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ANTIBIOTICS SENSITIVITY PATTERN OF *E. COLI* SPECIES ISOLATED AT TERTIARY CARE HOSPITAL

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ABSTRACT: Worldwide antimicrobial resistance is a serious problem and differs from region to region. This study has designed the determination of the incidence and sensitivity pattern of *Escherichia coli* (*E. coli*) and discusses the general issues related to antimicrobial resistance. Prospective study was carried out between JAN to April 2020. Samples of urine, blood, pus and miscellaneous (fluids, swabs and sputum) were collected from indoor patients. The present antimicrobial susceptibility of *E. coli*, the study was performed in the Department of Microbiology Narayana Super-specialty Gurugram, Total of 315 samples were tested out, 97 samples showed growth on culture and out of 97 samples, 40 *E. coli* have isolated and tested antibiotics sensitivity, it has been demonstrated that different samples showing a significant amount of antimicrobials Sensitivity and resistance to different antibiotics (Table 2). Antimicrobials, namely Meropenem have shown sensitivity (65%), imipenem (60%), Colistin (70%). Thus, there is a need for the implementation of strong antimicrobial stewardship program and stop misuse of antibiotics with an instant effect. Our region's present study helped us see the actuality of the antibiotic sensitivity pattern of bacteria, which is not good. To reduce antibiotic resistance, immediate steps must be taken as new antibiotic development is very quick. Otherwise, it will result in moving us to a pre-antibiotic era which would be a great step back for the human community.

INTRODUCTION: *Escherichia coli* is a gram-negative anaerobic, facultative, rod-shaped coli form bacterium of the genus *Escherichia* that is mostly found in the lower intestine of endotherm. Generally, *E. coli* are harmless, but few serotype ETEC and EPEC *etc.* causes serious food poisoning of its own hosts. In current year drug resistance to human pathogenic bacteria is commonly reported throughout world.

Antimicrobial resistance nowadays well known as scientific and public fitness problem³. It is an emerging issue, particularly in the health center and newly industrialized countries of Asia and the Pacific⁴. Gram-negative microorganism motive infections: pneumonia, bloodstream infections, wound or surgical web page infections, and meningitis in healthcare settings.

Gram-negative microorganisms are resistant to a couple of capsules, and more and more are resistant to the most reachable antibiotics. These microorganisms have built-in Competencies to locate new approaches to be resistant and ignore alongside genetic substances that permit different microorganisms to grow to be drug resistant. Gram-negative infections encompass these prompted by

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using *Klebsiella pneumonia* (*K. pneumoniae*), *Pseudomonas aeruginosa* (*P. aeruginosa*), *Escherichia coli* (*E. coli*) and *Acinetobacter* are as nicely as many different, less frequent bacteria. *Pseudomonas aeruginosa* is the most difficult microorganism to deal with resistance to a range of antibiotics. *Escherichia coli* (*E. coli*), the universal facultative Gram-negative bacillus in the human fecal flora, usually inhabit the colon as an innocuous commensal. Theoretically, the antimicrobial that treats *E. coli* is cotrimoxazole, fluoroquinolones, piperacillin, 0.33 technology cephalosporins, carbapenems and aminoglycosides. Surveillance statistics exhibit that resistance in *E. coli* is persistently absolute best for antimicrobial dealers that have been in use for the longest time in human and veterinary medicine. Enterobacter cloacae have been pronounced as necessary opportunistic and multi-resistant bacterial pathogens for people for the duration of the closing three many years in health facility wards.

E. cloacae and *E. hormaechei* are most often remoted from human medical specimens. *E. cloacae* complicated are no longer understood yet; this should be due to the shortage and the dispersion of records available. Its ability to structure biofilms and secrete a variety of cytotoxins (enterotoxins, hemolysins, pore-forming toxins) is vital for its pathogenicity¹¹. *Serratiamarcescens*, a gram-negative bacillus categorized as a member of the Enterobacteriaceae, has been recognized as a reason of hospital-acquired contamination for the ultimate too many years. *S. marcescens* used to be viewed in the beginning to be an innocuous, non-pathogenic saprophytic water organism and was once often used as an organic marker due to the fact its, without problems, known as pink colonies.

The colonies are 1 to 2mm, non-hemolytic, mcoid, easy and spherical on sheep's blood agar after 24 hours of incubation at 37°C¹⁵ MRSA is a precise pressure of the *S. aureus* is resistant to methicillin and all β -lactams. Later use of Oxacillin as a choice to methicillin in susceptibility assessments resulted in the time period 'Oxacillin-resistant *S. aureus*' (ORSA), which is resistant to severe antibiotics. Before the improvement of antibiotics, invasive infections prompted with the aid of *Staphylococcus aureus* have frequently been deadly¹⁸. *Candida*

species are the essential purpose of superficial to systemic fungal contamination in people and the important supply of contamination in fitness care facilities²⁰. *Albicans* is the fundamental pathogenic agent of systemic infections; however, for the duration of the current years, the charge of non-albicans *Candida* species has accelerated in many reports²¹. *Enterococcus faecalis* and *Enterococcus faecium* Enterococci reason large infections, together with intraabdominal sepsis, urinary tract infections, bacteremias and endocarditis. It has been described considerably with the aid of Hauser already in 1885 and is regarded to purpose a large spectrum of human diseases, in particular in the hospitalized patient. It bills for about 3% of nosocomial infections in the United States and ranks 1/3 as the reason of trouble-free cystitis, pyelonephritis, and prostatitis.²⁷

MATERIALS AND METHODS:

Sample Collection: This study was conducted at the Department of Microbiology in tertiary care hospital. From Jan-2020 to April 2020, different types of clinical specimens, Such as blood, urine, pus, swab, etc., were received from the inpatient. Antimicrobial susceptibility pattern *Escherichia coli* (*E. coli*), from Jan-2020 to June 2020, was analyzed and reported.

Identification of Bacteria: The bacteria were cultured on MacConkey's agar, Nutrient agar, blood agar and other selective media, followed by the recognition of the isolates based on their cultural characteristics, gram staining, motility, and reactions in standard biochemical tests.



FIG. 1: ESCHERICHIA COLI GROWTH ON CLAD AGAR MEDIUM

Antimicrobial agents: By the Kirby-Bauer disk diffusion technique on Muller Hinton Agar by

Filter Paper disks impregnated with antibiotics (Span diagnostics Limited, Surat, India) susceptibility. A pre-diffusion time of 30 min was allowed at room temperature, and the plates were incubated at 37°C for 24 h. The diameter of the zone of inhibition was measured and compared to that of standard strain, and the results were interpreted as sensitive or resistant, based on Clinical Laboratory Standard Institute 2014 guidelines. The percentage antimicrobial susceptibility of the isolated microorganism against different antimicrobials tested was calculated and interpreted as sensitive and resistant

organisms were isolated from 315. Out of all samples, the highest percentage was shown in urine (41.23%) followed by 21% in blood. Out of 97 positive samples of *E. coli*, 70 samples (72.16%) were isolated in women. In the present study, colistin shows the highest sensitivity 70%, meropenem has shown the 65%. The antimicrobial agents are losing their growth on culture, and out of 97 samples, 40 *E. coli*, were isolated and tested for antibiotics sensitivity; it has been demonstrated that different samples show a significant amount of antimicrobials Sensitivity and resistance to different antibiotics **Table 2**. Antimicrobials, namely Meropenem have shown sensitivity (65%), imipenem (60 %), Colistin (70%).

RESULTS AND DISCUSSION: During this study total sample collected was 315; a total 97

TABLE 1: PRESENCE OF *E. COLI* STRAINS IN OUTPATIENT POPULATION WITH POSITIVE URINE CULTURES. X²= 40.0071; P=0

Gender	Men	%	Women	%
Isolated <i>E. coli</i> strains	20/97	20.61%	70/97	72.16%

TABLE 2: ISOLATION OF ORGANISM FROM DIFFERENT CLINICAL SAMPLES

Nature of Sample	No of sample (N=97)	Percentage (%)
Blood	21	21.64948454
Urine	40	41.2371134
E/T Section	12	12.37113402
Pus	13	13.40206186
Bal	5	5.154639175
Sputum	2	2.06185567
As cite Fluid	1	1.030927835
Drain Fluid	2	2.06185567
Bile CBD Tip	1	1.030927835
Total	97	100

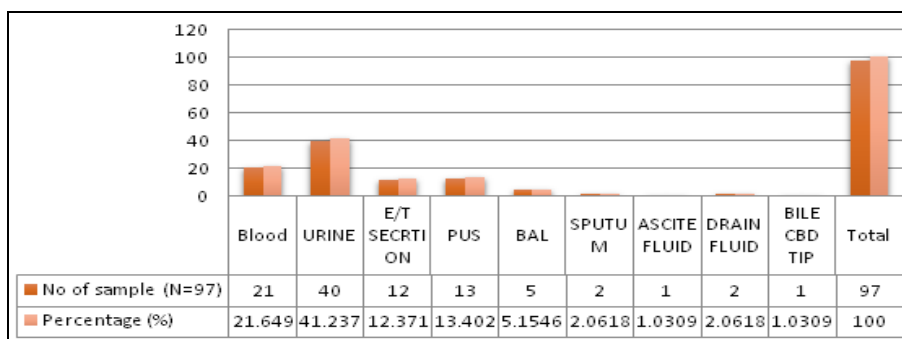


FIG. 2: ISOLATION OF ORGANISMS FROM DIFFERENT CLINICAL SAMPLES

TABLE 3: ANTIBIOTICS SENSITIVITY PATTERN

Antibiotics	Resistance	%	sensitivity	%	Intermediate	%
Meropenem	4	10	26	65	1	2.5
Imipennem	2	5	24	60	2	5
Ertapenem	4	10	22	55		
Gentamycin	4	10	23	57.5		
Amikacin	3	7.5	22	55		
Ciprofloxacin	14	35	13	32.5	3	7.5
Levofloxacin	1	2.5	1	2.5		
Colistin	0	0	28	70		

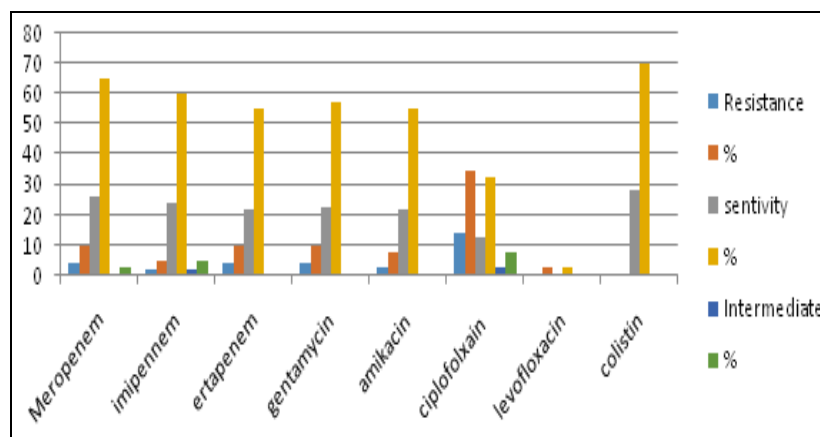


FIG. 3: ANTIBIOTICS SENSITIVITY PATTERN

CONCLUSION: In this study, the clinical isolation of *E. coli* indicates resistance to commonly used antibiotics and towards newer antibiotics. Because of the spread of resistant organisms and indiscriminate use of antibiotics, lack of awareness, patient noncompliance, and unhygienic conditions antimicrobial agents are losing their efficacy.

It is the need of time that antibiotic policies should be formulated to resist and overcome this emerging problem and to, implement a robust antimicrobial stewardship program, and stop the misuse of antibiotics with an instant effect. Our region's present study helped us see the actuality of the antibiotic sensitivity pattern of bacteria, which is not good. For reduction of the problem of antibiotic resistance, immediate steps must be taken as new antibiotic development is very quick. Otherwise, it will move us to the pre-antibiotic era, which would be a great step back for the human community.

Author's Contribution Statement: Mr. Vipin Kumar, Dr. Malvika, and Dr. Sachin Gupta studied the literature review and gathered the data about this work. Dr. Vipin Kumar Sharma, Dr. Swadesh, Dr. Kapil, and Dr. Ashwani & Rohan Kaushik analyzed these data and necessary inputs were given towards designing the manuscript. All Authors discussed the methodology and result and contributed to the final manuscript.

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CONFLICT OF INTEREST: The Authors confirm no conflict of interest.

REFERENCES:

- Jahan F, Lawrence R, Kumar V and Junaid M: Journal of Chemical and Pharmaceutical S Research 2011; 3(4): 777-789.
- Piddock KJV and Wise R: J Antimicrob Chemother 1989; 23: 475-483.
- Oteo J, Campos J and Baquero F: Antibiotic resistance in 1962 invasive isolates of *Escherichia coli* in 27 Spanish hospitals participating in the European Antimicrobial Resistance Surveillance System. J. Antimicrob Chemother 2002; 50: 945-52.
- Elizabeth B and Hirsch: Impact of Multidrug-resistant *Pseudomonas aeruginosa* Infection on Patient Outcomes. Expert Rev Pharmacoeconomics Outcomes Res 2010; 10(4): 441-6.
- US Food and Drug Administration. National antimicrobial resistance monitoring system- enteric bacteria (NARMS): 2008 executive report. Rockville (MD); 2010.
- Petri WA: Penicillins, Cephalosporins and Other B Lactam Antibiotics. In: Laurence LB, editor, Bruce AC, Bjorn CK, assoc. editors. Goodman and Gilman's The Pharmacological Basis of Therapeutics. Ed 12th China McGraw Hill 2011; 1472: 1480-1481.
- Jethwani U, Shah N and Trivedi P: Antibiotic Sensitivity Pattern of Gram Negative Bacilli Isolated from the Lower Respiratory Tract of Ventilated Patients in the Intensive Care Unit. Indian Medical Gazette 2014; 180-184.
- Niranjan V and Malini A: Antimicrobial resistance pattern in *Escherichia coli* causing urinary tract infection among inpatients, Indian Journal of Medical Research 2014; 139(6): 945-948.
- Joseph OA, Oluwatoyin O, Egbedokun A and Owolabi AT: Antibiotic susceptibility pattern of *Escherichia coli* isolated from outpatient individuals Attending the University College Hospital (UCH), Ibadan, Nigeria, Journal of Infectious Diseases and Treatment 2017; 3(1): 16.
- Bari MA, Arefin M, Nessa M and Mostofa G: Recent Antibiotic Sensitivity Pattern of *Escherichia coli* in Urinary Tract Infection, Journal of Teachers Association 30(1): 61-65.
- Onifade AK and Oladoja MA: Antibiotics Sensitivity Pattern of *Escherichia coli* isolated from Children of School Age in Ondo State, Nigeria, Science pub, 2015; 7(2): 1-8.

12. World Health Organization, WHO Global Report on Surveillance of Antimicrobial Resistance. Geneva Switzerland 2015.
13. Ranjana M, Rama S and Chaudhary U: Antimicrobial sensitivity pattern among clinical isolates of *Escherichia coli* in tertiary care centre of Northern India, *International J of Research in Medical Sciences* 2016; 4(2): 639-642.
14. Jackson CR, Jonathan GF and Sohyun C: Prevalence and characterization of *Escherichia coli* isolated from the Upper Oconee Watershed in Northeast Georgia, *PLoS ONE* 2018; 13(5): 307-315.
15. Probert WS, Miller GM and Ledin KE: Contaminated stream water as source for *Escherichia coli* O157 illness in children. *Emerge Infect Dis* 2017; 23(7): 1216–1218.
16. Metropolitan North Georgia Water Planning District. Water resource management plan. Appendix A: River basin profiles. Atlanta (GA): Metropolitan North Georgia Water Planning District 2016.
17. Clinical and Laboratory Standards Institute (CLSI). Performance standards for antimicrobial susceptibility testing; Ed 26th CLSI Supplement M100S. Wayne (PA): CLSI 2016.
18. Titilawo Y, Obi L and Okoh A: Antimicrobial resistance determinants of *Escherichia coli* isolates recovered from some rivers in Osun State, South-Western Nigeria: implications for public health. *Sci Total Environ* 2015; 523: 82–94.
19. Lupindu AM: Isolation and Characterization of *Escherichia coli* from Animals, Humans, and Environment, Open access peer-reviewed chapter 2017.
20. Ghassemi A, Farhangi H, Badiie Z, Banihashem A and Mosaddegh M: Evaluation of nosocomial infection in patients at hematology-oncology ward of Dr. Sheikh children's hospital. *Iran J PedHematol Oncol* 2015; 5: 179-85.
21. Gokcebay DG, Yarali N, Isik P, Bayram C, Ozkaya-Parlakay A and Kara A: *Candida* associated bloodstream infections in pediatric hematology patients: a single center experience. *Mediterr J Hematol Infect Dis* 2016; 8(1): 81-92.
22. Hegazi M, Abdelkader A, Zaki M and El-Deek B: Characteristics and risk factors of candidemia in pediatric intensive care unit of a tertiary care children's hospital in Egypt. *J Infect Dev Ctries* 2014; 8: 624–634.
23. Badiie P and Alborzi A: Assessment of a real-time PCR method to detect human non-cryptococcal fungal meningitis. *Arch Iran Med* 2011; 14: 381–384.
24. Murray BE: The life and times of the enterococcus. *Clin Microbiol Rev* 1990; 3: 46-65.
25. Gorkiewicz G: Nosocomial and antibiotic-associated diarrhea caused by organisms other than *Clostridium difficile*. *Int J Antimicrob Agents* 2009; 33: 537-541.
26. Centers for Disease Control and Prevention. National nosocomial infections surveillance (NNIS) report, data summary October 1986-April 1996, issued May 1996. *Am J Infect Control PubMed* 1996; 24: 381.
27. Stamm WE: Urinary tract infections. In: Root RK, editor. *Clinical infectious diseases: a practical approach*. Oxford University Press Inc New York NY 1999; 649-56.
28. Clinical and Laboratory Standards Institute. Performance Standards for Antimicrobial Susceptibility Testing; Twenty-Fourth Infor Sup (M100-S24); 2014; 34(1): 110-3.
29. Singh V, Davidson A and Peter J: Hume, Vassilis Koronakis. Pathogenic *Escherichia coli* Hijacks GTPase-Activated p21-Activated Kinase for Actin Pedestal Formation *mBio* 2019; 10(4).

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