonoid fractions showed activity at concentrations> $80 \mu g/ml$. Adriamycin, the

standard, showed activity at a concentration $<10\mu$ g/ml. **Fig. 27** gives the growth curve of the extracts, FRF, and the flavonoid fractions on the human colon cancer cell line (HT-29).



FIG. 27: GROWTH CURVE: HUMAN COLON CANCER CELL LINE HT-29

Human Lung Cancer Cell Line (A-549):

In the *in-vitro* cytotoxicity study on human lung cancer cell line A-549, the standard, Adriamycin, showed activity at $<10\mu$ g/ml. All the extracts, FRF,

and flavonoid fractions showed activity at concentrations>80 μ g/ml. **Fig. 28** gives the growth curve of the extracts, FRF, and the flavonoid fractions on human lung cancer cell line(A-549).



FIG. 28: GROWTH CURVE: HUMAN LUNG CANCER CELL LINE A-549

DISCUSSION: Phytochemical screening of the plant extracts were prepared using methanol and solvents in the increasing order of their polarity (petroleum ether, chloroform, ethyl acetate, methanol, and distilled water). The extracts were evaluated for their antioxidant potential. DPPH radical scavenging assay, nitric oxide scavenging assay and hydrogen peroxide scavenging assay

were performed. The total methanolic extract's total flavonoid content was estimated using aluminium chloride method. The flavonoid-rich extract was prepared from the total methanolic extract. Thinlayer chromatography was performed using a solvent system (Ethyl acetate: Formic acid: Glacial acetic acid: Water:: 10: 0.5: 0.5: 1.5). Four distinct flavonoid bands were obtained using TLC when

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observed under long wavelength UV radiation. LC-MS scan was performed for the flavonoid bands obtained through TLC. Cytotoxicity studies of the extracts and flavonoid fractions were performed on the cells lines: MCF-7, HT-29, and A-549.

Limitations: While *in-vitro* studies were performed in this study, animal models can be used to study antioxidant and cytotoxic potential of the extracts and the fractions. More sensitive methods of characterization like GC-MS can be used.

CONCLUSION: Extracts showed various phytochemicals like flavonoids, tannins and phenols, saponins, and alkaloids. Only petroleum ether extract showed the presence of steroids. Chloroform extract showed the presence of alkaloids, flavonoids, tannins and phenols, and saponins. Ethyl acetate extract and methanol extract showed the presence of flavonoids, tannins and phenols, and saponins. Ethyl acetate and methanol extract showed good antioxidant potential among all the extracts. LC-MS data from the existing database was acquired from the fractions obtained using TLC. F-01 fraction showed comparable activity with the standard (Adriamycin) on the MCF-7 cell line.

Financial Support: No funding was received for this work.

ACKNOWLEDGEMENT: The authors would like to thank the Advanced Centre for Treatment, Research, and Education in Cancer, India, for providing the cell line studies of the samples. We would like to express deep gratitude to IIT SAIF Bombay, India, for providing with analysis of samples by LC-MS. We would thank SVKM's Dr. Bhanuben Nanavati College of Pharmacy, India, for the facilities to complete the research work.

Data Availability: No supplementary file submitted.

CONFLICTS OF INTEREST: Nil

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How to cite this article:

Parmar D and Apte M: Evaluation of seeds of an Indian sacred plant *Aegle marmelos*, for their antioxidant and cytotoxic potential. Int J Pharm Sci & Res 2022; 13(8): 3261-74. doi: 10.13040/IJPSR.0975-8232.13(8).3261-74.

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