



Received on 18 January 2022; received in revised form, 25 April 2022; accepted 27 April 2022; published 01 September 2022

PLANT DERIVED ANTICANCER AGENTS FOR THE TREATMENT OF CANCER

Virender Kumar, Vandana Garg* and Harish Dureja

Faculty of Pharmaceutical Sciences, M. D. University, Rohtak - 124001, Haryana, India.

Keywords:

Phytoconstituents, Cancer, Bioactive compounds, Chemopreventive, Herbal drug

Correspondence to Author: Vandana Garg

Assistant Professor,
Faculty of Pharmaceutical Sciences,
M. D. University, Rohtak - 124001,
Haryana, India.

E-mail: Vandugarg@rediffmail.com

ABSTRACT: Cancer is defined as a condition where cancerous cells multiply out of control in the body. It severely affects the human population on a global basis. New therapies are constantly needed to prevent and treat this life-threatening disease. Natural-derived compounds are attracting scientific and research interest due to their substantially fewer side effects than the current cancer treatment. Plants produce secondary metabolites with anticancer properties and are being examined for their potential to be used as clinical drugs. Amazingly, antineoplastic activity has been reported even in most commonly found locally dwelling plants like ginger, turmeric, garlic, and Ashwagandha, besides their use in other illnesses by the local community. However, many medicinal plants are yet to be properly unveiled for their potential as anti-cancerous agents. They need extensive research and valuable resources for in-depth studies like identification, isolation, *in-situ* studies, bioequivalence studies, and *in-vitro*, *ex-vivo*, or *in-vivo* studies. This review summarizes the role of bioactive compounds in cancer, the mechanism by which compounds show anticancer activity, and the list of plants used for anticancer activity and clinical trials. The scientist will test plant-based anticancer agents and fill the research gap seeking chemopreventive substances.

INTRODUCTION: Today's modern era has significantly improved people's quality of life with newer established technologies and utilities, especially in the healthcare sector, like developing new diagnostic techniques, medical devices, treatment strategies, and surgical procedures. However, there are certain deadly diseases for which no suitable and appropriate treatment is available, although immense research is already on them.

Cancer is a life-threatening disease that costs about 8 million out of 14 million people worldwide of every age group as per the world health organization, especially in developing and developed countries¹. Cancer increases the mortality rate every day, with an estimated 13.1 to 17 million fatalities by 2030². Cancer has been ranked second after cardiovascular diseases for morbidity and mortality in the general population³.

Cancer is a situation in which the body's normal cells undergo uncontrolled division leading to tumor formation at the affected site, which damages and affects adjacent normal healthy tissues abnormally⁴. The prime reason behind the cancer transformation of normal cells is a mutation in DNA and hence, in genes, leading to the development of oncogenes in them.

<p>QUICK RESPONSE CODE</p> 	<p style="text-align: center;">DOI: 10.13040/IJPSR.0975-8232.13(9).3375-96</p> <hr/> <p style="text-align: center;">This article can be accessed online on www.ijpsr.com</p>
<p>DOI link: http://dx.doi.org/10.13040/IJPSR.0975-8232.13(9).3375-96</p>	

These can be caused by physical factors (X-rays), organic compounds, virus-infected pathogenic microorganisms, or behavioural and dietary variables such as a lack of physical activity, smoking, alcohol intake, and an unhealthy diet⁵. The cancer cells can stimulate tumor growth at the

local site (benign tumor) or can be significantly transmitted to various parts or organs of the body *via* the bloodstream (malignant tumor)⁶. Cancer can be classified into various types based on affected sites like cells, tissues, or organs, as represented in Table⁷.

TABLE 1: CLASSIFICATION OF CANCER

S. no.	Types of cancer	Affected / Associated part
1	Carcinoma	Skin, lungs, breast, pancreas, other organs & glands
2	Sarcoma	Bone, muscle, adipose tissue, blood vessels, cartilage or other soft, connective tissue
3	Melanoma	Melanocytes (pigmentation of the skin)
4	Lymphoma	Lymphocytes
5	Leukemia	Blood

Various methods are used to treat cancer, including traditional drugs⁸ as extract of plant⁹, chemotherapy¹, radiotherapy, and surgery. Treatment of cancer using certain chemicals, *i.e.*, chemotherapy, tends to cause side effects in cancer patients and severe adverse drug reactions like nausea, vomiting, hair loss, discomfort, and mental distress, simultaneously affecting other normal cells because of its non-selective approach and toxicity on slight over dosage⁷.

Patient non-compliance overdosage is also a significant problem in chemotherapy treatment. Nevertheless, surgery needs to be performed on terminal cancer patients. In current years, there has been a substantial shift towards traditional natural herbal medicines and the active compounds obtained after extraction from plants for cancer treatment because of various shortcomings of chemotherapy, as mentioned above.

Since, the herbal medicines are naturally obtained, they have lesser toxic side effects on patient health, are easily affordable, are considered safe to use, and produce deemed positive results¹⁰. Plants produce compounds like volatile oils, resins, gums, alkaloids, terpenoids, saponins, and secondary metabolites. The anti-cancerous evaluation and the efficacy have been determined with the help of various assays performed on several induced tumours as mentioned below:-

1. MTT (3 - (4, 5 - dimethylthiazol - 2 - yl) - 2, 5 - Diphenyltetrazolium bromide) Colorimetric Assay: The extract mixed with DMSO is added to a diluted cell culture media (pure cell lines) and seeded for incubation in 96 well-microtiter plates along with the addition of MTT. Cell viability and

absorbance were determined at definite time intervals¹¹.

2. WST - 1 (4 - [3 - (4 - iodophenyl) - 2 - (4 - nitrophenyl) - 2H - 5 - tetrazolio] - 1, 3 - benzene disulfonate) Colorimetric Assay: Firstly, pure cell lines are incubated in 96 well-microtiter plates with culture media. Later, definite concentrations of the extract are added and simultaneously dissolving the contents in DMSO. Finally, adding definite proportions of WST-1 to the above-solubilized mixture. The absorbance was determined at definite time intervals¹².

3. SRB (Sulforhodamine B) Method: Absorbance was determined for cultured RPMI 1640 medium of fetal bovine serum in which extract, TCA, and SRB solution were added simultaneously and kept for incubation¹³.

4. Alamar Blue Resazurin Reduction Assay: Serial dilutions of plant extract are prepared with a medium containing pure cell lines suspended in DMEM and seeded for incubation in 96 well-microtiter plates. Afterward, culture media along with resazurin is added and kept for further incubation. The cytotoxic effect of the extract is determined by analyzing the resultant intensity of the fluorescent dye.

Various researchers proposed herbal extract, and the compound derived from the plants showed anticancer effects by numerous mechanisms. They have oxidation-resistant properties or act as antioxidants, resulting in an antiproliferative effect and inducing apoptosis through various subcellular metabolic pathways like decreasing the lipid peroxidation level, acid phosphatase activity, or

inhibition of PI3K/Akt pathway¹⁴ or repairing DNA and enhancing body immunity¹⁵ Some of them are known to cause the arrest of the cell cycle at the metaphase stage by binding to tubulin protein, an essential constituent of microtubules that disrupts its functions and hence, mitotic spindle apparatus.

TABLE 2: FORMULATION OF ANTICANCER DRUG

S. no.	Active chemical constituent of plant	Formulated anticancer drug
1	Taxanes	Paclitaxel (taxol), docetaxel
2	Vinca alkaloids	Vinblastine, vindesine, vincristine
3	Camptothecin	Camptothecin derivatives like irinotecan
4	Epipodophyllotoxins	Etoposide, teniposide

In this way, they prevent cancer cell proliferation and suppress tumor growth. Any plant part like stem, leaves, bark, flowers, shoot, roots, rhizomes, and fruits, which have anti-cancerous properties, can prepare extracts and derive novel compounds for treating cancer *via* various extraction methods¹⁵ Although anti-cancerous activity has been detected in more than 3000 plants¹⁶ till today, in

the pharmaceutical market, only four major classes of drugs or medicinal agents obtained from plants¹⁷ are available for cancer treatment with suitable approved formulations for patients which are mentioned in the table below:-

Mechanisms Involved in Anticancer Activity of Herbal Drugs: Numerous herbal drugs show anticancer potential used in cancer therapy. Various mechanisms responsible for the anticancer potential of the herbal medication are described below:

1. Apoptosis: Several cancers are therapeutically targeted by apoptosis, the most common process of programmed cell death. Apoptosis is characterized by morphological changes such as heterochromatin mass core, shrinkage of the cell membrane, and losing cytoplasmic organelles' position. Researchers found that several dietary compounds inhibited carcinogenesis *via* induction of apoptosis. These compounds include curcumin, resveratrol, apigenin, quercetin, and ellagic acid^{18, 19}. Apoptosis induction is the most significant marker of the anticancer herbal drug²⁰.

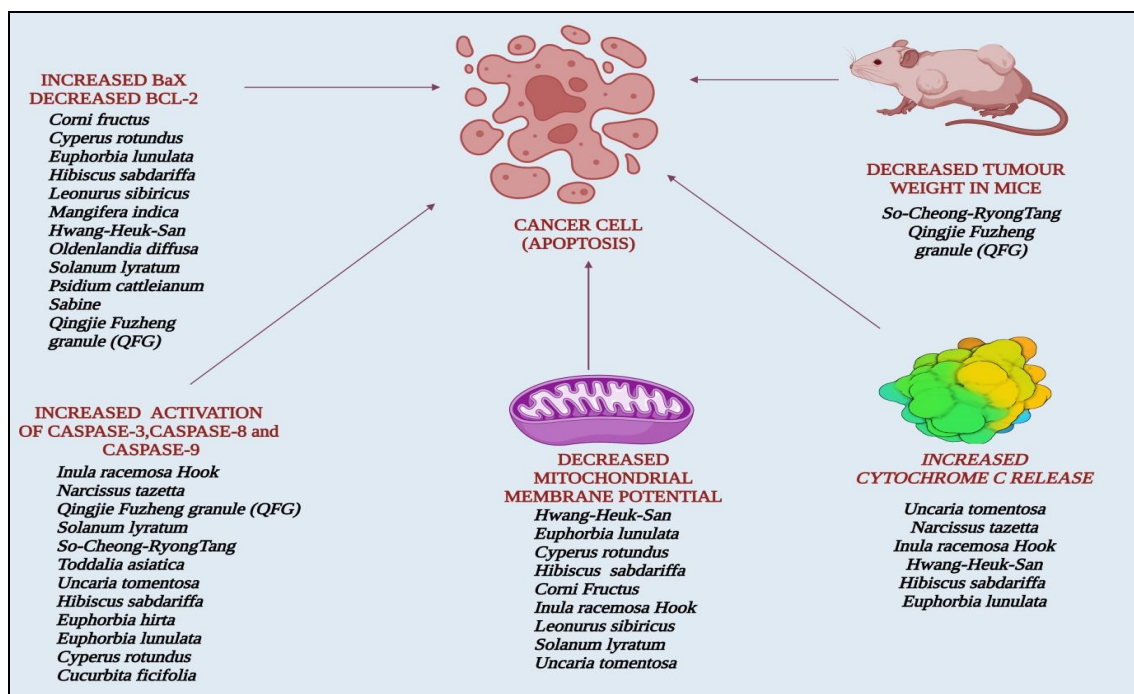


FIG. 1: HERBAL DRUG SHOWING ANTICANCER POTENTIAL VIA APOPTOSIS

2. Angiogenesis: It is the process which involves the development of new blood vessels from the old blood vessel resulting in increased blood flow²¹. Angiogenesis plays an essential role in cancer pathogenesis, and targeting is necessary for treating

the disease²². A variety of natural compounds have been recently shown to modulate tumor angiogenesis by inhibiting angiogenesis-associated cytokines or other mechanisms²³. These compounds include Resveratrol, Epicatechin,

Epigallocatechin, Genistein, Curcumin, *Triphala churna*, Red ginseng, and Sanguinarine²⁴. These plant-derived compounds act on various factors like pro-angiogenic (vascular growth endothelial factor), receptor-mediated signaling pathways

(PI3K/Akt), matrix protein, and hypoxia-inducible factor. Different molecular mechanisms have been used to describe many plant-based compounds that exhibit antiangiogenic properties, shown in **figure 2**.

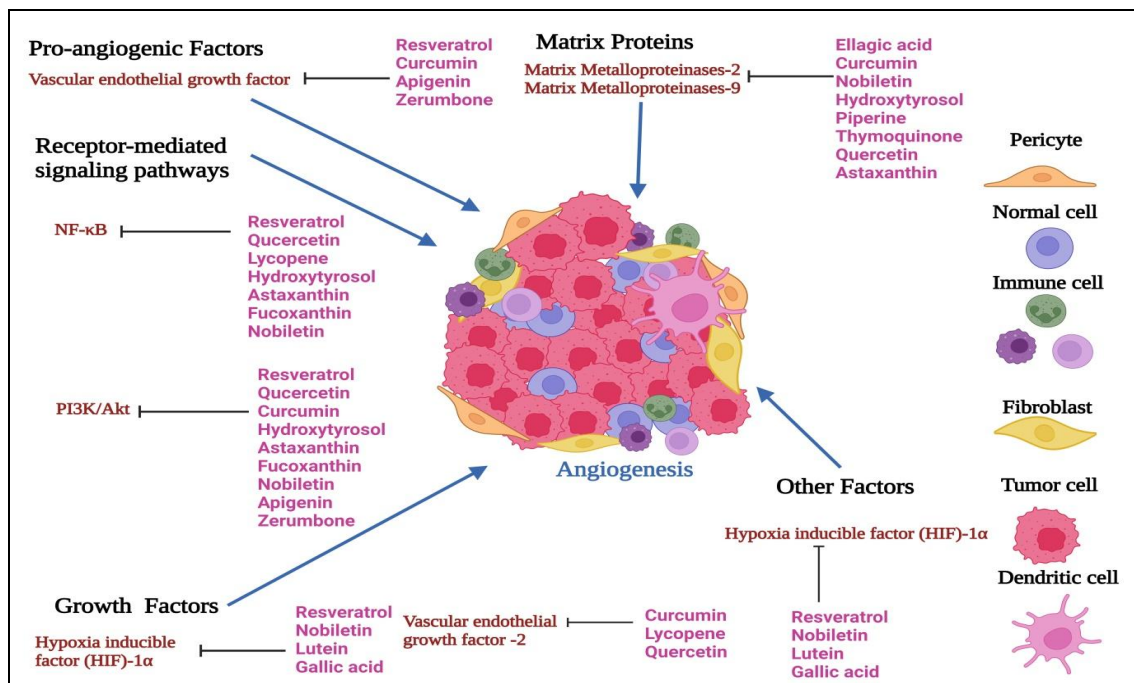


FIG. 2: HERBAL DRUG SHOWING ANTICANCER POTENTIAL VIA ANGIOGENESIS

3. Cell Cycle Arrest: In the cell cycle series of events occurs at the macromolecular level, *i.e.*, cell division and formation of two daughter cells²⁵. In a cell cycle process, a new piece of DNA is produced, chromosomes must be segregated, cells

undergo mitosis, and then they begin to divide²⁶. A cell's ability to adapt to a constantly changing microenvironment relies on regulating cell cycle progression^{27,28}.

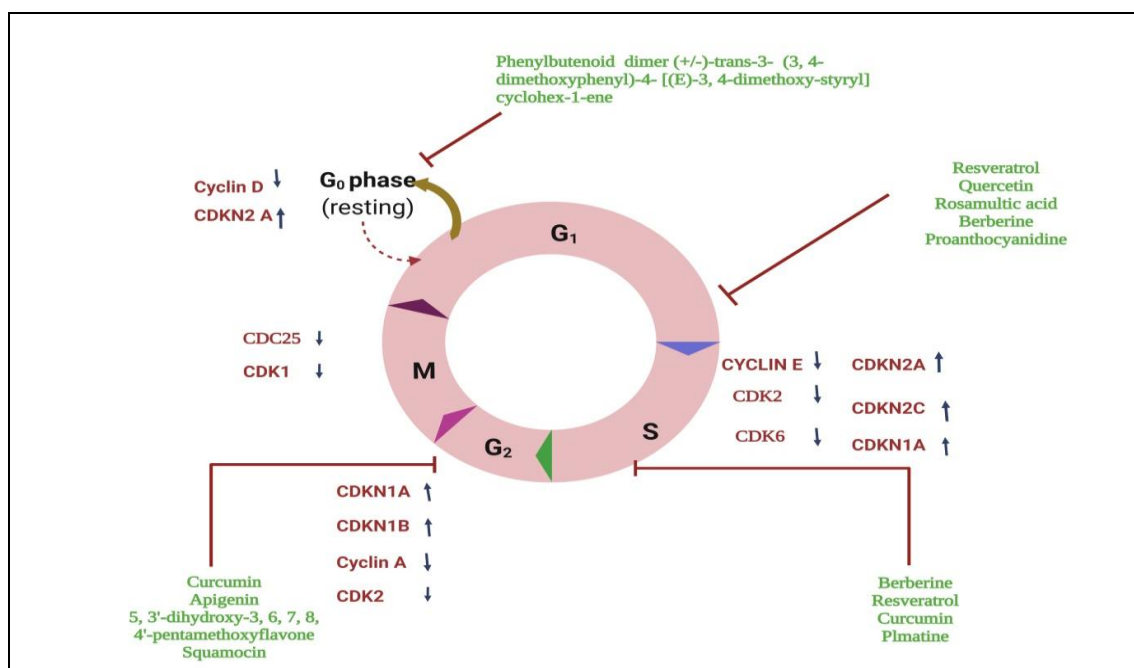


FIG. 3: HERBAL DRUG SHOWING ANTICANCER POTENTIAL VIA CELL CYCLE ARREST

Understanding how uncontrolled and rapid cell division contributes to cancer development is the key factor in selecting the anticancer drug. It has been shown *in-vivo*, *in-vitro* and in clinical studies that it is possible to kill cancer cells with natural compounds that inhibit the cell cycle arrest.

Various natural compounds berberine, quercetin, apigenin, chebulagic acid, squamocin, rosamultic acid, silibinin, bufotalin, palmatine, paclitaxel, rhizoxin, and tryprostatin acting on the cell cycle at different checkpoints of the cell cycle. Different checkpoints are affected by various herbal drugs, as shown in diagram 4.

4. Autophagy: Autophagy is a catabolic process that evolved and delivered cytoplasmic components

to the lysosome for degradation in lysosomes²⁹. The main objective of autophagy is to maintain cell vitality by eliminating damaged proteins^{30, 31}. There has been evidence that natural products play an essential role in modulating autophagy, correlated to pro-survival autophagy or autophagic cell death^{32, 33}.

These compounds include Quercetin, Kaempferol, Genistein, Resveratrol, Rottlerin, Ursolic Acid, Camptothecin, Neferine, and Thymoquinone³⁴.

To develop novel therapeutic approaches, autophagy targets must be identified. Several natural products can induce or inhibit autophagy and target various stages of autophagy, as shown in the figure.

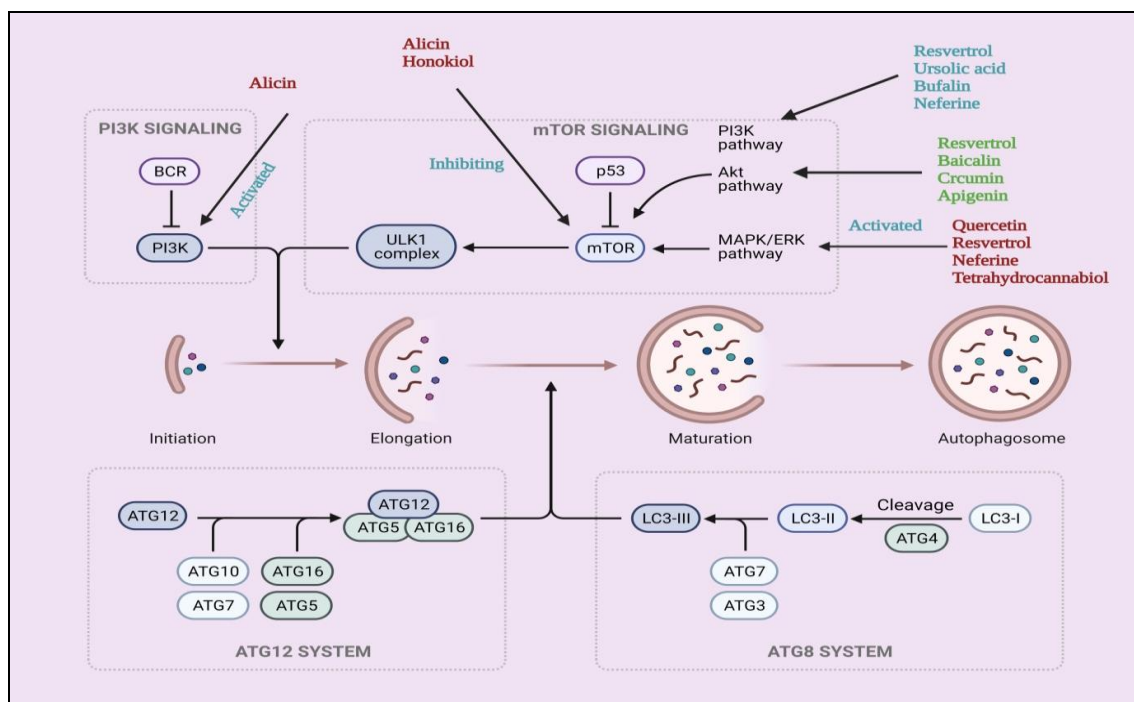


FIG. 4: HERBAL DRUG SHOWING ANTICANCER POTENTIAL VIA AUTOPHAGY

Phytochemicals and the active constituents derived from different plant parts, *i.e.*, leaf, stem, rhizome, fruit, seed, root, pericarps, and fruit, show different pharmacological properties and therapeutic potential.

The different experimental studies found that plant extracts and their isolated phytoconstituents can exert chemopreventive actions³⁵.

These herbal drugs act by different approaches, *i.e.*, made more responsive cancer cells to herbal chemotherapeutic drugs, increase the accumulation

of herbal drugs in cancerous cells, and reduce the adverse effects of chemotherapeutic agents³⁶.

Herbal drugs from natural sources are excellent candidates for cancer treatment because of their capacity to affect different targets such as migration and growth of cells and numerous molecular pathways.

Updated information about plant binomial name, common name, family name, part of the plant used, active constituents, and the mechanism responsible for anticancer activity are given in **Table 3**.

TABLE 3: MEDICINAL PLANT USED AS ANTICANCER DRUG

S. no.	Plant binomial name	Common name	Family	Plant part used	Effective against specific cancer type	Active chemical constituent	Mechanism of action	Ref.
1.	<i>Zingiber officinale</i>	Ginger, inguru	Zingiberaceae	Rhizome	Gastrointestinal, esophageal, liver	Gingerols converted to shogaols, paradols, zingerone Curcumin	Suppresses TNF α & reduces the expression of NF κ B Downregulation of COX-2 enzyme, apoptosis via caspase-3 activity & increasing level of ROS, Ca ²⁺	37
2.	<i>Curcuma longa</i>	Turmeric, kaha	Zingiberaceae	Rhizome	Colon, hepatocellular, MCF-7 & ZR-75 breast type, renal, prostate, T-cell leukemia, B cell lymphoma	Curcumin	Upregulation of CD83 and activation of caspases 3&9, inhibition of glutathione S-transferaseP expression	38
3.	<i>Hemidesmus indicus</i>	Indian sarsaparilla	Apocynaceae	Root, leaves	Leukemia, liver, uterine breast, HepG2	Ledol, nerolidol, caryophyllene, camphor, borneol, lupeol, dodecanoic acid	Inhibits PI3K, AKT, Mtor, stat3 responsible for cell proliferation	39-42
4.	<i>Munronia pinnata</i>	Bin kohomba	Meliaceae	Leaves, root & even whole plant	Lung, brain, LLC, T47D, S-180, MethA, PANC-1	B-caryophyllene, caryophyllene oxide ganoderiol F	Antioxidant activity, upregulation of p53 tumor suppressor gene, downregulation of Bcl2	43-44
5.	<i>Smilax zeylanica</i>	Kumarika, kabarossa	Smilacaceae	Root, stem	Lung cancer, Breast cancer (MDA-MB 231, MCF-7)	Diosgenin, smilagenin, β -sitosterol, phenolics, flavonoids, tannins	Inhibits cell cycle at G ₁ & G ₂ by suppression of cyclin D1, anti-apoptotic protein Bcl-xL	45
6.	<i>Tinospora cordifolia</i>	Heart-leaved moonseed	Menispermaceae	Stem	Leukemia, Hela	Berberine, choline, tembetarine, tetrahydropalmatine, β -sitosterol, cordiofolioside A, giloinsterol, furanolactone, palmatine.	Free radical scavenger activity, antioxidant effect, arrest of G ₂ phase (prevents cell proliferation) Reduction of glutathione level, apoptosis, antioxidant activity by quenching free radicals	46-47
7.	<i>Adenanther apavonina</i>	Red lucky seed, madatiya	Leguminosae	Bark	Leukemia (HL-60), lymphoma, Hela cells, HCT-116 cells	β -sitosterol, arginine, cysteine, arachidonic acid, dulcitol, lignoceric acid, malonic acid	Induces caspase 3/7 & upregulated Fas protein, inhibiting NF κ B	48
8.	<i>Thespesiapo pulnea</i>	Malvaceae	Tulip tree, gansuriya	Leaves	Leukemia, lymphoma, B16-F10 melanoma	,Lupeol, β -sitosterol, Kaempferol.	Inhibits S, G ₁ phase of cell cycle, apoptosis Inducement	49-50
9.	<i>Phyllanthus emblica</i>	Phyllanthaceae	Gooseberry, nelli	Fruit or fruit juice	Throat, lung cancers, A549 (lung), HepG2, Hela, SW620 (colon)	Pyrogallol, quercetin, Kaempferol, geraniin.	Generation of ROS, Topoisomerase-I inhibited, MAP kinase activated	51-52
10.	<i>Boerhavia diffusa</i>	Nyctaginaceae	Hog weed, karichcharani	leaves	Gastric, liver, MCF-7 cells	Rhamnoside, quercetin, borhaavone.	Alterations in the levels of glutathione, Glutathione-S-transferase activity increased	53
11.	<i>Ziziphusnumulariawig ht</i>	Rhamnaceae	Harbor, wild jujube	Root, bark, stem, flowers, seeds	p53 mutant cells	Botulin, betulinic acid		54-55
12.	<i>Andrographispaniculata</i>	Acanthaceae	The king of bitters, kiryat	Roots, leaves	KB, P388, MCF7, HCT-116, HT29	Diterpenes, flavonoids, stigmasterols, andrographolide		56-59

13.	<i>Centella asiatica</i>	Apiaceae (umbelliferae)	Brahmamandu ki, Asiatic pennywort, gotu kola	Leaves, whole plant	Ehrlich ascites cells	Vallerine, pectic acid, sterol, flavonoids, ascorbic acid, asiaticoside,	DNA synthesis inhibition, increase the level of Ca ²⁺ ATPase	60-61
14.	<i>Annona atemoya</i>	Annonaceae	Sour-sop of America, mamaphal	Root, bark, leaf, fruit	A-549, MCF-7, HT-29, HepG2	Bullatacin, annonuricin A&B,	Induces apoptosis via antioxidant activity	62-64
15.	<i>Mappia foetida</i>	Icacinaceae	Amruta, kalgur, narkya	Whole plant, leaves	Hela cells, L-120 cells, breast carcinoma	Camptothecin	Inhibits nucleic acid synthesis, DNA topoisomerase-I	65-68
16.	<i>Withania somnifera</i>	Solanaceae	Ashwagandha, winter cherry	Roots, leaves	HL-60, sarcoma	Withanolide A, withaferin A	Apoptosis activated and generation of NO, increase in production of inflammatory mediators (CD4 ⁺ , IFN- γ , IL-2)	69-73
17.	<i>Cedrus deodara</i>	Pinaceae	Devdar (timber of gods), Himalayan cedar	bark	HL-60 cells, leukemia	Oleo-resin, wiktstromal, matairesinol, dibenzylbutyrolactol	Activation of caspase 3,8,9, increase in level of NO, sub G ₀ cells concentration and formation of apoptotic body.	74-75
18.	<i>Boswellia serrata</i>	Burseraceae	Indian olibanum, salaiguggul	Stem, bark,	Hela cells, carcinoma	Oleo gum resin, boswellic acid	Excessive OS, NO formation, increased sub G ₀ fraction, Bcl-2 cleavage, expression of CDR4, apoptosis	76-80
19.	<i>Aloe barbadensis</i>	Cape aloe, mussabar	Asphodelaceae (liliaceae)	Whole plant	HepG2, breast and cervical cancer	Aloin, emodin, volatile oil, barbaloin, chrysophanic acid, β -barbaloin, isobarbaloin	Downregulation of cyclin D1, CYP1A1, CYP1A2	81
20.	<i>Arbutus andrachne</i>	Greek strawberry tree	Ericaceae	Leaves, aerial parts	MCF-7 cell line, CACO-2 and HRT18 cell line	Arbutin, flavanoids, polyphenols	Antioxidant activity, antiproliferative and lipid peroxidation	82
21.	<i>Aristolochia longa</i>	Barratzam	Aristolochiaceae	Roots	VCREMS, breast cancer	Aristolochic acid, aristolctams, aporphines, isoquinolones, flavanoids, coumarins, terpenoids	Induced apoptosis	83
22.	<i>Asplenium nidus</i>	Bird's nest fern, langsuyar	Aspleniaceae	Whole plant	HepG2, HeLa	Gliciridin-7-O-hexoside, phenols, flavanoids, quercetin, kaempferol, myricetin-3-O-rhamnoside	Antiproliferative effect	84
23.	<i>Averrhoa bilimbi</i>	Star fruit, carambola	Oxalidaceae	Fruit, leaves	MCF-7	B-sitosterol, apigenin, fucopyranoside, flavanoids	Induced apoptosis via antioxidant and inhibits DNA replication	85-86
24.	<i>Azadirachta indica</i>	Neem, idian lilac, margosa	Meliaceae	Leaves, seeds	EACC, HBP carcinoma	Nimbin, nimbolide, nimbidin, limonoids, β -sitosterol, quercetin, ascorbic acid, polyphenolic compounds, azadirachtin	Antioxidant activity, Modulated the activity of VEGF, p53 and NF-kB	87-88
25.	<i>Barleria grandiflora</i>	Devkaranti, shemmul	Acanthaceae	Leaves	A-549, DLA	Alkaloidal, phenolics, flavanoids, balarenone, barlerinoside, lupulinoside, lupeol, β -sitosterol	Antioxidant activity leading to induced cell death via ROS generation, decline in tumor growth by antiproliferative effect	89
26.	<i>Berberis aristata</i>	Indian barberry, daruhaldi, tree urmeric	Berberidaceae	Root, stem	Colo205, Hop62, HT29, SiHa, MIA-PA-CIA-2, DWD, T24, PC3, A549, ZR75-I, A2780, DU145,	Alkaloidal, flavanoids, berberine, berbamine, columbamine, oxyacanthine, chelidonic acid	Influencing the mitochondrial Transmembranepotential, MMP regulation, p53 activation, NF-kB signal activated,	90

					MCF7, K562		inhibition of telomerase.	
27.	<i>Caesalpinia sappan</i>	Sappan wood, brazil wood	Caesalpinaceae (fabaceae)	Heartwood, leaves	MCF7, A549 cell lines	Ombuin, quercetin, rhamnetin, sappanchalcone, sappanol, 8-methoxy bonducellin, chromanomones, brazilein, brazilin	Induced S phase & G ₂ /M phase accumulation, acts on p53 independent pathway & induce apoptosis	91-92
28.	<i>Calligonum camosum</i>	Fire bush, arta	Polygonaceae	Whole plant	HepG2	Catechin, dehydrocatechin, kaempferol, quercetin, isoquercetrin, mequilianin, β -sitosterol, lauric acid, myristic acid, palmitic acid	Apoptosis induced via ROS generation, induced G ₀ /G ₁ , antioxidant enzyme catalase	93-95
29.	<i>Cenchrus ciliaris</i>	African foxtail grass	Poaceae	Aerial parts, roots	HepG2, CACO, A-549	Glycosides, flavanoids, steols, triterpenes, anthraquinones	Antioxidant and antiproliferative activity	96-97
30.	<i>Cenchrus antiochia</i>	Sand burs, sand spur	Asteraceae	Aerial parts	Vero, HeLa	Alkaloidal, phenolics, sterols, anthraquinones	Antioxidant activity and altering the caspase enzyme activation	98
31.	<i>Vitexagnus castus</i>	Monk's pepper, abraham's balm	Lamiaceae	Fruits, leaves	MCF-7 breast cancer	1,8- cineole, phenolic compounds, sabinene, β -caryophyllene, flavanoids (vitexin, casticin), iridoid glycoside (agnuside, aucubin)	Potent antioxidant, cytotoxic & apoptotic activity	99-100
32.	<i>Chrysanthemum coronarium</i>	Chop-suey greens	Asteraceae	All parts	T47D, HRT18, CACO-2, A375.MCF-7	Camphor, α -pinene, lyratyl acetate, β -pinene, chrysanthenylactetae, β -farnesene, germacrene, camphor, perillaldehyde, isoitalicene	Radical scavenging activity relative to the strong antioxidant effect, also exhibits antiproliferative effect	101-102
33.	<i>Cocculus hirsutus</i>	Broom creeper, patalgarudi	Menispermaceae	Aerial parts	MCF-7, Dalton's lymphoma ascites	Flavanoids rutin, liquiritin, quercetin, hirsudioltrilobine, magnofluorine, ginnol, β -sitosterol	Endogenous antioxidant mechanism	103
34.	<i>Cordia alliodora</i>	Indian cherry, bird lime tree	Boraginaceae	Leaves	PC3, MCF-7	Stearic acid, betulin, octacosanol, oleic acid, α -amyriols, lupeol-3rhamnoside.	Antioxidant activity leading to decrease in cell viability	104
35.	<i>Cotinus coggygria</i>	Smoke tree	Anacardiaceae	Leaves	Vero, HeLa	Quercetin, gallic acid, disulfuretin, myricetin, taxifolin, phenolics, flavanoids, tannins, limonene, α -pinene	Lipid peroxidation inhibition	105
36.	<i>Crataegus microphylla</i>	Hawthorn	Rosaceae	Leaves	Vero, HeLa	phenolic compounds, chlorogenic acid, hyperoside and epicatechin	Potent antioxidant effect leading to apoptosis	106
37.	<i>Croton caudatus</i>	Scandent shrub	Euphorbiaceae	Leaves	DL, MCF7, HeLa	ethyl hexatriacontanoate, palmitic acid, stearic acid, zheberiesinol, vanillin, vanillic acid, syringic acid, octacosanoic acid, succinic acid, inosine, isosinensetin	Antioxidant effect, free radical scavenging activity leading to cellular toxicity	106
38.	<i>Delphinium staphisagaria</i>	Stavesacre	Ranunculaceae	Seeds	H56, N2A	Isoazitine, azitine, 19-oxodihydroatisine, dihydroatisine, 22-O-acetyl-19-	Used for hair loss treatment in cancer patients and have cytotoxic and	107-108

						oxodihydroatisine.	angiogenic potential	
39.	<i>Dillenia pentagyna</i>	Dog teak	Dilleniaceae	Stem, bark	DL, MCF7, HeLa	phenolics, flavonoids, tannin, saponin, alkaloid, and terpenoids	apoptosis via NF-Kb pathway through the increase of the bax to Bcl ₂ ratio, leading to fall of MMP & subsequently induced release of cytC , activation of caspase-3 followed by nuclear fragmentation in A549 cells	109
40.	<i>Euphorbia tirucalli</i>	Indian tree spurge, pencil tree, finger tree, petroleum plant	Euphorbiaceae	Leaves, stem	MiPaCa2	Terpenes, sterols, taraxasterol, tirucallol, euphol, α -euphorbol, tiglane, ingenane	Inhibition of protein kinase activity, induce genotoxicity, and changes in antioxidant gene expression	110-111
41.	<i>Ficus racemosa</i>	Goolar, cluster fig	Moraceae	Roots, bark	HL60	α -amyrin acetate , tannin, wax, cerylbehenate,saponingluanol acetate, β -sitosterol (A).	Antioxidant effect observed in-vitro	112-113
42.	<i>Helicteres isora</i>	Enthaniavartani, marodphali,	Sterculiaceae	Plant (Whole)	H60, PN-15,HeLa-B75.	Cucurbitacin b ,gallic acid, vanillin.	Induced apoptosis, generation of ROS & antioxidation activity	114-115
43.	<i>Hibiscus micranthus</i>	Tiny flower hibiscus, rose mallow	Malvaceae	Aerial parts	HepG2, MCF-7	β -sitosterol, phenolic acids, fatty alcohols and acids	Increase oxidative stress, decrease MMP, selectively induced apoptosis via ROS production, though the exact mechanism is not known	116
44.	<i>Hypericum kotschyannum</i>	St. john's wort, goat weed	Hypericaceae	Aerial parts	Vero , HeLa	Hypericin	Apoptosis Induce	117-118
45.	<i>Inulavis cosa</i>	False yellowhead, woody fleabane	Compositae	Flowers	MCF-7, Hep-2	Sesquiterpene lactones (tomentosin, inuviscolide)	Inhibition of cell proliferation	119-121
46.	<i>Jasmiun sambac</i>	Arabian jasmine	Oleaceae	Flowers	MCF-7, Hep-2	benzyl acetate, benzyl alcohol, linalool, geraniol.	Dose dependent tumor cell proliferation inhibition activity	122-123
47.	<i>Lavandulaa ngustifolia</i>	True lavender, narrow-leaved lavender, garden lavender	Lamiaceae	Flowers	MCF-7, Hep-2	Camphor,linalool, terpine-4-ol,	Antiproliferative effect as the main mechanism	124
48.	<i>Leea indica</i>	Bandicoot berry	Vitaceae	Leaves	DU-145, PC-3	Phthalic acid, palmitic acid, ursolic acid, gallic acid, β -sitosterol	Augmentation of bax/Bcl2 ratio, induced mitochondrial-mediated apoptosis	125
49.	<i>Limonium densiflorium</i>	Sea-lavender, marsh-rosemary	Plumbaginaceae	Shoots	A549, DLD-1	3-hydroxycinnamic acid, myricetin, isorhamnetin	Highest antioxidant effect recorded	126
50.	<i>Luffa cylindrica</i>	Sponge guard	Araceae	Aerial parts	MCF-7, Hep-2	Lucyosides C, Lucyosides F, Lucyosides H	Minimise recurrence and metastasis through antiproliferative effect	126-130
51.	<i>Manilkara</i>	Chiku,	Sapotaceae	Flower,	MCF-7	Lupeol acetate,	Induce apoptosis via	131

	<i>zapota</i>	sapodilla, naseberry, chicle gum		leaves		caffeic acid oleanolic acid	antioxidant effect	
52.	<i>Mirabilis jalapa</i>	Marvel of peru, four o'clock flower	Nyctaginaceae	Roots, stem	Hep-2, MCF-7	Phytosterols, β -sitosterols, stigmasterol, brassicasterol, ursolic, oleanolic acid, betulinic acid, Miraxanthin-III,II,V, vulgaxanthin-I	LDHA inhibition activity in silico	132
53.	<i>Morus nigra</i>	Black mulberry, blackberry	Moraceae	Leaves	HeLa, MCF-7	β -Sitosterol, flavanoids	Antioxidant & cytoprotective effect, decrease tumor growth via antiproliferative effect	132
54.	<i>Narcissus tazetta</i>	Daffodil, Chinese sacred lily	Amaryllidaceae	Aerial parts, flowers	MCF-7, Hep-2	Heptanal, myrcene, cineole, ocimene, linalool, nonanal, benzyl acetate, terpineol	Antioxidant as the major mechanism	133
55.	<i>Nepetaita lica</i>	True catnip	Laminaceae	Aerial parts	Vero , HeLa	α -Pinene, 4 α β ,7 α ,7 α β -nepetalactone, 4 α α ,7 α ,7 α β -nepetalactone, α -amorphene, γ -cadinene, cis-calamenene, β -pinene, β -caryophyllene	Effect on lipid peroxidation, functions of oxidative enzymes	134-135
56.	<i>Ocimum sanctum</i>	Tulsi, holy basil	Laminaceae	Leaves	HFS-1080	Oleanolic acid, ursolic acid, linalool, carvacrol, eugenol	Cytotoxic effect on leukemia cell lines	136-138
57.	<i>Oldenlandia corymbosa</i>	Flat-top mille grains, dimond flower	Rubiaceae	Leaves	K562	Geniposide, 6- α -hydroxygeniposide	Significant cytotoxic effect with <i>Parthenium hysterophorus</i> on cancer cell lines	139
58.	<i>Olea europea</i>	Common olive, Indian olive	Oleaceae	Leaves	MCF-7, B16F10, HeLa	Secoiridoids such as oleuropein, ligstroside, methyloleuropein, flavanoids	Antiproliferative effect leading to decrease in tumor growth	140-141
59.	<i>Ononis hirta</i>	Restharrow	Fabaceae	Parts (Aerial)	Breast cancer cell line MCF-7	Flavanoids, terpenoids, phenolic compounds, alkaloidal	Solid tumor necrosis on combination with <i>Bifiridobacteriumlongum</i>	142
60.	<i>Origanum sipyleum</i>	Amaracusgl ed, Turkish oregano	Laminaceae	Aerial parts	Vero , HeLa	γ -Terpinene, pcymene	Inhibition of cancer cell migration, proapoptotic activity, inhibit tumor growth	143
61.	<i>Parthenium hysterophorus</i>	Santa-maria, white top weed, famine weed, congress grass	Asteraceae	Leaves	K562	Germacrene- D, trans- β -ocimene	Considerable potential effect as cytotoxic & antioxidant action, inhibits lipid peroxidation, increase activity of caspase-3,6,9.	144
62.	<i>Phagna lonrupstre</i>	Gnaphalonlo we	Asteraceae	Aerial parts	MCF-7, Hep-2	Dimethylallyl-hydroquinone glucoside,	Antioxidation effect leading to generation of ROS, induces apoptosis	145
63.	<i>Picrorhiza kurroa</i>	Kutki, katuka, picrorhiza	Plantaginaceae	Root	Colo205, Hop62, HT29, SiHa, MIA-PA-CIA-2, DWD, T24, PC3, A549, ZR75-I, A2780, DU145, MCF7, K562	Picoside I, II, d-mannitol, kutkiol, kutkisterol, apocynin, androsin	Regulate the expression of cyclinD1 & CDKs help protect cells against carcinogenesis, inhibition of NF-Kb	146
64.	<i>Piper longum</i>	Pipli, Indian long pepper	Piperaceae	Fruit	Colo205, Hop62, HT29, SiHa, MIA-PA-CIA-2, DWD, T24, PC3,	α -pinene, caryophyllene, Limonene, α -copaene,	Selectively induced caspase independent apoptosis	147

65.	<i>Plectranthus stocksii</i>	Spur-flower	lamiaceae	Stem part and leaves, Flowers	A549, ZR75-I, A2780, DU145, MCF7, K562 MCF-7, RAW 264.7, Caco cell line MCF-7, Hep-2	Ferulic acid, caffeic acid, catechin	Antioxidant effect as the major mechanism	
66.	<i>Populus alba</i>	Silver poplar, white poplar	salicaceae	Flowers	MCF-7, Hep-2	Tremuloidin, populin, chaenomeloidin, salicin, tremulacin, catechol, benzoic acid, salicylol	Strong antiproliferative, cytotoxic & potent antioxidant properties	148
67.	<i>Pterocephalus pulvereus</i>	Wing head shaped	Dipsacaceae	Aerial parts	MCF-7, Hep-2	Flavonoids, alkaloidal, polyphenols, quercetin	Antioxidant effect and antimigratory effect shown against cancer cell lines	149
68.	<i>Rosa damascena</i>	Damask rose, bulgarian rose	Rosaceae	Flowers	Vero, HeLa	Flavonoids, carboxylic acid, glycosides, quercetin	Cell viability decreased	150
69.	<i>Diploaxis harra</i>	Middle east harra, forssk.	Brassicaceae	Whole plant, aerial parts	HCT116, HepG2, MCF7	Quercetin	Antioxidant effect	151
70.	<i>Salvia officinalis</i>	Culinary sage, garden sage	Lamiaceae	Aerial parts	MCF-7, B1610, HeLa	Thujone, β -caryophyllene, cineole, α -humulene, β -pinene, β -thujone, camphor, allo-aromadendrene, borneol, α -pinene	Regulates p53 signalling, cell cycle regulation by G ₁ /S phase arrest	152-153
71.	<i>Saururus chinensis</i>	Lizard's tail	Saururaceae	Roots	MCF-7	Glucosides, hydrolyzable tannins, indolealkaloidal, sitosterols, aristolactam A, ellagic acid, corilagin, lyoniside, neolignans	Antiproliferative activity and antiangiogenic action	154-155
72.	<i>Scorzonera omentosa</i>	Scorzonera	Asteraceae	Aerial parts	Vero, HeLa	Flavanoidglycones, glycosides, phenolic acids, triterpenoids, bibenzyl derivatives	Antioxidant activity leading to apoptosis	156-158
73.	<i>Senecio scandens</i>	Climbing senecio, saiek-hlo	Asteraceae	Leaves	MCF-7, DL, HeLa	Senecainin A, 3-methoxyisonicotinic acid, monoepoxy lignanolate, pinosresinol	High ration of Bax/Bcl-2 promotes apoptosis activity, upregulation of capase-3,9 proteins	159-160
74.	<i>Solanum khasianum</i>	Nightshade	Solanaceae	Fruit	MCF-7, DL, HeLa	Dillapiole, α -cardinol, para-cymene, β -damascenone, α -phellandrene, β -pinene, α -bisabololactate	Cytotoxic effect on cancerous cells and induces cell death and arrest of cell division	161
75.	<i>Citrullus colocynthis</i>	Vine of Sodom, bitter apple, bitter cucumber, desert gourd	Cucurbitaceae	Root, bark, leaves	HepG2 hepatoma cells, colorectal cancer	Cucurbitacin, flavanoids, alkaloidal, phenolic acids	Increase in caspase-3 activity and inhibiting STAT3 function	162-163
76.	<i>Syringa vulgaris</i>	Common lilac	Oleaceae	Seeds	MCF-7 cell line, Hep-2	Iridoids, lignans, phenylpropanoids, phenylethanoids	Oxidative stresses induces apoptosis in cancer cells	164-165
77.	<i>Syzygium cumini</i>	Indian blackberry, jamun	Myrtaceae	Seeds	A2780, MCF7, PC-3, H460	Anthocyanins, glucoside, ellagic acid, isoquercetin, kaemferol, myrecetin	Antiproliferative and antioxidant activity	166-168
78.	<i>Tabernaemontana divaricata</i>	East Indian rosebay, moon beam, coffee rose	Apocynaceae	Flowers	NIH3T3, HeLa	A-amyrin acetate, α -amyryloctadecanoate, taraxasterol acetate, calycosin, formononetin, farnisin	Superoxide anion scavenging activity, reduced GSH levels	169-170

79.	<i>Tecoma stans</i>	Yellow trumpetbush, yellow bells	Bignoniaceae	Leaves, flowers	A549	Alkaloidal, flavanoids, phenylbenzopyrone	inhibit aromatase & disturbs cell division at telophase	171-172
80.	<i>Terucrium polium</i>	Felty germander	Laminaceae	Stems, Leaves	MCF-7, CACO-2, Hep-2,	Agarospinol, caryophyllene, β -caryophyllene, α -humulene, β -bisabolene, β -sesquiphellandrene, dolichodial	Inhibition of DNA oxidation, lipid oxidation	173-175
81.	<i>Tillandsia recurvata</i>	Ball moss	Bromeliaceae	Plant	HL60	Caffeic acid	FMS-like tyrosine kinase 3 get inhibited	176-177
82.	<i>Verbascums inuatum</i>	Wavy-leaf mullein	Scrophulariaceae	Aerial parts, flowers	MCF-7, Hep-2	Verbathasin A, luteolin, 3-O-fucopyranosylsaikogenin F, verbascoside	Apoptosis enhanced	178-179
83.	<i>Vitisa inifera</i>	European wine grape, common grape	Vitaceae	Sap of stem	MCF-7, HeLa	Phenolic compounds, flavanoids	Antioxidant, antiproliferative effect because of epicatechin-3-O-gallate present	180-181
84.	<i>Zea mays</i>	Indian corn	Gaminaea	leaves	Hep2	Quercetin diglucoside, maizenic acid, rutin, chlorogenic acid, hydroxycinnamic acid, flavanoids, saponins, phytosterol, eugenol	Oxidative stress induces apoptosis, upregulation of p53	182-183
85.	<i>Allium savitum</i>	Garlic	Amaryllidaceae	Bulb	Skin, colon, lung, prostate, leukemia, breast cancer	S-allylcysteine, allicin, S-allylmercapto-L-cysteine	High radical scavenging activity, antiproliferative growth, inhibits tumor growth	184-186
86.	<i>Acronychia baueri</i>	Chakkimaram, mootanari, aspen	Rutaceae	Bark	Carcinoma, colon cancer	Alkaloids (normelicopidine, melicopine, acronycine), triterpenelupeol	Antiproliferative and antioxidant effect	187
87.	<i>Alstonia boonei</i>	Cheese wood, pattern wood, stool wood	Apocynaceae	Stem, bark, leaf, root	Pancreas, lung, prostate, colon cancer	Echitamine, eugenol, 1,2 benzene dicarboxylic acid, alstibooinine	Antiproliferative and cytotoxic effect	188-189
88.	<i>Thymus vulgaris</i>	Thyme	Lamiaceae	Bulb, leaves	MCF-7 cell lines, HeLa	Diallyl disulphide, diallyltrisulphide, allicin, tannins, triterpenes, styerols, flavonoids, glycosides	By controlling interferon signalling, biosynthesis of N-glycxan	190-191
89.	<i>Tithonia diversifolia</i>	Gaint Mexican sunflower	Composiae	leaves	HL-60, U373, Col2 colon cancer cells	Tagitinin C, 1- β -methoxydiversifolin, 3-O-methyl ether, 1- β - & 2- α -epoxytagitinin C	Inhibit cell proliferation, induces cell death	192

Clinical Trials on Isolated Constituents (<https://clinicaltrials.gov/>): Herbal drugs are integral to the health care system. Clinical trials are essential to assess the risks associated with herbal drugs.

In clinical trials, herbal medicines must be proven safe and effective before their effectiveness can be determined. Clinical studies about herbal anticancer are interventional, observational, and other types mentioned in the literature. In this article, we consider the clinical studies related to herbal drugs used in cancer.

An early phase 1 (NCT01568996) study on Sulforaphane was done to determine the chemopreventive action and modulate the melanoma.

Another phase 1 (NCT03980509) study is performed on curcumin to study its effect on breast cancer. Effect of quercetin (NCT03980509) has been tested on polyphenols uptake subject's suffering from prostate cancer. Similarly, clinical trials on the herbal drug used as an anticancer drug are listed in **Table 4**.

TABLE 4: CLINICAL TRIAL ON HERBAL DRUGS

Description	Type of study	Phase of study	Design of study	Age (years)	Subject participated	Status of study	Reference
Traditional Chinese Medicine in breast and other cancers	Interventional	Phase 2	Randomized	≥21	80	Recruiting	NCT04104113
Teng-Long-Bu-Zhong-Tang herbal along With Chemotherapy in Colorectal cancer patients	Interventional	Phase 1	Randomized	18-70	72	Completed	NCT01975454
Herbal drug products in patients of breast cancers for reduction of dermatitis induced by radiation	Interventional	Phase 2	Randomized	≥20	150	Completed	NCT02922244
TPE-1 herbal formula effect in breast cancer patients	Interventional	Not Applicable	Randomized	18-70	60	Completed	NCT01142479
KD018 and Sorafenib in patients with hepatic carcinoma	Interventional	Phase 2	Not applicable	≥18	18	Completed	NCT01666756
Dendrobium Huoshanense Granules effects in rectal cancer patients	Interventional	Phase 3	Randomized	18-70	210	Recruiting	NCT04394598
Traditional Chinese Medicine regulation in cancer patients	Interventional	Phase 1	Not applicable	≥20	300	Recruiting	NCT04438564
Da Huang Gang Tsao Tang effect for improving appetite in patients of cancer at late-stage	Interventional	Phase 2	Not applicable	≥20	4	Completed	NCT01503346
Effect of Chinese herbal therapy in women of breast cancer undergoing chemotherapy	Interventional	Not Applicable	Randomized	≥18	Unknown	Completed	NCT00028964
Herbal drug therapy in patients of prostate cancer	Interventional	Phase 2	Not applicable	≥18	43	Completed	NCT00669656
Effect of ACAPH in patients of Intraepithelial neoplasia having smoking history	Interventional	Phase 1	Randomized	45-75	90	Completed	NCT00522197
Effect of combination of chinese and western medicine in cancer patients having constipation	Interventional	Phase 2	Randomized	≥18	60	Completed	NCT02795390
TCM-TSKSR effects in patients having cancer at different stages	Interventional	Phase 2	Randomized	18-75	400	Recruiting	NCT03716518
Herbal Mouth rinse in patients of cancer having mucositis	Interventional	Phase 2	Randomized	18-89	50	Completed	NCT01898091
Angelica Sinensis effect in prostate cancer for treatment of hot fleshes	Interventional	Phase 2	Randomized	≥18	44	Completed	NCT00199485
Traditional and complementary medicine effect in Ovarian Cancer	Interventional	Phase 3	Not applicable	≥18	28	Completed	NCT01419210
Food supplements in cancer threapies	Interventional	Phase 2	Non-Randomized	≥18	60	Recruiting	NCT04474951
Effect of KD018 in colorectal cancer	Interventional	Phase 4	Randomized	≥18	33	Completed	NCT00730158
Sho-Saiko in hepatic cancer	Interventional	Not Applicable	Not applicable	≥18		Completed	NCT00040898
Traditional chinese medicine effect in triple-negative breast cancer	Interventional	Not Applicable	Randomized	18-70	200	Recruiting	NCT04403529
Effect of SH003 in Solid Cancer	Interventional	Phase 2	Sequential Assignment	≥19	11	Completed	NCT03081819
Blue Citrus in breast cancer	Interventional	Phase 2	Randomized	≥18	30	Completed	NCT00702858
Dietary supplement effect in breast cancer	Observational	Phase 3	Cohort	≥18	200	Completed	NCT03959618
Fuzheng Yiliu effect in chemotherapy patients	Interventional	Phase 1	Non-Randomized	18-75	189	Recruiting	NCT04459754
Chemotherapy and Traditional chinese medicine effect on Lung Cancer	Observational	Not Applicable	Not Applicable	65-80	82	Completed	NCT01780181
Sipjeondaebo-tang in patients of cancer having anorexia	Interventional	Not Applicable	Randomized	20-80	32	Completed	NCT02468141

Siliphos in hepatic cancer patients	Interventional	Phase 2	Non-Randomized	≥18	3	Completed	NCT01129570
Artemi Coffee in Patients of ovarian cancer at advanced stage	Interventional	Not Applicable	Non-Randomized	≥18	18	Recruiting	NCT04805333
Oligo-Fucoidan effect in hepatic cancer	Interventional	Not Applicable	Randomized	≥18	100	Recruiting	NCT04066660
Chamomile gel in prevention of oral mucositis induced by chemotherapy	Interventional	Phase 1	Randomized	20-70	45	Recruiting	NCT04317183
Effect of curcumin on radiation induced dermatitis among the person suffering from breast cancer	Interventional	Phase 1	Randomized	21	35	Completed	NCT01042938
Effect of curcumin on pancreatic cancer was studied.	Interventional	Phase 2	Not applicable	18	50	Completed	NCT00094445
Resveratrol effect on patients with colon cancer.	Interventional	Phase 2	Not applicable	18	11	Completed	NCT00256334
Artesunate effect in Colorectal Cancer at different stages.	Interventional	Phase 2	Randomized	18-70	200	Recruiting	NCT03093129
The effect of Chinese herbs on syndrome differentiation with reduced side effects and increased antitumour effect was studied.	Interventional	Phase 2	Randomized	18	200	Unknown	NCT02737735
This study aims to know the effect of berberine to prevent colorectal adenomas in patients with previous colorectal cancer.	Interventional	Phase 1	Randomized	18-80	1000	Unknown	NCT03281096
To study the effect of lycopene in colorectal carcinoma patients.	Interventional	Phase 2	Randomized	18	28	Completed	NCT03167268
This study aims to know the chemoprevention effect of Sulforaphane in lung cancer in former smokers.	Interventional	Phase 1	Randomized	55-75	72	Recruiting	NCT03232138
The effect of Chinese herbs on syndrome differentiation with reduced side effects and increased antitumour effect was studied	Interventional	Phase 2 Phase 3	Randomized	18	200	Unknown	NCT02737735
This study aims to know the effect of berberine to prevent colorectal adenomas in patients with previous colorectal cancer.	Interventional	Phase 2	Randomized	18-80	1000	Unknown	NCT03281096
This study aims to know the chemoprevention effect of Sulforaphane in lung cancer in former smokers.	Interventional	Phase 2	Randomized	55-75	72	Recruiting	NCT03232138
To study the effect of epigallocatechin gallate colorectal cancer patients.	Interventional	Early Phase 1	Randomized	≥18	50	Recruiting	NCT02891538
A Phase 1 Dose Escalation of Artemi Coffee in Patients With Advanced Ovarian Cancer	Interventional	Phase 1	Non-Randomized	≥18	18	Recruiting	NCT04805333
Kanglaite (Coix Seed Oil) in head and neck Cancer	Interventional	Phase 2	Not applicable	18-80	53	Completed	NCT03101514
Effect of Kanglaite Injection with gemcitabine in pancreatic cancer	Interventional	Phase 2	Randomized	18-75	85	Completed	NCT00733850

CONCLUSION: Throughout the world, cancer threatens millions of lives each year. Aside from affecting a patient's physical health and quality of life, treatments such as surgery and chemotherapy do not dramatically improve the chances of survival. Even though modern medicine has greatly influenced people's lives and synthetic drugs have

become more readily available; still, most of the people depend on plant remedies. WHO data shows that greater than 75% of the population rely on plant remedies or extracts for health care needs. Research results have directed to expanding plant-derived products after achieving excellent results and are now conducting clinical trials. An effective

way to inhibit cancer cells is with plant-derived anticancer agents, making them highly valuable. To meet demand and remain sustainable, herbal drugs must be exploited efficiently. The current review may provide researchers with a great deal of information about herbs, their effects on disease, and the safety of using them.

ACKNOWLEDGEMENT: None

CONFLICTS OF INTEREST: The authors declare no conflict of interest.

REFERENCES:

- Kooti W, Servatyari K, Behzadifar M, Asadi-Samani M, Sadeghi F and Nouri B: Effective Medicinal Plant in Cancer Treatment, Part 2: Review Study. *Journal of Evidence-based Complementary & Alternative Medicine* [Internet]. 2017 Oct 1 [cited 2022 Jan 15]; 22(4):982. Available from: <https://pubmed.ncbi.nlm.nih.gov/26297173/>
- Muniyandi K, George E, Mudili V, Kalagatur NK, Anthuvan AJ and Krishna K: Antioxidant and anticancer activities of *Plectranthus stocksii* Hook. f. leaf and stem extracts. *Agriculture and Natural Resources* 2017; 51(2): 63–73.
- Ghagane SC, Puranik SI, Kumbar VM, Nerli RB, Jalalpure SS and Hiremath MB: *In-vitro* antioxidant and anticancer activity of *Leea indica* leaf extracts on human prostate cancer cell lines. *Integrative medicine research* [Internet]. 2017 Mar [cited 2022 Jan 15]; 6(1):79–87. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/28462147>
- Cytotoxic and Anticancer Activity of *F. Racemosa* Fruit Extract on MCF7 Human Breast Cancer Cell Line by SRB Method - JAR [Internet]. [cited 2022 Jan 15]. Available from: <https://journalanimalresearch.com/Journal/abstract/id/MTI5MQ==>
- Kola P, Metowogo K, Kantati YT, Lawson-Evi P, Kpemissi M and El-Hallouty SM: Ethnopharmacological survey on medicinal plants used by traditional healers in central and kara regions of togo for antitumor and chronic wound healing effects. *Evidence-based Complementary and Alternative Medicine* 2020; 2020.
- Newman DJ and Cragg GM: Natural products as sources of new drugs over the last 25 years. *Journal of natural products* [Internet]. 2007 Mar [cited 2022 Jan 15]; 70(3): 461–77. Available from: <https://pubmed.ncbi.nlm.nih.gov/17309302/>
- Solowey E, Lichtenstein M, Sallon S, Paavilainen H, Solowey E and Lorberboum-Galski H: Evaluating medicinal plants for anticancer activity. *TheScientificWorldJournal* [Internet]. 2014 [cited 2022 Jan 15]; 2014. Available from: <https://pubmed.ncbi.nlm.nih.gov/25478599/>
- Taneja SC and Qazi GN: Bioactive Molecules in Medicinal Plants: A Perspective on their Therapeutic Action. *Drug Discovery and Development* [Internet]. 2006 May 18 [cited 2022 Jan 15]; 1–50. Available from: <https://onlinelibrary.wiley.com/doi/full/10.1002/9780470085226.ch17>
- Pratheeshkumar P, Sreekala C, Zhang Z, Budhraj A, Ding S and Son YO: Cancer Prevention with Promising Natural Products: Mechanisms of Action and Molecular Targets. *Anticancer agents in medicinal chemistry* [Internet]. 2012 Nov 11 [cited 2022 Jan 15]; 12(10):1159. Available from: <https://pubmed.ncbi.nlm.nih.gov/25478599/>
- Noh S, Choi E, Hwang CH, Jung JH, Kim SH and Kim B: Dietary Compounds for Targeting Prostate Cancer. *Nutrients* [Internet]. 2019 Oct 1 [cited 2022 Jan 15]; 11(10). Available from: <https://pubmed.ncbi.nlm.nih.gov/3535786/>
- Mahmudur Rahman A: A Review on Medicinal Plants with Anticancer Activity Available In Bangladesh. 2018 [cited 2022 Jan 15]; Available from: <http://www.ethnobotanybd.com/>
- Adair TH, Montani J-P. Overview of Angiogenesis. 2010 [cited 2022 Jan 15]; Available from: <https://www.ncbi.nlm.nih.gov/books/NBK53238/>
- Fallah A, Sadeghinia A, Kahroba H, Samadi A, Heidari HR and Bradaran B: Therapeutic targeting of angiogenesis molecular pathways in angiogenesis-dependent diseases. *Biomedicine & Pharmacotherapy* 2019; 110: 775–85.
- Ziyad S and Iruela-Arispe ML: Molecular Mechanisms of Tumor Angiogenesis. *Genes & Cancer* [Internet]. 2011
- Kooti W, Servatyari K, Behzadifar M, Asadi-Samani M, Sadeghi F and Nouri B: Effective Medicinal Plant in Cancer Treatment, Part 2: Review Study. *Journal of Evidence-based Complementary & Alternative Medicine* [Internet]. 2017 Oct 1 [cited 2022 Jan 15]; 22(4):982. Available from: <https://pubmed.ncbi.nlm.nih.gov/26297173/>
- Mousavi SM, Gouya MM, Ramazani R, Davanlou M, Hajsadeghi N and Seddighi Z: Cancer incidence and mortality in Iran. *Annals of oncology : official journal of the European Society for Medical Oncology* [Internet]. 2009 [cited 2022 Jan 15]; 20(3):556–63. Available from: <https://pubmed.ncbi.nlm.nih.gov/19073863/>
- Preventing chronic diseases : a vital investment : WHO global report [Internet]. [cited 2022 Jan 15]. Available from: <https://apps.who.int/iris/handle/10665/43314>
- Greenwell M and Rahman PKSM: Medicinal Plants: Their Use in Anticancer Treatment. *International journal of pharmaceutical sciences and research* [Internet]. 2015 Oct 1 [cited 2022 Jan 15]; 6(10):4103. Available from: <https://pubmed.ncbi.nlm.nih.gov/26297173/>
- Kuruppu AI, Paranagama P and Goonasekara CL: Medicinal plants commonly used against cancer in traditional medicine formulae in Sri Lanka. *Saudi pharmaceutical journal : SPJ : the official publication of the Saudi Pharmaceutical Society* [Internet]. 2019 May 1 [cited 2022 Jan 15]; 27(4):565–73. Available from: <https://pubmed.ncbi.nlm.nih.gov/31061626/>
- Cooper GM: The Development and Causes of Cancer. 2000 [cited 2022 Jan 15]; Available from: <https://www.ncbi.nlm.nih.gov/books/NBK9963/>
- Saranya K, Manivasagan V, Kanakadurga R, Babu VPM and Ramesh Babu NG: A survey on anticancer properties of indian medicinal plants-a broad spectrum analysis. *International Journal of Pharmaceutical Sciences and Research* [Internet]. 2019 [cited 2022 Jan 15]; (8):3635. Available from: <https://ijpsr.com/bft-article/a-survey-on-anticancer-properties-of-indian-medicinal-plants-a-broad-spectrum-analysis/>
- Poonam S and Chandana M: A Review on Anticancer Natural Drugs.
- (PDF) Anticancer activity of some medicinal plants from high altitude evergreen elements of Indian Western Ghats [Internet]. [cited 2022 Jan 15]. Available from: https://www.researchgate.net/publication/230559795_Anti_cancer_activity_of_some_medicinal_plants_from_high_altitude_evergreen_elements_of_Indian_Western_Ghats
- Asadi-Samani M, Kooti W, Aslani E and Shirzad H: A systematic review of iran's medicinal plants with anticancer effects. *Journal of evidence-based complementary & alternative medicine* [Internet]. 2016 Apr 1 [cited 2022 Jan 15]; 21(2):143–53. Available from: <https://pubmed.ncbi.nlm.nih.gov/26297173/>

- Dec [cited 2022 Jan 15]; 2(12):1085. Available from: [/pmc/articles/PMC3411131/](#)
24. Lu K, Bhat M and Basu S: Plants and their active compounds: natural molecules to target angiogenesis. *Angiogenesis* [Internet]. 2016 Jul 1 [cited 2022 Jan 15]; 19(3): 287. Available from: [/pmc/articles/PMC4930694/](#)
 25. Lew DJ: Cell Cycle. *Brenner's Encyclopedia of Genetics: Second Edition* 2013; 456–64.
 26. Introduction to the Cell Cycle. *Cell Biology* 2017; 697–711.
 27. Wang M, Zhao J, Zhang L, Wei F, Lian Y and Wu Y: Role of tumor microenvironment in tumorigenesis. *Journal of Cancer* [Internet]. 2017 [cited 2022 Jan 15]; 8(5):761. Available from: [/pmc/articles/PMC5381164/](#)
 28. Hanahan D and Weinberg RA: Hallmarks of Cancer: The Next Generation. *Cell* 2011; 144(5): 646–74.
 29. Cao W, Li J, Yang K and Cao D: An overview of autophagy: Mechanism, regulation and research progress. *Bulletin du Cancer* 2021; 108(3): 304–22.
 30. Hayat MA: Overview of Autophagy. *Autophagy: Cancer, Other Pathologies Inflammation Immunity Infection and Aging* 2016; 3–84.
 31. Todde V, Veenhuis M and van der Klei IJ: Autophagy: Principles and significance in health and disease. *Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease* 2009; 1792(1): 3–13.
 32. Bhat P, Kriel J, Shubha Priya B, Basappa, Shivananju NS and Loos B: Modulating autophagy in cancer therapy: Advancements and challenges for cancer cell death sensitization. *Biochemical pharmacology* [Internet]. 2018 Jan 1 [cited 2022 Jan 15]; 147: 170–82. Available from: <https://pubmed.ncbi.nlm.nih.gov/29203368/>
 33. Kocaturk NM, Akkoc Y, Kig C, Bayraktar O, Gozuacik D and Kutlu O: Autophagy as a molecular target for cancer treatment. *European Journal of Pharmaceutical Sciences*. 2019; 134: 116–37.
 34. Al-Bari MAA, Ito Y, Ahmed S, Radwan N, Ahmed HS and Eid N: Targeting Autophagy with Natural Products as a Potential Therapeutic Approach for Cancer. *International Journal of Molecular Sciences* [Internet]. 2021 Sep 1 [cited 2022; 22(18). Available from: [/pmc/articles/PMC8467030/](#)
 35. Wang H, Oo Khor T, Shu L, Su ZY, Fuentes F and Lee JH: Plants vs. cancer: a review on natural phytochemicals in preventing and treating cancers and their druggability. *Anticancer agents in medicinal chemistry* [Internet]. 2012 Nov 11 [cited 2022 Jan 15]; 12(10):1281–305. Available from: <https://pubmed.ncbi.nlm.nih.gov/22583408/>
 36. Yin SY, Wei WC, Jian FY and Yang NS: Therapeutic applications of herbal medicines for cancer patients. *Evidence-based complementary and alternative medicine: eCAM* [Internet]. 2013 [cited 2022 Jan 15]; 2013. Available from: <https://pubmed.ncbi.nlm.nih.gov/23956768/>
 37. Shukla Y and Singh M: Cancer preventive properties of ginger: a brief review. *Food and chemical toxicology: an international journal published for the British Industrial Biological Research Association* [Internet]. 2007 May [cited 2022 Jan 15]; 45(5): 683–90. Available from: <https://pubmed.ncbi.nlm.nih.gov/17175086/>
 38. Williamson EM: *Dabur Research Foundation., Dabur Ayurved Limited. Major herbs of ayurveda.* Edinburgh New York Churchill Livingstone 2002; 361.
 39. Turrini E, Catanzaro E, Muraro MG, Governa V, Trella E and Mele V: *Hemidesmus indicus* induces immunogenic death in human colorectal cancer cells. *Oncotarget* [Internet]. 2018 May 1 [cited 2022 Jan 15]; 9(36):24443–56. Available from: <https://explore.elsevier.com/search/publication?pid=10.18632/oncotarget.25325>
 40. Das S and Singh Bisht S: The bioactive and therapeutic potential of *Hemidesmus indicus* R. Br. (Indian Sarsaparilla) root. *Phytotherapy research: PTR* [Internet]. 2013 Jun [cited 2022 Jan 15]; 27(6):791–801. Available from: <https://pubmed.ncbi.nlm.nih.gov/22887725/>
 41. Nandy S, Mukherjee A, Pandey DK, Ray P and Dey A: Indian Sarsaparilla (*Hemidesmus indicus*): Recent progress in research on ethnobotany, phytochemistry and pharmacology. *Journal of Ethnopharmacology* 2020; 254.
 42. Turrini E, Calcabrini C, Tacchini M, Efferth T, Sacchetti G and Guerrini A: *In-vitro* study of the cytotoxic, cytostatic, and antigenotoxic profile of *Hemidesmus indicus* (L.) r.br. (apocynaceae) crude drug extract on t lymphoblastic cells. *Toxins* 2018; 10(2).
 43. Dharmadasa RM, Premakumara GAS, Hettiarachchi PL and Ratnasooriya WD: Cytotoxicity and *in-vivo* antimalarial activity of aqueous whole plant extract of *Munronia pinnata* (Wall.) Theob. (Meliaceae) in mice. *Research Journal of Medicinal Plant* 2012; 6(3): 267–73.
 44. Kuruppu AI, Paranagama P and Goonasekara CL: Medicinal plants commonly used against cancer in traditional medicine formulae in Sri Lanka. *Saudi Pharmaceutical Journal: SPJ* [Internet]. 2019 May 1 [cited 2022 Jan 15]; 27(4):565. Available from: [/pmc/articles/PMC6488922/](#)
 45. Prashith Kekuda T, Sahana B, Saema Noorain G and Raghavendra H: A Comprehensive Review on the Ethnobotanical uses, Phytochemistry and Pharmacological Activities of *Smilax zeylanica* L. (Smilacaceae). Available online www.jocpr.com *Journal of Chemical and Pharmaceutical Research* [Internet]. 2018 [cited 2022 Jan 15]; 10(9): 55–63. Available from: www.jocpr.com
 46. Saha S and Ghosh S: *Tinospora cordifolia*: One plant, many roles. *Ancient Science of Life* [Internet]. 2012 [cited 2022 Jan 15]; 31(4):151. Available from: [/pmc/articles/PMC3644751/](#)
 47. Biswasroy P, Panda S, Das C, Das D, Kar DM, Ghosh G. *Tinospora cordifolia*— a plant with spectacular natural immunobooster. *Research Journal of Pharmacy and Technology* 2020; 13(2): 1035–8.
 48. Lindamulage IKS and Soysa P: Evaluation of anticancer properties of a decoction containing *Adenanthera pavonina* L. and *Thespesia populnea* L. *BMC Complementary and Alternative Medicine* [Internet]. 2016 Feb 20 [cited 2022 Jan 15]; 16(1). Available from: [/pmc/articles/PMC4761162/](#)
 49. Phanse MA, Patil MJ and Abbulu K: review on pharmacological studies of *Thespesia populnea* linn.
 50. Kuruppu AI, Paranagama P and Goonasekara CL: Medicinal plants commonly used against cancer in traditional medicine formulae in Sri Lanka. *Saudi Pharmaceutical Journal* [Internet]. 2019 May 1 [cited 2022 Jan 15]; 27(4):565–73. Available from: https://www.researchgate.net/publication/331007535_Medicinal_plants_commonly_used_against_cancer_in_traditional_medicine_formulae_in_Sri_Lanka
 51. Liu X, Zhao M, Wu K, Chai X, Yu H and Tao Z: Immunomodulatory and anticancer activities of phenolics from emblica fruit (*Phyllanthus emblica* L.). *Food Chemistry* [Internet]. 2012 Mar 15 [cited 2022 Jan 15]; 2(131):685–90. Available from: <https://www.infona.pl/resource/bwmeta1.element.elsevier-9b05e080-447c-3fcb-815e-2977691fb0cf>
 52. De A, De A, Papasian C, Hentges S, Banerjee S and Haque I: *Embolica officinalis* extract induces autophagy and

- inhibits human ovarian cancer cell proliferation, angiogenesis, growth of mouse xenograft tumors. PloS one [Internet]. 2013 Aug 15 [cited 2022 Jan 15]; 8(8). Available from: <https://pubmed.ncbi.nlm.nih.gov/24133573/>
53. Mishra S, Aeri V, Gaur PK and Jachak SM: Phytochemical, therapeutic, and ethnopharmacological overview for a traditionally important herb: *Boerhavia diffusa* Linn. BioMed research international [Internet]. 2014 [cited 2022 Jan 15]; 2014. Available from: <https://pubmed.ncbi.nlm.nih.gov/24949473/>
 54. Liu WK, Ho JCK, Cheung FWK, Liu BPL, Ye WC and Che CT: Apoptotic activity of betulinic acid derivatives on murine melanoma B16 cell line. European Journal of Pharmacology 2004; 498(1-3): 71-8.
 55. Cichewicz RH and Kouzi SA: Chemistry, biological activity, and chemotherapeutic potential of betulinic acid for the prevention and treatment of cancer and hiv infection. Medicinal Research Reviews 2004; 24(1): 90-114.
 56. Taylor WC: Cytotoxic Diterpenoid Constituents from *Andrographis paniculata* Ness. Leaves. Science Asia 1992; 18(4): 187.
 57. Ajaya Kumar R, Sridevi K, Vijaya Kumar N, Nanduri S and Rajagopal S: Anticancer and immunostimulatory compounds from *Andrographis paniculata*. Journal of Ethnopharmacology 2004; 92(2-3): 291-5.
 58. Rajagopal S, Kumar RA, Deevi DS, Satyanarayana C and Rajagopalan R: Andrographolide, a potential cancer therapeutic agent isolated from *Andrographis paniculata*. Journal of experimental therapeutics & oncology [Internet]. 2003 May [cited 2022 Jan 15]; 3(3): 147-58. Available from: <https://pubmed.ncbi.nlm.nih.gov/14641821/>
 59. Islam MT, Ali ES, Uddin SJ, Islam MA, Shaw S and Khan IN: Andrographolide, a diterpene lactone from *Andrographis paniculata* and its therapeutic promises in cancer. Cancer Letters 2018; 420: 129-45.
 60. Naidoo DB, Chuturgoon AA, Phulukdaree A, Guruprasad KP, Satyamoorthy K and Sewram V: *Centella asiatica* modulates cancer cachexia associated inflammatory cytokines and cell death in leukaemic THP-1 cells and peripheral blood mononuclear cells (PBMC's). BMC Complementary and Alternative Medicine [Internet]. 2017 Aug 1 [cited 2022 Jan 15]; 17(1). Available from: <https://pubmed.ncbi.nlm.nih.gov/285540453/>
 61. Babu TD, Kuttan G and Padikkala J: Cytotoxic and anti-tumour properties of certain taxa of Umbelliferae with special reference to *Centella asiatica* (L.) Urban. Journal of ethnopharmacology [Internet]. 1995 Aug 11 [cited 2022 Jan 15]; 48(1):53-7. Available from: <https://pubmed.ncbi.nlm.nih.gov/8569247/>
 62. Wu FE, Gu ZM, Zeng L, Zhao GX, Zhang Y and McLaughlin JL: Two new cytotoxic monotetrahydrofuran Annonaceous acetogenins, anomuricins A and B, from the leaves of *Annona muricata*. Journal of natural products [Internet]. 1995 [cited 2022 Jan 15]; 58(6):830-6. Available from: <https://pubmed.ncbi.nlm.nih.gov/7673926/>
 63. Chang FR and Wu YC: Novel cytotoxic annonaceous acetogenins from *Annona muricata*. Journal of natural products [Internet]. 2001 [cited 2022 Jan 15]; 64(7): 925-31. Available from: <https://pubmed.ncbi.nlm.nih.gov/11473425/>
 64. Liaw CC, Chang FR, Lin CY, Chou CJ, Chiu HF and Wu MJ: New cytotoxic monotetrahydrofuran annonaceous acetogenins from *Annona muricata*. Journal of Natural Products 2002; 65(4): 470-5.
 65. Wall ME and Wani MC: Camptothecin. Discovery to clinic. Annals of the New York Academy of Sciences [Internet]. 1996 [cited 2022; 803:1-12. Available from: <https://pubmed.ncbi.nlm.nih.gov/8993495/>
 66. Puri SG, Verma V, Amna T, Qazi GN and Spiteller M: An endophytic fungus from *Nothapodytes foetida* that produces camptothecin. Journal of natural products [Internet]. 2005 Dec [cited 2022 Jan 15]; 68(12): 1717-9. Available from: <https://pubmed.ncbi.nlm.nih.gov/16378360/>
 67. Wall ME and Wani MC: Antineoplastic agents from plants. Annual review of pharmacology and toxicology [Internet]. 1977 [cited 2022 Jan 15]; 17: 117-32. Available from: <https://pubmed.ncbi.nlm.nih.gov/326159/>
 68. Slichenmyer WJ, Rowinsky EK, Donehower RC and Kaufmann SH: The current status of camptothecin analogues as antitumor agents. Journal of the National Cancer Institute [Internet]. 1993 Feb 17 [cited 2022 Jan 15]; 85(4): 271-91. Available from: <https://pubmed.ncbi.nlm.nih.gov/8381186/>
 69. Jayaprakasam B, Zhang Y, Seeram NP and Nair MG: Growth inhibition of human tumor cell lines by withanolides from *Withania somnifera* leaves. Life sciences [Internet]. 2003 Nov 21 [cited 2022 Jan 15]; 74(1):125-32. Available from: <https://pubmed.ncbi.nlm.nih.gov/14575818/>
 70. Singh N, Yadav SS, Rao AS, Nandal A, Kumar S and Ganaie SA: Review on anticancerous therapeutic potential of *Withania somnifera* (L.) Dunal. Journal of Ethnopharmacology 2021; 270.
 71. Mukherjee PK, Banerjee S, Biswas S, Das B, Kar A and Katiyar CK: *Withania somnifera* (L.) Dunal - Modern perspectives of an ancient Rasayana from Ayurveda. Journal of Ethnopharmacology 2021; 264.
 72. Malik F, Kumar A, Bhushan S, Khan S, Bhatia A and Suri KA: Reactive oxygen species generation and mitochondrial dysfunction in the apoptotic cell death of human myeloid leukemia HL-60 cells by a dietary compound withaferin A with concomitant protection by N-acetyl cysteine. Apoptosis: an international journal on programmed cell death [Internet]. 2007 Nov [cited 2022 Jan 15]; 12(11):2115-33. Available from: <https://pubmed.ncbi.nlm.nih.gov/17874299/>
 73. Malik F, Singh J, Khajuria A, Suri KA, Satti NK and Singh S: A standardized root extract of *Withania somnifera* and its major constituent withanolide-A elicit humoral and cell-mediated immune responses by up regulation of Th1-dominant polarization in BALB/c mice. Life sciences [Internet]. 2007 Mar 27 [cited 2022 Jan 15]; 80(16): 1525-38. Available from: <https://pubmed.ncbi.nlm.nih.gov/17336338/>
 74. Shashi B, Jaswant S, Madhusudana RJ, Kumar SA and Nabi QG: A novel lignan composition from *Cedrus deodara* induces apoptosis and early nitric oxide generation in human leukemia Molt-4 and HL-60 cells. Nitric oxide: biology and chemistry [Internet]. 2006 Feb [cited 2022 Jan 15]; 14(1): 72-88. Available from: <https://pubmed.ncbi.nlm.nih.gov/16288976/>
 75. Singh SK, Shanmugavel M, Kampasi H, Singh R, Mondhe DM and Rao JM: Chemically standardized isolates from *Cedrus deodara* stem wood having anticancer activity. Planta medica [Internet]. 2007 Jun [cited 2022 Jan 15]; 73(6): 519-26. Available from: <https://pubmed.ncbi.nlm.nih.gov/17534788/>
 76. Bhushan S, Kumar A, Malik F, Andotra SS, Sethi VK and Kaur IP: A triterpenediol from *Boswellia serrata* induces apoptosis through both the intrinsic and extrinsic apoptotic

- pathways in human leukemia HL-60 cells. Apoptosis: an international journal on programmed cell death [Internet]. 2007 Oct [cited 2022 Jan 15]; 12(10): 1911–26. Available from: <https://pubmed.ncbi.nlm.nih.gov/17636381/>
77. Syrovets T, Gschwend JE, Büchele B, Laumonier Y, Zugmaier W and Genze F: Inhibition of IkappaB kinase activity by acetyl-boswellic acids promotes apoptosis in androgen-independent PC-3 prostate cancer cells in vitro and in vivo. The Journal of biological chemistry [Internet]. 2005 Feb 18 [cited 2022 Jan 15]; 280(7): 6170–80. Available from: <https://pubmed.ncbi.nlm.nih.gov/15576374/>
 78. Liu JJ, Nilsson Å, Oredsson S, Badmaev V, Zhao WZ and Duan RD: Boswellic acids trigger apoptosis via a pathway dependent on caspase-8 activation but independent on Fas/Fas ligand interaction in colon cancer HT-29 cells. Carcinogenesis [Internet]. 2002 Dec 1 [cited 2022 Jan 15]; 23(12): 2087–93. Available from: <https://pubmed.ncbi.nlm.nih.gov/12507932/>
 79. Zhao W, Entschladen F, Liu H, Niggemann B, Fang Q and Zaenker KS: Boswellic acid acetate induces differentiation and apoptosis in highly metastatic melanoma and fibrosarcoma cells. Cancer detection and prevention [Internet]. 2003 [cited 2022 Jan 15]; 27(1): 67–75. Available from: <https://pubmed.ncbi.nlm.nih.gov/12600419/>
 80. Syrovets T, Büchele B, Gedig E, Slupsky JR and Simmet T: Acetyl-boswellic acids are novel catalytic inhibitors of human topoisomerases I and IIalpha. Molecular pharmacology [Internet]. 2000 [cited 2022 Jan 15]; 58(1): 71–81. Available from: <https://pubmed.ncbi.nlm.nih.gov/10860928/>
 81. Hussain A, Sharma C, Khan S, Shah K and Haque S: Aloe vera inhibits proliferation of human breast and cervical cancer cells and acts synergistically with cisplatin. Asian Pacific journal of cancer prevention: APJCP [Internet]. 2015 [cited 2022 Jan 15]; 16(7): 2939–46. Available from: <https://pubmed.ncbi.nlm.nih.gov/25854386/>
 82. Tavares L, Fortalezas S, Carrilho C, McDougall GJ, Stewart D and Ferreira RB: Antioxidant and antiproliferative properties of strawberry tree tissues. Journal of Berry Research 2010; 1(1): 3–12.
 83. Akindele AJ, Wani Z, Mahajan G, Sharma S, Aigbe FR and Satti N: Anticancer activity of *Aristolochia ringens* Vahl. (Aristolochiaceae). Journal of Traditional and Complementary Medicine 2015; 5(1): 35–41.
 84. Jarial R, Thakur S, Sakinah M, Zularisam AW, Sharad A and Kanwar SS: Potent anticancer, antioxidant and antibacterial activities of isolated flavonoids from *Asplenium nidus*. Journal of King Saud University - Science 2018; 30(2): 185–92.
 85. Yan and Asmah R: Anti-proliferation of MDA-MB-231 Cells by Averrhoa bilimbi Extract is Associated with G0/G1 Perturbation and Mitochondria-mediated Apoptosis Independent of p53. International Food Research Journal 2017; 24(3): 1331–7.
 86. Lee KH: Anticancer drug design based on plant-derived natural products. Journal of Biomedical Science 1999; 6(4): 236–50.
 87. Moga MA, Bălan A, Anastasiu CV, Dimienescu OG, Neculoiu CD and Gavriș C: An Overview on the Anticancer Activity of *Azadirachta indica* (Neem) in Gynecological Cancers. International Journal of Molecular Sciences [Internet]. 2018 Dec 1 [cited 2022 Jan 15]; 19(12). Available from: <https://pubmed.ncbi.nlm.nih.gov/30416351/>
 88. Moga MA, Bălan A, Anastasiu CV, Dimienescu OG, Neculoiu CD and Gavriș C: An Overview on the Anticancer Activity of *Azadirachta indica* (Neem) in Gynecological Cancers. International Journal of Molecular Sciences 2018; 19: 3898 [Internet]. 2018 Dec 5 [cited 2022 Jan 15]; 19(12):3898. Available from: <https://www.mdpi.com/1422-0067/19/12/3898/htm>
 89. Hussein A, Granica S, Gangaram S, Naidoo Y, Dewir YH, El-Hendawy S. Phytochemicals and Biological Activities of Barleria (Acanthaceae). Plants 2022, Vol 11, Page 82 [Internet]. 2021 Dec 28 [cited 2022 Jan 15]; 11(1):82. Available from: <https://www.mdpi.com/2223-7747/11/1/82/htm>
 90. Pai KSR, Srilatha P, Suryakant K, Setty MM, Nayak PG and Rao CM: Anticancer activity of *Berberis aristata* in Ehrlich ascites carcinoma-bearing mice: A preliminary study. <http://dx.doi.org/10.3109/138802092011599035> [Internet]. 2012 Mar [cited 2022 Jan 15]; 50(3): 270–7. Available from: <https://www.tandfonline.com/doi/abs/10.3109/13880209.2011.599035>
 91. Naik Bukke A, Nazneen Hadi F, Babu KS and Shankar PC: *In-vitro* studies data on anticancer activity of *Caesalpinia sappan* L. heartwood and leaf extracts on MCF7 and A549 cell lines. Data in Brief [Internet]. 2018 Aug 1 [cited 2022 Jan 15]; 19: 868. Available from: [https://www.sciencedirect.com/journal/data-in-brief/article/pii/S2352-3440\(18\)30299-1](https://www.sciencedirect.com/journal/data-in-brief/article/pii/S2352-3440(18)30299-1)
 92. Li Y, Dong M, Wu Z, Huang Y, Qian H and Huang C: Activity Screening of the Herb *Caesalpinia sappan* and an Analysis of Its Antitumor Effects. Evidence-based Complementary and Alternative Medicine 2021; 2021.
 93. Alehaideb Z, AlGhamdi S, Yahya W Bin, Al-Eidi H, Alharbi M and Alaujan M: Antiproliferative and Pro-Apoptotic Effects of *Calligonum comosum* (L'Her.) Methanolic Extract in Human Triple-Negative MDA-MB-231 Breast Cancer Cells. Journal of evidence-based integrative medicine [Internet]. 2020 [cited 2022 Jan 15]; 25. Available from: <https://pubmed.ncbi.nlm.nih.gov/33302699/>
 94. Abdo W, Hirata A, Shukry M, Kamal T, Abdel-Sattar E and Mahrous E: *Calligonum comosum* extract inhibits diethylnitrosamine-induced hepatocarcinogenesis in rats. Oncology Letters [Internet]. 2015 Aug 1 [cited 2022 Jan 15]; 10(2):716–22. Available from: <http://www.spandidos-publications.com/10.3892/ol.2015.3313/abstract>
 95. Shalabi M, Khilo K, Zakaria MM, Elsebaei MG, Abdo W and Awadin W: Anticancer activity of *Aloe vera* and *Calligonum comosum* extracts separately on hepatocellular carcinoma cells. Asian Pacific Journal of Tropical Biomedicine 2015; 5(5): 375–81.
 96. Ke Y, Podio M, Conner J and Ozias-Akins P: Single-cell transcriptome profiling of buffelgrass (*Cenchrus ciliaris*) eggs unveils apomictic parthenogenesis signatures. Scientific Reports 2021 11:1 [Internet]. 2021 May 10 [cited 2022 Jan 15]; 11(1):1–17. Available from: <https://www.nature.com/articles/s41598-021-89170-y>
 97. Alothman EA, Awaad AS, Al-Qurayn NA, Al-Kanhal HF, El-Meligy RM and Zain YM: Anticancer effect of *Cenchrus ciliaris* L. Saudi Pharmaceutical Journal: SPJ: the official publication of the Saudi Pharmaceutical Society [Internet]. 2018 Nov 1 [cited 2022 Jan 15]; 26(7):952–5. Available from: <https://pubmed.ncbi.nlm.nih.gov/30416351/>
 98. Mazumder K, Biswas B, Raja IM and Fukase K: A review of cytotoxic plants of the Indian subcontinent and a broad-spectrum analysis of their bioactive compounds. Molecules 2020, Vol 25, Page 1904 [Internet]. 2020 Apr 20 [cited 2022 Jan 15]; 25(8):1904. Available from: <https://www.mdpi.com/1420-3049/25/8/1904/htm>

99. Ohyama K, Akaike T, Hirobe C and Yamakawa T: Cytotoxicity and apoptotic inducibility of *Vitex agnus-castus* fruit extract in cultured human normal and cancer cells and effect on growth. *Biological & pharmaceutical bulletin* [Internet]. 2003 Jan [cited 2022 Jan 18]; 26(1):10–8. Available from: <https://pubmed.ncbi.nlm.nih.gov/12520164/>
100. Ohyama K, Akaike T, Imai M, Toyoda H, Hirobe C and Bessho T: Human gastric signet ring carcinoma (KATO-III) cell apoptosis induced by *Vitex agnus-castus* fruit extract through intracellular oxidative stress. *International J of Biochemis and Cell Biology* 2005; 37(7): 1496–510.
101. Bardaweel SK, Hudaib MM, Tawaha KA and Bashatwah RM: Studies on the *in-vitro* antiproliferative, antimicrobial, antioxidant and acetylcholinesterase inhibition activities associated with chrysanthemum coronarium essential oil. *Evidence-based Complementary and Alternative Medicine* 2015; 2015.
102. Cheng W, Li J, You T and Hu C: Anti-inflammatory and immunomodulatory activities of the extracts from the inflorescence of *Chrysanthemum indicum* Linné. *Journal of Ethnopharmacology* 2005; 101(1–3): 334–7.
103. Thakkar KN, Nayak J, Iyer S V, Kumar S and Prasad AK: Antioxidant and *in-vitro* cytotoxic activity of extracts of aerial parts of *Cocculus hirsutus* (L) using cell line cultures (breast cell line). *The Journal of Phytopharmacology* [Internet]. 2014 [cited 2022 Jan 18]; 3(6): 395–9. Available from: www.phytopharmajournal.com
104. Rahman MA, Sahabjada and Akhtar J: Evaluation of anticancer activity of *Cordia dichotoma* leaves against a human prostate carcinoma cell line, PC3. *Journal of Traditional and Complementary Medicine* [Internet]. 2017 Jul 1 [cited 2022 Jan 18]; 7(3): 315. Available from: <https://pubmed.ncbi.nlm.nih.gov/3066664/>
105. Yurdakök B and Baydan E: Cytotoxic effects of *Eryngium kotschyi* and *Eryngium maritimum* on Hep2, HepG2, Vero and U138 MG cell lines. *Pharmaceutical Biology* 2013; 51(12): 1579–85.
106. Rosangkima G and Jagetia: *In-vitro* anticancer screening of medicinal plants of mizoram state, india, against dalton's lymphoma, mcf-7 and hela cells. [cited 2022 Jan 18]; Available from: <http://www.recentscientific.com>
107. Koparal AT and Bostancıoğlu RB: Promotion of Hair Growth by Traditionally Used Delphinium Staphisagria Seeds through Induction of Angiogenesis. *Iranian Journal of Pharmaceutical Research: IJPR* [Internet]. 2016 Mar 1 [cited 2022 Jan 18]; 15(2):551. Available from: <https://pubmed.ncbi.nlm.nih.gov/28283/>
108. Yin T, Cai L and Ding Z: An overview of the chemical constituents from the genus *Delphinium* reported in the last four decades. *RSC Advances* [Internet]. 2020 Apr 3 [cited 2022 Jan 18]; 10(23):13669–86. Available from: <https://pubs.rsc.org/en/content/articlehtml/2020/ra/d0ra00813c>
109. De D, Chowdhury P, Panda SK and Ghosh U: Ethanolic extract of leaf of *Dillenia pentagyna* reduces *in-vitro* cell migration and induces intrinsic pathway of apoptosis via downregulation of NF- κ B in human NSCLC A549 cells. *Journal of cellular biochemistry* [Internet]. 2019 Dec 1 [cited 2022 Jan 18]; 120(12):19841–57. Available from: <https://pubmed.ncbi.nlm.nih.gov/31318086/>
110. Alves Da Paz DP, Nagamine MK, Grande MP Del, Leite JVP, Sobreira FMG and Bacchi EM: Inhibitory Effects of *Euphorbia tirucalli* Lineu (Euphorbiaceae) Diluted Latex on Human and Canine Melanoma Cells. *Evidence-based Complementary and Alternative Medicine* 2020; 2020.
111. Munro B, Vuong QV, Chalmers AC, Goldsmith CD, Bowyer MC and Scarlett CJ: Phytochemical, antioxidant and anticancer properties of *euphorbia tirucalli* methanolic and aqueous extracts. *Antioxidants* (Basel, Switzerland) [Internet]. 2015 Dec 1 [cited 2022 Jan 18]; 4(4):647–61. Available from: <https://pubmed.ncbi.nlm.nih.gov/26783950/>
112. Choudhari AS, Suryavanshi SA and Kaul-Ghanekar R: The aqueous extract of *Ficus religiosa* induces cell cycle arrest in human cervical cancer cell lines SiHa (HPV-16 Positive) and apoptosis in HeLa (HPV-18 positive). *PLoS one* [Internet]. 2013 Jul 26 [cited 2022 Jan 18]; 8(7). Available from: <https://pubmed.ncbi.nlm.nih.gov/23922932/>
113. (PDF) anticancer effect of fig fruit ficus racemosa extract against human hepatocellular carcinoma (HepG-2) CELL LINE [Internet]. [cited 2022 Jan 18]. Available from: https://www.researchgate.net/publication/332301636_Anti_cancer_Effect_Of_Fig_Fruit_Ficus_Racemosa_Extract_Against_Human_Heptocellular_Carcinoma_Hepg-2_Cell_Line
114. Rattanamaneeerumee A, Thirapanmethee K, Nakamura Y, Bongcheewin B and Chomnawang MT: Chemopreventive and biological activities of *Helicteres isora* L. fruit extracts. *Research in Pharmaceutical Sciences* [Internet]. 2018 [cited 2022 Jan 18]; 13(6):484. Available from: <https://pubmed.ncbi.nlm.nih.gov/3066664/>
115. Raffoul JJ, Kucuk O, Sarkar FH and Hillman GG: Dietary agents in cancer chemoprevention and treatment. *Journal of Oncology* 2012;
116. Nguyen C, Baskaran K, Pupulin A, Ruvinov I, Zaitoon O and Grewal S: Hibiscus flower extract selectively induces apoptosis in breast cancer cells and positively interacts with common chemotherapeutics. *BMC Complementary and Alternative Medicine* [Internet]. 2019 May 6 [cited 2022 Jan 18]; 19(1). Available from: <https://pubmed.ncbi.nlm.nih.gov/33386/>
117. P A, AV, WM and PA de W: Hypericin in cancer treatment: more light on the way. *The international journal of biochemistry & cell biology* [Internet]. 2002 [cited 2022 Jan 18]; 34(3). Available from: <https://pubmed.ncbi.nlm.nih.gov/11849990/>
118. Mirmalek SA, Azizi MA, Jangholi E, Yadollah-Damavandi S, Javidi MA and Parsa Y: Cytotoxic and apoptogenic effect of hypericin, the bioactive component of *Hypericum perforatum* on the MCF-7 human breast cancer cell line. *Cancer Cell International* [Internet]. 2016 Feb 9 [cited 2022 Jan 18]; 16(1):1–9. Available from: <https://pubmed.ncbi.nlm.nih.gov/28283/>
119. Bar-Shalom R, Bergman M, Grossman S, Azzam N, Sharvit L and Fares F: *Inula viscosa* Extract Inhibits Growth of Colorectal Cancer Cells *in-vitro* and *in-vivo* Through Induction of Apoptosis. *Frontiers in oncology* [Internet]. 2019 [cited 2022 Jan 18]; 9(APR). Available from: <https://pubmed.ncbi.nlm.nih.gov/31024836/>
120. Virdis P, Migheli R, Galleri G, Fancello S, Cadoni MPL and Pintore G: Antiproliferative and proapoptotic effects of *Inula viscosa* extract on Burkitt lymphoma cell line. *Tumor Biology* 2020; 42(2).
121. Merghoub N, El Btaouri H, Benbacer L, Gmouh S, Trentesaux C and Brassart B: *Inula viscosa* extracts induces telomere shortening and apoptosis in cancer cells and overcome drug resistance. *Nutrition and Cancer* 2016; 68(1): 131–43.
122. Kalaiselvi M, Narmadha R, Ragavendran P, Vidya B, Gomathi D and Raj CA: Chemopreventive effect and

- HPTLC fingerprinting analysis of *Jasminum sambac* (L.) Ait. Extract against DLA-induced lymphoma in experimental animals. Applied biochemistry and biotechnology [Internet]. 2013 Feb [cited 2022 Jan 18]; 169(4): 1098–108. Available from: <https://pubmed.ncbi.nlm.nih.gov/23306882/>
123. Wu LC, Lin CL, Peng CC, Huang TL, Tsai TH and Kuan YE: Development from *Jasminum sambac* Flower Extracts of Products with Floral Fragrance and Multiple Physiological Activities. Evidence-based Complementary and Alternative Medicine 2021; 2021.
 124. Zhao Y, Chen R, Wang Y, Qing C, Wang W and Yang Y: *In-vitro* and *In-vivo* Efficacy Studies of Lavender *angustifolia* Essential Oil and Its Active Constituents on the Proliferation of Human Prostate Cancer. Integrative cancer therapies [Internet]. 2017 Jun 1 [cited 2022 Jan 18]; 16(2):215–26. Available from: <https://pubmed.ncbi.nlm.nih.gov/27151584/>
 125. Ghagane SC, Puranik SI, Kumbar VM, Nerli RB, Jalalpure SS and Hiremath MB: *In-vitro* antioxidant and anticancer activity of *Lea indica* leaf extracts on human prostate cancer cell lines. Integrative Medicine Research [Internet]. 2017 Mar [cited 2022 Jan 18]; 6(1):79. Available from: [/pmc/articles/PMC5395687/](https://pubmed.ncbi.nlm.nih.gov/30995545/)
 126. Medini F, Bourgou S, Lalancette KG, Snoussi M, Mkadmini K and Coté I: Phytochemical analysis, antioxidant, anti-inflammatory, and anticancer activities of the halophyte *Limonium densiflorum* extracts on human cell lines and murine macrophages. South African Journal of Botany 2015; 99: 158–64.
 127. Abdel-Salam IM, Abou-Bakr AA and Ashour M: Cytotoxic effect of aqueous ethanolic extract of *Luffa cylindrica* leaves on cancer stem cells CD44 +/24 - in breast cancer patients with various molecular sub-types using tissue samples *in-vitro*. Journal of ethnopharmacology [Internet]. 2019 Jun 28 [cited 2022 Jan 18]; 238. Available from: <https://pubmed.ncbi.nlm.nih.gov/30995545/>
 128. Abdel-Salam IM, Awadein NES and Ashour M: Cytotoxicity of *Luffa cylindrica* (L.) M. Roem. extract against circulating cancer stem cells in hepatocellular carcinoma. J of Ethnopharmacology 2019; 229: 89–96.
 129. Abdel-Salam IM, Ashmawy AM, Hilal AM, Eldahshan OA and Ashour M: Chemical Composition of Aqueous Ethanol Extract of *Luffa cylindrica* Leaves and its Effect on Representation of Caspase-8, Caspase-3 and the Proliferation Marker Ki67 in Intrinsic Molecular Subtypes of Breast Cancer *in-vitro*. Chemistry and Biodiversity 2018; 15(8).
 130. Yehia S, Abdel-Salam IM, Elgamel BM, El-Agamy B, Hamdy GM and Aldesouki HM: Cytotoxic and Apoptotic Effects of *Luffa Cylindrica* Leaves Extract against Acute Lymphoblastic Leukemic Stem Cells. Asian Pacific Journal of Cancer Prevention 2020; 21(12): 3661–8.
 131. Tan BL, Norhaizan ME and Chan LC: *Manilkara zapota* (L.) P. Royen leaf water extract induces apoptosis in human hepatocellular carcinoma (HepG2) Cells via ERK1/2/Akt1/JNK1 Signaling Pathways. Evidence-based Complementary and Alternative Medicine 2018; 2018.
 132. Kusumawati R, Nasrullah AH, Pesik RN, Muthmainah and Indarto D: Secondary metabolites of *Mirabilis jalapa* structurally inhibit Lactate Dehydrogenase A in silico: a potential cancer treatment. IOP Conference Series: Materials Science and Engineering [Internet]. 2018 Mar 1 [cited 2022 Jan 18]; 333(1):012078. Available from: <https://iopscience.iop.org/article/10.1088/1757-899X/333/1/012078>
 133. Liu J, Li Y, Ren W and Hu WX: Apoptosis of HL-60 cells induced by extracts from *Narcissus tazetta* var. *chinensis*. Cancer letters [Internet]. 2006 Oct 8 [cited 2022 Jan 18]; 242(1): 133–40. Available from: <https://pubmed.ncbi.nlm.nih.gov/16427186/>
 134. Abbas JA, El-Oqlah AA and Mahasneh AM: Herbal plants in the traditional medicine of Bahrain. Economic Botany. 1992; 46(2): 158–63.
 135. Abu-Darwish MS, Efferth T. Medicinal plants from near east for cancer therapy. Frontiers in Pharmacology 2018; 9: 56.
 136. Luke AM, Patnaik R, Kuriadom ST, Jaber M and Mathew S: An *in-vitro* study of *Ocimum sanctum* as a chemotherapeutic agent on oral cancer cell-line. Saudi Journal of Biological Sciences 2021; 28(1): 887–90.
 137. Akhtar MS and Swamy MK: Anticancer plants: Properties and application. Anticancer plants: Properties and Application 2018; 1: 1–582.
 138. Kaushal N, Rao S, Ghanghas P, Abraham S, George T and D'souza S: Usefulness of *Ocimum sanctum* Linn. in Cancer Prevention: An Update. Anticancer plants: Properties and Application [Internet]. 2018 Jun 29 [cited 2022 Jan 18]; 1: 415–29. Available from: https://link.springer.com/chapter/10.1007/978-981-10-8548-2_18
 139. Mabberley DJ: Mabberley's Plant-book: a portable dictionary of plants, their classification and uses / David J. Mabberley. - 3rd ed., completely rev. 3rd ed. 2008 [cited 2022 Jan 18]; 1021. Available from: https://www.researchgate.net/publication/331223183_Botanical_features_phytochemical_and_pharmacological_over_views_of_Oldenlandia_corymbosa_Linn_A_brief_review
 140. Albogami S and Hassan AM: Assessment of the Efficacy of Olive Leaf (*Olea europaea* L.) Extracts in the Treatment of Colorectal Cancer and Prostate Cancer Using *In Vitro* Cell Models. Molecules (Basel, Switzerland) [Internet]. 2021 Jul 1 [cited 2022 Jan 18]; 26(13). Available from: <https://pubmed.ncbi.nlm.nih.gov/34279409/>
 141. Fares R, Bazzi S, Baydoun SE, Abdel-Massih RM. The Antioxidant and Antiproliferative Activity of the Lebanese *Olea europaea* extract. Plant Foods for Human Nutrition 2011; 66(1): 58–63.
 142. Talib WH and Mahasneh AM: Antiproliferative activity of plant extracts used against cancer in traditional medicine. Scientia pharmaceutica [Internet]. 2010 [cited 2022 Jan 18]; 78(1):33–45. Available from: <https://pubmed.ncbi.nlm.nih.gov/21179373/>
 143. Chishti S, Kaloo ZA and Sultan P: Journal of Pharmacognosy and Phytotherapy Medicinal importance of genus *Origanum*: A review. 2013 [cited 2022 Jan 18]; 5(10):170–7. Available from: <http://www.academicjournals.org/JPP>
 144. Kumar S, Chashoo G, Saxena AK and Pandey AK: *Parthenium hysterophorus*: A Probable Source of Anticancer, Antioxidant and Anti-HIV Agents. BioMed Research International [Internet]. 2013 [cited 2022 Jan 18]; 2013. Available from: [/pmc/articles/PMC3848086/](https://pubmed.ncbi.nlm.nih.gov/24279409/)
 145. Góngora L, Máez S, Giner RM, Carmen Recio M, Gray AI and Ríos JL: Phenolic glycosides from *Phagnalon rupestre*. Phytochemistry 2002; 59(8): 857–60.
 146. Soni D and Grover A: Picrosides from *Picrorhiza kurroa* as potential anti-carcinogenic agents. Biomedicine & Pharmacotherapy 2019; 109: 1680–7.
 147. Kumar Sharma A, Kumar S, Chashoo G, Saxena AK and Pandey AK: Cell cycle inhibitory activity of *Piper longum*

- against A549 cell line and its protective effect against metal-induced toxicity in rats. *Indian Journal of Biochemistry & Biophysics* 2014; 51: 358–64.
148. Gezici S, Sekeroglu N and Kijjoa A: *In-vitro* anticancer activity and antioxidant properties of essential oils from *Populus alba* L. and *Rosmarinus officinalis* L. from South Eastern Anatolia of Turkey. *Indian Journal of Pharmaceutical Education and Research* [Internet]. 2017 Jul 1 [cited 2022 Jan 18]; 51(3):S498–503. Available from: https://www.researchgate.net/publication/320248735_In_vitro_Anticancer_Activity_and_Antioxidant_Properties_of_Essential_Oils_from_Populus_alba_L_and_Rosmarinus_officinalis_L_from_South_Eastern_Anatolia_of_Turkey
 149. Ma J, Gao Y, Jiang T and Tian F: Excellent Anti-lung Cancer Activity of *Populus nigra* and Phylogenetic Analysis. *Journal of oleo science* [Internet]. 2021 [cited 2022 Jan 18]; 70(12):1783–9. Available from: <https://pubmed.ncbi.nlm.nih.gov/34866109/>
 150. Talib WH and Mahasneh AM: Antiproliferative Activity of Plant Extracts Used Against Cancer in Traditional Medicine. *Scientia Pharmaceutica* [Internet]. 2010 [cited 2022 Jan 18]; 78(1): 33. Available from: <https://pubmed.ncbi.nlm.nih.gov/21894557/>
 151. (PDF) Cytotoxic flavonoids from *Diplotaxis harra* (Forssk.) Boiss. growing in Sinai [Internet]. [cited 2022 Jan 18]. Available from: https://www.researchgate.net/publication/230635715_Cytotoxic_flavonoids_from_Diplotaxis_harra_Forssk_Boiss_growing_in_Sinai
 152. Zare H: Effects of *Salvia officinalis* Extract on the Breast Cancer Cell Line. *Sci Medicine Journal* [Internet]. 2019 Mar 1 [cited 2022 Jan 18]; 1(1): 25–9. Available from: <https://scimedjournal.org/index.php/SMJ/article/view/7>
 153. Sertel S, Eichhorn T, Plinkert PK and Effertth T: [Anticancer activity of *Salvia officinalis* essential oil against HNSCC cell line (UMSCC1)]. *HNO* [Internet]. 2011 Dec [cited 2022 Jan 18]; 59(12): 1203–8. Available from: <https://pubmed.ncbi.nlm.nih.gov/21894557/>
 154. Jeong HJ, Koo BS, Kang TH, Shin HM, Jung S, Jeon S. Inhibitory effects of *Saururus chinensis* and its components on stomach cancer cells. *Phytomedicine: international journal of phytotherapy and phytopharmacology* [Internet]. 2015 Feb 15 [cited 2022 Jan 19]; 22(2): 256–61. Available from: <https://pubmed.ncbi.nlm.nih.gov/25765830/>
 155. Gao X, He J, Wu X De, Peng LY, Shao LD and Li Y: Sauruchinenols A and B, unprecedented monocyclic diterpenes with new carbon skeleton from the aerial parts of *Saururus chinensis*. *Fitoterapia* 2017; 116: 116–20.
 156. Küpeli Akkol E, Bahadır Acikara Ö, Süntar I, Ergene B and Saltan Çitoğlu G: Ethnopharmacological evaluation of some *Scorzonera* species: *In-vivo* anti-inflammatory and antinociceptive effects. *Journal of Ethnopharmacology*. 2012; 140(2): 261–70.
 157. Lenzion K, Gornowicz A, Bielawski K and Bielawska A: Phytochemical Composition and Biological Activities of *Scorzonera* Species. *International Journal of Molecular Sciences* 2021, Vol 22, Page 5128 [Internet]. 2021 May 12 [cited 2022 Jan 19];22(10):5128. Available from: <https://www.mdpi.com/1422-0067/22/10/5128/htm>
 158. Dall'Acqua S, Ak G, Sut S, Ferrarese I, Zengin G and Yıldızıtugay E: Phenolics from *Scorzonera tomentosa* L.: Exploring the potential use in industrial applications via an integrated approach. *Industrial Crops and Products* 2020; 154: 112751.
 159. Dou C, Zhang B, Han M, Jin X, Sun L and Li T: Antitumor activity of polysaccharides extracted from *senecio scandens* Buch, -Ham root on hepatocellular carcinoma. *Tropical Journal of Pharmaceutical Research* [Internet]. 2017 Jan 1 [cited 2022 Jan 19]; 16(1):43–9. Available from: https://www.researchgate.net/publication/313492449_Antitumor_activity_of_polysaccharides_extracted_from_Senecio_scandens_Buch_Ham_root_on_hepatocellular_carcinoma
 160. Arroyo-Acevedo JL, Herrera-Calderon O, Rojas-Armas JP, Chávez-Asmat R, Calva J and Behl T: Histopathological evaluation of *Senecio rhizomatus* Rusby in 7,12-dimethylbenz(α) anthracene-induced breast cancer in female rats. *Veterinary World* [Internet]. 2021 Mar 1 [cited 2022 Jan 19]; 14(3): 569. Available from: <https://pubmed.ncbi.nlm.nih.gov/34866109/>
 161. Uddin A and Chaturvedi HC: Abscisic Acid from Berries of *Solanum khasianum* Cl. *Zeitschrift für Pflanzenphysiologie* 1981; 102(5): 471–2.
 162. Ayyad SEN, Abdel-Lateff A, Alarif WM, Patacchioli FR, Badria FA and Ezmirly ST: *In-vitro* and *in-vivo* study of cucurbitacins-type triterpene glucoside from *Citrullus colocynthis* growing in Saudi Arabia against hepatocellular carcinoma. *Environmental toxicology and pharmacology* [Internet]. 2012 Mar [cited 2022 Jan 19]; 33(2): 245–51. Available from: <https://pubmed.ncbi.nlm.nih.gov/22245841/>
 163. Liu M, Yan Q, Peng B, Cai Y, Zeng S and Xu Z: Use of cucurbitacins for lung cancer research and therapy. *Cancer Chemotherapy and Pharmacology* 2021; 88(1).
 164. Mazumder K, Biswas B, Raja IM and Fukake K: A Review of Cytotoxic Plants of the Indian Subcontinent and a Broad-Spectrum Analysis of Their Bioactive Compounds. *Molecules* 2020, Vol 25, Page 1904 [Internet]. 2020 Apr 20 [cited 2022 Jan 19]; 25(8): 1904. Available from: <https://www.mdpi.com/1420-3049/25/8/1904/htm>
 165. Wozniak M, Michalak B, Wyszomierska J, Dudek MK and Kiss AK: Effects of phytochemically characterized extracts from *Syringa vulgaris* and isolated secoiridoids on mediators of inflammation in a human neutrophil model. *Frontiers in Pharmacology* [Internet]. 2018 Apr 11 [cited 2022 Jan 19]; 9(APR):349. Available from: <https://pubmed.ncbi.nlm.nih.gov/313492449/>
 166. Yadav SS, Meshram GA, Shinde D, Patil RC, Manohar SM and Upadhye MV: Antibacterial and Anticancer Activity of Bioactive Fraction of *Syzygium cumini* L. Seeds. *HAYATI Journal of Biosciences* 2011; 18(3): 118–22.
 167. Banerjee A, Dasgupta N and De B: *In-vitro* study of antioxidant activity of *Syzygium cumini* fruit. *Food Chemistry* 2005; 90(4): 727–33.
 168. Barh D: *Syzygium cumini* Inhibits Growth and Induces Apoptosis in Cervical Cancer Cell Lines: A Primary Study. *Ecancer Medical Science* 2008.
 169. Kumar A and Selvakumar S: Antiproliferative efficacy of *Tabernaemontana divaricata* against HEP2 cell line and Vero cell line. *Pharmacognosy Magazine* [Internet]. 2015 [cited 2022 Jan 19]; 11(Suppl 1): S46. Available from: <https://pubmed.ncbi.nlm.nih.gov/25765830/>
 170. Shini SS and Devi MP: Studies on the *in-vitro* anticancer activity of *Tabernaemontana divaricata* extract against colon Cancer cell line. *International Journal of Pharma and Bio Sciences* 2017; 8(2).
 171. Anburaj G, Marimuthu M, Rajasudha V and Manikandan R: *In-vitro* anticancer activity *Tecoma stans* against human

- breast cancer yellow elder (*Tecoma stans*). ~ 331 ~ Journal of Pharmacognosy and Phytochemistry 2016; 5(4).
172. Sa Marzouk M, Gamal-Eldeen AM, Mohamed MA and El-Sayed MM: Antioxidant and Antiproliferative Active Constituents of *Tecoma stans* against Tumor Cell Lines.
 173. Rahmouni F, Saoudi M and Rebai T: Therapeutics studies and biological properties of *Teucrium polium* (Lamiaceae). *Bio Factors* 2021; 47(6): 952–63.
 174. Tabatabaie PS and Yazdanparast R: *Teucrium polium* extract reverses symptoms of streptozotocin-induced diabetes in rats via rebalancing the Pdx1 and FoxO1 expressions. *Biomedicine and Pharmacology* 2017; 93: 1033–9.
 175. Movahedi A, Basir R, Rahmat A, Charaffedine M and Othman F: Remarkable Anticancer Activity of *Teucrium polium* on Hepatocellular Carcinogenic Rats. Evidence-based complementary and alternative medicine: eCAM [Internet]. 2014 [cited 2022 Jan 19]; 2014. Available from: <https://pubmed.ncbi.nlm.nih.gov/25197311/>
 176. Lowe HIC, Toyang NJ, Watson CT, Ayeah KN and Bryant J: HLB-100: A highly potent anticancer flavanone from *Tillandsia recurvata* (L.). *Cancer Cell International* 2017; 17(1).
 177. Cabrera GM and Seldes AM: Hydroperoxycycloartanes from *Tillandsia recurvata*. *Journal of Natural Products* [Internet]. 2004 [cited 2022 Jan 19]; 58(12): 1920–4. Available from: <https://pubs.acs.org/doi/abs/10.1021/np50126a020>
 178. Tariq A, Sadia S, Pan K, Ullah I, Mussarat S and Sun F: A systematic review on ethnomedicines of anticancer plants. *Phytotherapy Research* [Internet]. 2017 Feb 1 [cited 2022 Jan 19]; 31(2):202–64. Available from: <https://onlinelibrary.wiley.com/doi/full/10.1002/ptr.5751>
 179. Riaz M, Zia-Ul-Haq M and Jaafar HZE: Common mullein, pharmacological and chemical aspects. *Revista Brasileira de Farmacognosia* 2013; 23(6): 948–59.
 180. Grace Nirmala J, Evangeline Celsia S, Swaminathan A, Narendhirakannan RT and Chatterjee S: Cytotoxicity and apoptotic cell death induced by *Vitis vinifera* peel and seed extracts in A431 skin cancer cells. *Cytotechnology* [Internet]. 2018 Apr 1 [cited 2022 Jan 19]; 70(2):537–54. Available from: <https://pubmed.ncbi.nlm.nih.gov/28983752/>
 181. Nassiri-Asl M and Hosseinzadeh H: Review of the pharmacological effects of *Vitis vinifera* (grape) and its bioactive compounds. *Phytotherapy Research* 2009; 23(9): 1197–204.
 182. Al-Oqail MM, Al-Sheddi ES, Farshori NN, Al-Massarani SM, Al-Turki EA and Ahmad J: Corn Silk (*Zea mays* L.) Induced Apoptosis in Human Breast Cancer (MCF-7) Cells via the ROS-Mediated Mitochondrial Pathway. *Oxidative medicine and cellular longevity* [Internet]. 2019 [cited 2022 Jan 19]; 2019. Available from: <https://pubmed.ncbi.nlm.nih.gov/31781357/>
 183. Balasubramanian K and Padma PR: Anticancer activity of *Zea mays* leaf extracts on oxidative stress-induced Hep2 cells. *JAMS Journal of Acupuncture and Meridian Studies* [Internet]. 2013 [cited 2022 Jan 19]; 6(3): 149–58. Available from: https://www.researchgate.net/publication/236604881_Anticancer_Activity_of_Zea_mays_Leaf_Extracts_on_Oxidative_Stress-induced_Hep2_Cells
 184. Lau BHS, Tadi PP and Tosk JM: *Allium sativum* (Garlic) and cancer prevention. *Nutrition Research*. 1990; 10(8): 937–48.
 185. Özkan İ, Koçak P, Yıldırım M, Ünsal N, Yılmaz H and Telci D: Garlic (*Allium sativum*)-derived SEVs inhibit cancer cell proliferation and induce caspase mediated apoptosis. *Scientific Reports* 2021; 11(1).
 186. Lee JH, Yang HS, Park KW, Kim JY, Lee MK and Jeong IY: Mechanisms of thiosulfates from *Allium tuberosum* L.-induced apoptosis in HT-29 human colon cancer cells. *Toxicology Letters* 2009; 188(2): 142–7.
 187. Epifano F, Fiorito S and Genovese S: Phytochemistry and pharmacognosy of the genus *Acronychia*. *Phytochemistry* 2013; 95: 12–8.
 188. Obiagwu MO, Ihekwereme CP, Ajaghaku DL, Okoye FBC. The Useful Medicinal Properties of the Root-Bark Extract of *Alstonia boonei* (Apocynaceae) May Be Connected to Antioxidant Activity. *ISRN Pharmacology* 2014; 2014: 1–4.
 189. Adotey JPK, Adukpo GE, Opoku Boahen Y and Armah FA: A Review of the Ethnobotany and Pharmacological Importance of *Alstonia boonei* De Wild (Apocynaceae). *ISRN Pharmacology* 2012; 2012: 1–9.
 190. Kubatka P, Uramova S, Kello M, Kajo K, Samec M and Jasek K: Anticancer Activities of *Thymus vulgaris* L. in experimental breast carcinoma *in-vivo* and *in-vitro*. *International Journal of Molecular Sciences* [Internet]. 2019 Apr 1 [cited 2022 Jan 19]; 20(7). Available from: [/pmc/articles/PMC6479806/](https://pubmed.ncbi.nlm.nih.gov/31781357/)
 191. Li Y, Li S, Meng X, Gan RY, Zhang JJ and Li H Bin: Dietary natural products for prevention and treatment of breast cancer. *Nutrients* 2017; 9(7).
 192. Gu JQ, Gills JJ, Park EJ, Mata-Greenwood E, Hawthorne ME and Axelrod F: Sesquiterpenoids from *Tithonia diversifolia* with potential cancer chemopreventive activity. *Journal of natural products* [Internet]. 2002 [cited 2022 Jan 19]; 65(4): 532–6. Available from: <https://pubmed.ncbi.nlm.nih.gov/11975495/>

How to cite this article:

Kumar V, Garg V and Dureja H: Plant derived anticancer agents for treatment of cancer. *Int J Pharm Sci & Res* 2022; 13(9): 3375-96. doi: 10.13040/IJPSR.0975-8232.13(9).3375-96.

All © 2022 are reserved by International Journal of Pharmaceutical Sciences and Research. This Journal licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported License.

This article can be downloaded to **Android OS** based mobile. Scan QR Code using Code/Bar Scanner from your mobile. (Scanners are available on Google Playstore)