



Received on 22 February 2022; received in revised form, 24 April 2022; accepted 21 September 2022; published 01 October 2022

PHYTOCHEMICAL AND PHARMACOGNOSTICAL SCREENING OF *JASMINUM MULTIFLORUM* (BURM. F.) AND *JASMINUM MESNYI* (HANCE) STEMS

Preeti Garg¹ and Vandana Garg^{*2}

Department of Pharmacognosy and Phytochemistry¹, Hindu College of Pharmacy, Sonapat - 131001, Haryana, India.

Department of Pharmaceutical Sciences², Maharshi Dayanand University, Rohtak - 124001, Haryana, India.

Keywords:

Jasminum multiflorum, *Jasminum mesnyi* standardization, Pharmacognostical, Phytochemical

Correspondence to Author:

Dr. Vandana Garg

Assistant Professor,
Department of Pharmaceutical
Sciences, Maharshi Dayanand
University, Rohtak - 124001,
Haryana, India.

E-mail: vandugarg@rediffmail.com

ABSTRACT: *Jasminum multiflorum* and *Jasminum mesnyi*, the ornamental shrubs belonging to family Oleaceae are widely distributed throughout India. Despite their use in the essence industry for making perfumes, oils, and creams, these species are also used in the healing wounds and ulcers, constipation, flatulence, skin diseases, rheumatism, stomatitis, diabetes, stress, antiseptic, diuretics, and gastric disturbance. The present study is designed to establish the authenticity of both species by evaluating various standardization parameters. The stems of both species were collected and dried under shade. Freehand transverse sections were taken for microscopic examination. The standardization parameters such as Moisture content, Ash values, Extractive values, Heavy metal content, Aflatoxin content, Microbial infestation and, Pesticides residue have been determined as per WHO guidelines. Results showed the *J. multiflorum* stem was greenish-grey in color, cylindrical shaped, fragile and have aromatic odor. Microscopy showed the presence of glandular multicellular trichomes, epidermis, collenchyma, endodermis, medullary rays, vessels, xylem cells, parenchyma, fibre bundle and calcium oxalate crystals. The stem of *J. mesnyi* is yellowish-green in color the surface is smooth and pubescent. The epidermis, collenchymas, endodermis, medullary rays, xylem cells, vessels, trichomes and parenchyma are also present in the microscopy. Preliminary phytochemical screening of both plants confirmed the presence of alkaloids, anthraquinone glycosides, saponins glycosides, cardiac glycosides, flavonoids, steroids, terpenoids, tannins, and phenolic compounds. The analysis of Heavy metal, Aflatoxin, microbial contamination and pesticide residual values were found within limits. The result of our study confirmed the authenticity of both the species and the parameters studied can be used in the future as a part of the monograph.

INTRODUCTION: Ayurveda is one of the most renowned traditional systems of medicine that has survived and flourished for ages to date. In the modern time, when the allopathic system of medicine has reached almost the top because of

validated research and advanced techniques still herbal medicines are the choice of drug for several chronic diseases such as cancer, diabetes, arthritis, and asthma. The various species of the genus *Jasminum* are used by people for their effectiveness.

The genus *Jasminum* belongs to the Oleaceae family having 200 species throughout the world and out of which around 40 species are native to India¹. *Jasminum multiflorum* is also known as winter jasmine or musk jasmine; which is an ever green winner shrub with young branches dressed

	QUICK RESPONSE CODE DOI: 10.13040/IJPSR.0975-8232.13(10).4035-43
	This article can be accessed online on www.ijpsr.com
DOI link: http://dx.doi.org/10.13040/IJPSR.0975-8232.13(10).4035-43	

with velvety pubescence. *Jasminum mesnyi*, well known as Yellow jasmine, is a perennial shrub^{2,3}. Being known for their fragrances, the flowers are used in aromatherapy for uplifting the mood. Both the species are found throughout India, in the forests of Western Ghats, sub-Himalayan tracts up to 1300-1500 meters altitude and are also cultivated as garden plants⁴. Traditionally these plants are used to treat a variety of ailments like treat anxiety, anorexia, diabetes, depression, gastric disturbance, muscular pain, nocturnal emission, oral sores, stress, ulcer, uropathy, cephalalgia, cardiac disorders, constipation, indigestion, inflammation, rheumatism, lactifuge, weakness of sight, and snake bite⁵⁻⁹.

The various secondary metabolites namely alkaloids, glycosides, flavonoids, steroids, terpenoids, tannins and saponins are substantiated in different parts of the plants^{5, 10, 11}. The pharmacological studies proved their efficacy in *J. multiflorum* towards antioxidant^{12,13}, antimicrobial¹⁴, antihelmintic¹⁵, nematocidal¹⁶, GI¹⁷, in cardiovascular diseases^{18, 19} and effect on central nervous system^{20, 21} and *J. mesnyi* revealed its wound healing potential of roots²², antioxidant^{23, 24}, antiulcer²⁵, antihyperglycemic²⁶, antihelmintic²⁷ and antimicrobial²⁸ efficacy of leaves were substantiated. The main hindrance to the acceptance of herbal medicines is the lack of documentation and quality control. The establishment of pharmacognostic standards for the traditional plants is a prerequisite for its detailed phytochemical and pharmacological investigation. The present study was done for establishing pharmacognostic and physicochemical standards of the plant stem as per WHO Guidelines which will further help in drug identification as well as in detecting/establishing adulteration.

MATERIALS AND METHODS:

Plant Collection and Authentication: *J. multiflorum* and *J. mesnyi* stems were collected from the Botanical Garden of Hindu College of Pharmacy, Sonapat. The plants were authenticated by Dr. Sunita Garg (Emeritus Scientist, CSIR-NISCAIR, and New Delhi, India). The voucher specimen (NISCAIR/RHMD/CONSULT/2018/329-6-97-1 and 3296-97-2 for *J. multiflorum*, *J. mesnyi* respectively) was deposited in the Department of Pharmacognosy, Hindu College of Pharmacy,

Sonepat for future reference. The plant materials were dried under shade by placing in a single layer and coarsely powdered by hand mixer and pass through sieve no #60.

Macroscopy: Untreated powdered samples were observed under daylight and the color of samples was recorded. The powder was rubbed slowly between fingers and odor was examined. Tastes of the powder were also observed. The surface was examined by softly touching it²⁹.

Microscopy: The thin transverse sections of both fresh stems were cut with the help of a sharp blade and cleared with chloral hydrate solution. Freehand sections were stained with phloroglucinol, ruthenium red, safranin and hydrochloric acid and further mounted in glycerine and covered with coverslip. For powder microscopy, the powder of stems was passed through sieve no # 60 and taken on glass slides. The same procedure was followed as mentioned above to prepare the slides. The slides were observed under a compound microscope and photographs were taken using Labomed ATC- 2000 microscope attached with camera²⁹⁻³².

Qualitative Analysis: The procedure recommended by Indian Pharmacopoeia was used for determining the total ash, water-soluble ash, acid insoluble ash, water-soluble extractive, ethanol-soluble extractive, loss on drying, swelling index, crude fibre content and foaming index²⁹⁻³², where as aflatoxin content, microbial contamination, heavy metal analysis and pesticidal residues were evaluated according to WHO guidelines³². A fluorescence study was performed as per standard procedure^{29, 33}.

Preliminary Phytochemical Screening: For the preliminary phytochemical screening, extracts of both species were prepared according to the polarity of the solvents (petroleum ether, chloroform, ethanol and aqueous) as per the standard procedure. The preliminary phytochemical screenings of extracts were performed according to standard procedure^{29, 33}.

RESULTS AND DISCUSSION:

Macroscopy: *J. multiflorum* of outer bark is thin and greenish-grey in color. The stem is buff colored with the central brown part.

The outer surface is rough, longitudinal wrinkles are present with alternate nodes and yellowish spots are present on the bark. Fracture is easy and uneven. The stem is thin and cylindrical in shape as shown in **Fig. 1**. It is bitter in taste and odor is aromatic. The stem of *J. mesnyi* yellowish-green

in color, the surface is smooth and pubescent. The nodes are present in the opposite positions; internodes are uniform about 4 to 5cm as shown in **Fig. 2**. The internal color of the stem is brown. The dried stem is woody and the fracture is smooth and easy to smash.



FIG. 1: (A) *J. MULTIFLORUM* PLANT SHOWING LEAVES, BUD, FLOWER AND STEM; (B) DRIED STEM



FIG. 2: (A) *J. MESNYI* PLANT SHOWING LEAVES FLOWER AND STEM; (B) DRIED STEM

Microscopy: Transverse section of *J. multiflorum* stem showed the presence of non-glandular, multicellular trichomes covering the epidermis. Collenchyma lies beneath the epidermis layer followed by endodermis. Medullary rays, vessels, xylem cells, fibre bundle, and parenchyma were observed under the endodermis layer. Vessels are mostly isolated or present in a group of two to four. Some cells were observed with calcium oxalate

crystals as shown in **Fig. 3, 4**. The transverse section of the *J. mesnyi* stem showed a quadrangular shape showing entire epidermis. Under the epidermis, the collenchyma layer is presently followed by endodermis. Medullary rays, xylem cells, and vessels and parenchyma were observed under the endodermis layer. Vessels are mostly isolated and unicellular trichomes were present as shown in **Fig. 5, 6**.

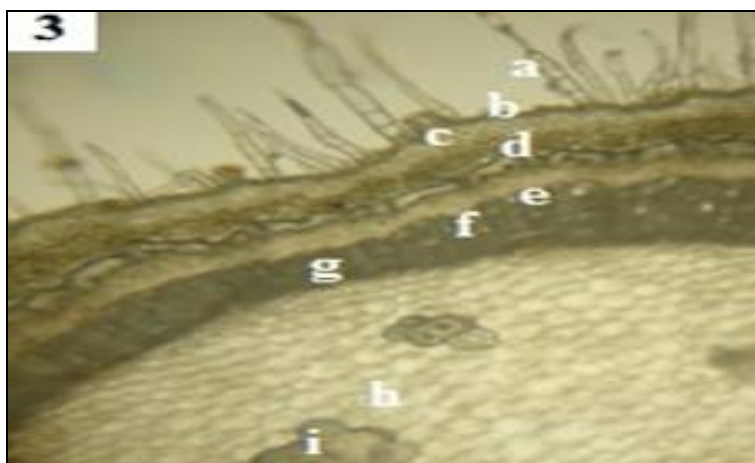


FIG. 3: REPRESENTATIVE PHOTOMICROGRAPH OF TRANSVERSE SECTION STEM *J. MULTIFLORUM* AT 10 X SHOWING (A) AGLANDULAR MULTICELLULAR TRICHOMES; (B) EPIDERMIS; (C) COLLENCHYMAS; (D) ENDODERMIS; (E) VESSELS; (F) XYLEM CELLS; (G) MEDULLARY RAYS; (H) PARENCHYMA AND (I) FIBRE BUNDLE

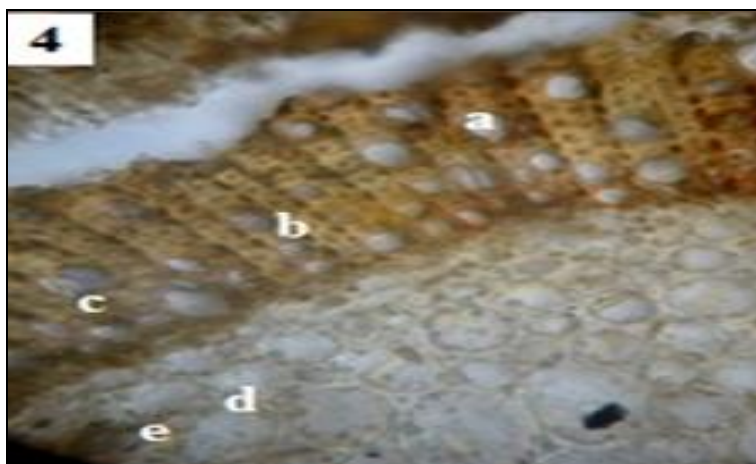


FIG. 4: REPRESENTATIVE PHOTOMICROGRAPH OF TRANSVERSE SECTION OF STEM OF *J. MULTIFLORUM* AT 45 X SHOWING (A) VESSELS; (B) XYLEM CELLS; (C) MEDULLARY RAYS; (D) PARENCHYMA; (E) FIBRE BUNDLE; (F) CALCIUM OXALATE CRYSTAL

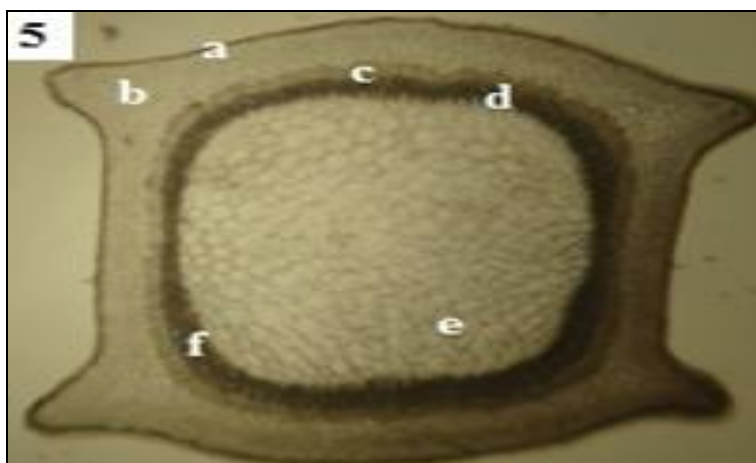


FIG. 5: MICROSCOPY OF TRANSVERSE SECTION OF *J. MESNYI* SHOWING AT 10X (A) EPIDERMIS; (B) COLLENCHYMAS; (C) ENDODERMIS; (D) MEDULLARY RAYS; (E) PARENCHYMA (F) XYLEM CELLS AND VESSELS

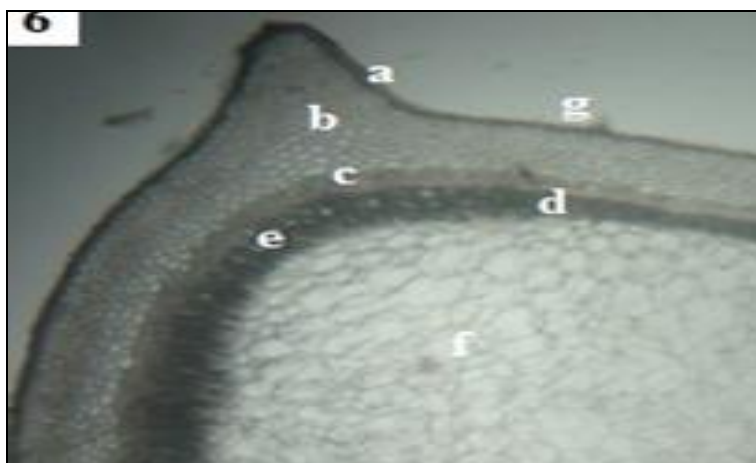


FIG. 6: MICROSCOPY OF TRANSVERSE SECTION OF *J. MESNYI* SHOWING AT 45X (A) EPIDERMIS; (B) COLLENCHYMAS; (C) ENDODERMIS; (D) MEDULLARY RAYS; (E) XYLEM CELLS AND VESSELS; (F) PARENCHYMA AND (G) TRICHOMES

Powder microscopic characters of the stem of *J. multiflorum* showed the presence of trichomes, parenchymatous cells and xylem fibre as shown in

Fig. 7 and *J. mesnyi* showed the presence of parenchymatous cells and xylem fibre as shown in **Fig. 8.**

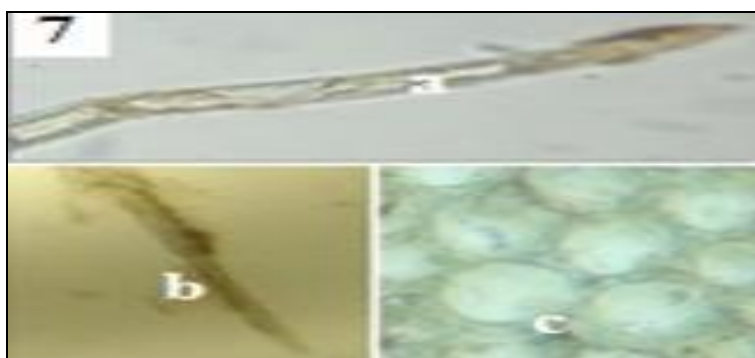


FIG. 7: POWDER MICROSCOPY OF STEM OF *J. MULTIFLORUM* AT 45X (A) TRICHOME; (B) XYLEM FIBRE AND (C) PARENCHYMATOUS CELL

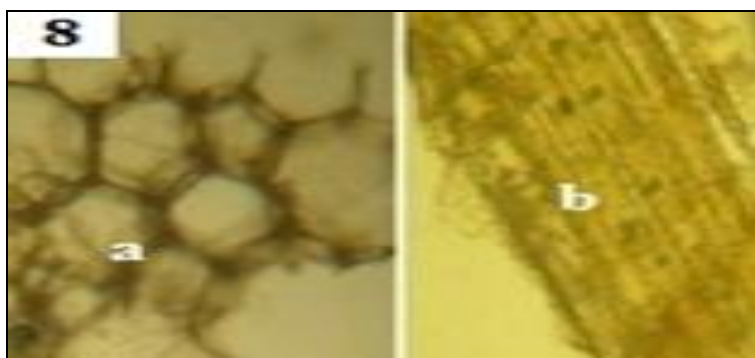


FIG. 8: POWDER MICROSCOPY OF STEM OF *J. MESNYI* AT 45X (A) PARENCHYMATOUS CELLS AND (B) XYLEM FIBRE

Fluorescence Study: The fluorescence behavior of the powder of both stems, moistened with different chemical reagents and solvents; under UV and daylight as shown in **Table 1**.

Physicochemical Parameters and Preliminary Phytochemical Screening: The ash values (total ash, water-soluble ash and acid-insoluble ash

content), extractive values (ethanol-soluble and water-soluble extractive values), loss on drying, swelling index, foaming index and crude fibre content were determined in **Table 2**. The powder of stem was extracted successively using solvents of increasing polarity *i.e.* petroleum ether, chloroform, ethanol and water respectively.

TABLE 1: FLOURESCENCE STUDY OF STEM POWDER WITH DIFFERENT REAGENTS

Reagent	<i>J. multiflorum</i>			<i>J. mesnyi</i>		
	Day light	Short UV (254nm)	Long UV (365nm)	Day light	Short UV (254nm)	Long UV (365nm)
Dry drug	Dark green	Blackish	Green	Light green	Blackish green	Green
Drug + Distilled water	Green	Blackish	Blackish green	Green	Blackish	Blackish green
Drug +1N HCl	Dark green	Blackish	Blackish green	Dark green	Blackish grey	Dark green
Drug+ 50% HCl	Brownish green	Blackish	Blackish green	Green	Blackish	Blackish green
Drug + Acetic acid	Dark green	Blackish	Blackish green	Light green	Blackish	Brownish green
Drug + 50% H ₂ SO ₄	Green	Blackish	Blackish green	Dark green	Smoky black	Blackish green
Drug +1N H ₂ SO ₄	Brownish green	Blackish	Blackish green	Brownish green	Blackish	Brownish green
Drug +1N NaOH (Aq)	Brown	Blackish	Blackish green	Black green	Blackish	Brownish green
Drug +1N NaOH (Alc)	Black green	Blackish	Blackish green	Brown	Blackish	Brownish green
Drug+ 1NHNO ₃	Brown	Blackish	Dark Green	Brownish green	Blackish	Dark Green
FeCl ₃ 5% (Alc)	Brownish green	Blackish green	Blackish green	Green	Blackish green	Blackish green
FeCl ₃ 5% (Aq)	Brownish green	Blackish green	Blackish green	Green	Blackish green	Blackish green

Color and percentage yield of each were observed in **Table 3**. The Preliminary phytochemical screening depicted in **Table 4**, showed the presence of alkaloids, anthraquinone glycosides, saponins glycosides, cardiac glycosides, flavonoids, steroids, terpenoids, tannins and phenolic compounds. The Heavy metal content, aflatoxins and microbial content were observed within given limits as per WHO as shown in **Table 5**. The pesticide residues were observed according to WHO guidelines as given in **Table 6**.

TABLE 2: RESULT FOR PHYSICOCHEMICAL PARAMETERS

Parameters	<i>J. multiflorum</i>	<i>J. mesnyi</i>
Loss on drying	5.73%	6.54%
Total ash	3.92%	4.56%
Water soluble ash	1.75%	2.34%
Acid insoluble ash	1.28%	1.66%
Ethanol extractive	5.67%	8.81%
Water extractive	4.64%	7.16%
Swelling Index	Nil	Nil
Crude fiber content	7.83%	6.53%
Foaming Index	Less than 100	Less than 100

TABLE 3: RESULT FOR SUCCESSIVE EXTRACTS

Extract	<i>J. multiflorum</i>		<i>J. mesnyi</i>	
	Colour	Extractive value (%w/w)	Colour	Extractive value (%w/w)
Petroleum Ether	Blackish yellow	5.85	Yellowish brown	3.81
Choloroform	Yellowish green	6.74	Greenish black	4.56
Ethanol	Yellowish green	8.23	Dark brown	9.98
Aqueous	brownish	7.12	Dark brown	8.96

TABLE 4: RESULTS OF PRELIMINARY PHYTOCHEMICAL SCREENING

Compounds	<i>J. multiflorum</i>				<i>J. mesnyi</i>			
	PE	CH	Et.	Aq.	PE	CH	Et.	Aq.
Alkaloids	-	-	++	++	-	-	++	-
Carbohydrates	-	-	++	++	-	-	++	++
Anthraquinone glycoside	-	-	-	-	-	-	-	-
Saponin glycoside	-	-	++	++	-	-	++	++
Cardiac glycoside	-	-	++	++	-	-	++	++
Coumarin glycoside	-	-	++	++	-	-	++	++
Flavanoids	-	-	++	++	-	-	++	++
Tannins & Phenolic	-	-	++	++	-	-	++	++
Protein and amino acids	-	-	++	++	-	-	++	++
Steroids & Terpenoids	++	++	++	-	++	++	++	-

Key: (++) = Present (-) = Absent # PE- Petroleum ether, CH- Chloroform, Et- Ethanol, Aq- Aqueous.

TABLE 5: PHYSICOCHEMICAL PARAMETERS

Parameter	Specified limit	Value	
		<i>J. multiflorum</i>	<i>J. mesnyi</i>
Microbial Contamination Test			
Total Bacterial Count	1 X 10 ⁵ c.f.u./g	700	500
Total yeast and mould count	1 X 10 ³ c.f.u./g	Nil	Nil
<i>E. coli</i>	Nil	Nil	Nil
<i>Salmonella sp.</i>	Nil	Nil	Nil
<i>S. aureus</i>	Nil	Nil	Nil
<i>P. aeruginosa</i>	Nil	Nil	Nil
Aflatoxin Content			
Aflatoxin B1	0.5ppm	Nil	Nil
Aflatoxin B2	0.1ppm	Nil	Nil
Aflatoxin G1	0.5ppm	Nil	Nil
Aflatoxin G2	0.1ppm	Nil	Nil
Heavy Metal Analysis			
Arsenic	5ppm	ND	0.01
Cadmium	0.3ppm	0.02	0.02
Lead	10ppm	0.01	0.01
Mercury	0.2ppm	ND	ND

ND- not detected.

TABLE 6: DETERMINATION OF PESTICIDES

Pesticides	Specified limit	Value	
		<i>J. multiflorum</i>	<i>J. mesnyi</i>
Alachlor	0.02	ND	ND
Atrazine	-	ND	ND
BHC (sum of all isomers)	0.3	ND	ND
Bifenthrin	-	ND	ND
Butachlor	-	ND	ND
Carbofuran	-	ND	ND
Carbofuran, 3-Hydroxy	-	ND	ND
Chlordane (sum of cis-, alpha-)	0.05	ND	ND
Cypermethrin peak 1	1.0	ND	ND
DDD(sum of all isomers)	1.0	ND	ND
DDE (sum of all isomers)	1.0	ND	ND
Dieldrin	0.05	ND	ND
Dimethoate	0.5	ND	ND
Edifenphos	-	ND	ND
Endosulfan peak 1	3.0	ND	ND
Endosulfan peak 2	3.0	ND	ND
Endosulfan sulphate	3.0	ND	ND
Endrin	0.05	ND	ND
Ethion	2.0	ND	ND
Fenthion	0.5	ND	ND
Fenvalerate	1.5	ND	ND
Heptachlor	0.05	ND	ND
Heptachlor epoxide	0.05	ND	ND
Malathion	1.0	ND	ND
Methoxychlor	-	ND	ND
Parathion-methyl	0.2	ND	ND
Phorate	-	ND	ND
Phoratesulfone	-	ND	ND
Phosalone	0.1	ND	ND

#ND- not detected.

DISCUSSION: Despite the availability of various analytical tools in the field of modern pharmaceuticals, still lacking pharmacognostical standards for traditionally used plants is the major lacuna. Precisely for the reason, the WHO has recommended various physicochemical methods for authentication of plant drugs. These parameters can even help in the detection of adulteration hence they should be adopted to confirm the identity, purity, and quality of plant drugs³⁴.

Macroscopic parameters *viz.*, color, taste, texture, odour, size and shape are helpful for the identification of plant materials. So, quality control of herbal drugs has traditionally been done based on appearance or morphological evaluation; but today microscopic evaluation is indispensable in the initial identification of herbs, as well as in identifying small fragments of crude or powdered herbs, and detection of foreign matter and adulterants. Physicochemical parameters are important parameters in detecting adulteration and

are adopted to confirm the identity, purity, and quality of the drug. Ash values are particularly important parameters as these show the presence or absence of foreign matters like metallic salts and silica etc. The values of total ash, water-soluble ash, and acid-insoluble ash are 3.92%, 1.75% and 1.28% (*J. multiflorum*) and 4.56%, 2.34% and 1.66% (*J. mesnyi*). Loss on drying for both the stem was nearly 5.73% and 6.54% respectively.

It signifies the considerable amount of moisture in the stem. The moisture content of a drug should be below in order to prevent decomposition of crude drug either due to chemical change of constituents or microbial contamination. The result of fluorescence analysis of stem powder showed their characteristic fluorescent color in different organic and inorganic solvents. The fluorescence behavior of powdered drugs plays a vital role in the determination of the identity, quality, and purity of the drug material.

Extractive values are primarily useful for the determination of exhausted or adulterated drugs. The result suggests that the drug has high ethanol-soluble extractive value compared to water-soluble extractive value. Preliminary Phytochemical Evaluation showed the presence of alkaloids, glycosides, saponins, steroids, terpenoids, flavonoids, carbohydrates, protein and amino acid, tannins and phenolic compounds in the stem of both plants. Heavy metal like mercury is not detected in any of the species however the level of cadmium, arsenic and lead are within the limits as per WHO guidelines. The results of microbial content, aflatoxin content, and pesticide residue revealed that both species are free from above the natural contaminants and therefore used for the preparation of formulation³⁴.

CONCLUSION: Standardization studies help in identification, authentication and establishing the quality of the plant material and will be useful in preparing a monograph of the plant. Further, it will act as a tool to detect adulterants and substituents and is helpful in maintaining the reproducibility of quality and efficacy of natural drugs.

ACKNOWLEDGEMENTS: None

CONFLICTS OF INTEREST: The authors have no conflicts of interest regarding this investigation.

REFERENCES:

- Mittal A, Sardana S and Pandey A: Ethnobotanical, Phytochemical and Pharmacological Profile of *Jasminum sambac* (L.): Journal of Pharmaceutical and Biomedical Sciences 2011; 11: 1-7.
- Chatterjee A and Prakash CS: The treatise of indian medicinal plants. National institute of science communication and information resources, CSIR 6th Edition New Delhi 2013; 76-77.
- Nadkarni KM: The Indian material medica popular Prakashan. Second Edition Bombay India 2000; 1: 703.
- Srivastava HC and Karmakar PG: An inventory of jasmine. Indian Horticulture 1989; 35-36.
- Chopra RN, Nayar SL and Chopra IC: Glossary of Indian Medicinal Plants. Council of Scientific & Industrial Research New Delhi 1986; 144.
- Singh D: *J. multiflorum* (Burm.f.) Andr. Botany, Chemistry and Pharmacology. Asian Journal of Chemistry 2016; 28(12): 2575-2578.
- Wiert C: Medicinal Plants of China, Korea and Japan. Bioresources For Tomorrow's Drugs and Cosmetics CRS Press Inc 2012; 454.
- Mamillian HF: Tropical Planting and Gardening, Macmillan Co. Ltd. London 1954; 2-20.
- Bera P, Kotamreddy JN, Samanta T, Malti S and Mitra A: Interspecies variation in headspace scent volatiles

- composition of four commercially cultivated jasmine flowers. Natural Product Research 2015; 29: 1328.
- Pahari SK, Ghosh S, Saha H, Gupta PK and Mondal S: Pharmacognostic and Qualitative Evaluation of the Root of the Plant *Jasminum multiflorum* Andr. Der Pharma Chemica 2017; 9(23): 8-11.
- Ganatra SH, Durge SP and Ramteke AM: Phytochemical Investigation and TLC Analysis of *Jasminum multiflorum* leaves. International Journal of Pharmaceutical Sciences and Research 2013; 4(3): 1135-1139.
- Kumaresan M, Kannan M, Sankari A, Chandrasekha CN and Vasanthi D: Phytochemical screening and Antioxidant activity of *Jasminum multiflorum* (Pink Kakada) leaves and flowers. Journal of Pharmacognosy and Phytochemistry 2019; 8(3): 1168-1173.
- Khidzir MK, Cheng SF and Chuah CH: Interspecies variation of chemical constituents and antioxidant capacity of extracts from *Jasminum sambac* and *Jasminum multiflorum* grown in Malaysia. Industrial Crops and Products 2015; 74: 635-641.
- Sharma A, Sati SC, Rawat S and Tomar A: Phytochemical study and antimicrobial activities of *Jasminum multiflorum*. World Journal of Pharmacy and Pharmaceutical Sciences 2014; 3(4): 735-742.
- Pal DK, Pahari SK and Mishra AK: Anthelmintic Activities of Roots of *Cocos nucifera* and aerial Parts of *Jasminum multiflorum*. Asian Journal of Chemistry 2007; 19(7): 5089-5092.
- Romabati N, Joymati L and Dhanachand C: Effect of the plants extract on *Meloidogyne incognita*. Journal of Applied Zoological and Research 1999; 10: 133.
- Verma R, Kumari R, Agnihotri R and Katiyar A: Therapeutic role of Indian flowers in treatment of gastrointestinal Diseases caused by *Vibrio* species. Research Inventory. International Journal of Engineering and Science 2013; 3(3): 20-24.
- Shen C YA, Lin CY and Chen Chung Hsiung: Secoirridoids glycoside from *Jasminum multiflorum*. Phytochemistry 1990; 29(9): 2905-2912.
- Chen HY, Shen Yc and Chen CH: Novel Secoirridoid lactones from *Jasminum multiflorum* Journal of Natural Products 1991; 54(5): 1087.
- Addae J, Pingal R, Walkins K, Cruickshank R, Youssef F F, Navak SB: Effects of *Jasminum multiflorum* leaf extract on rodent models of epilepsy, motor coordination and anxiety. Epilepsy Research 2017; 131: 58-63.
- Pal DK and Pahari SK: Evaluation of CNS Activities of aerial Parts of *Jasminum multiflorum* Andr. Asian Journal of Chemistry 2007; 19(6): 4452-4458.
- Sani P, Verma PK: Evaluation of the Wound Healing Properties of *Jasminum mesnyi* Hance in Diabetic Rats. Annals of Pharmacology and Pharmaceutics 2017; 18: 1-3.
- Borar S, Punia P, Kalia AN: Antioxidant potential of n-butanol fraction from extract of *Jasminum mesnyi* Hance leaves. Indian Journal of Experimental Biology 2011; 49: 39-43.
- Poonia P, Niazi J, Chaudhary G and Kalia AN: *In-vitro* antioxidant potential of *Jasminum mesnyi* Hance (Leaves) extracts. Research Journal of Pharmaceutical Biological and Chemical Sciences 2011; 1: 348-357.
- Farheen M and Farheen S: Phytochemical screening and Antiulcer activity of *Jasminum mesnyi* and *Triticum aestivum* leaves in albino Wistar rats. International Journal of Farming and Allied Sciences 2015; 1: 1-11.
- Bhushan B, Sardana S and Bansal G: Anti-Diabetic Potentials of *Clerodendrum inerme*, *Jasminum mesnyi*

- Hance* and *Callistemon citrinus* on Nicotinamide-Streptozotocin Induced Type 2 Diabetic Rats. International Journal of Phytomedicine 2015; 2: 136-141.
27. Dullu V: Anthelmintic activity of Ethanolic leaf extract of *Jasminum mesnyi*. Asian Pacific Journal of Tropical Diseases 2014; 1: 273-275.
 28. Verma R, Balaji BS and Dixit A: Phytochemical analysis and broad spectrum antimicrobial activity of ethanolic extract of *Jasminum mesnyi* Hance, leaves and its solvent partitioned fractions. Bioinformation 2018; 8: 430-438.
 29. Kokate CK: Practical Pharmacognosy. First edition Vallabh Prakashan. New Delhi 1994; 15-30.
 30. Trease and Evans: Pharmacognosy W.B. Saunders. International Edition 2005; 15: 456-48.
 31. The Ayurvedic Pharmacopoeia of India, Ministry of Health and Family Welfare, Government of India, Department of Indian system of medicines and Homeopathy. First edition New Delhi; A53-A55.
 32. WHO: Quality control methods for medicinal plant material, Geneva, APTBS publisher and distributor. New Delhi 2002; 46: 22-34.
 33. Khandelwal KR: Practical Pharmacognosy: Techniques and Experiment. Thirteen Edition Nirali Prakashan. Pune 2005.
 34. Mittal A, Sardana S and Pandey A: Pharmacognostical Profiling of *Jasminum grandiflorum* Linn. Leaves. International Journal of Phytomedicine 2015; 7: 100-105.

How to cite this article:

Garg P and Garg V: Phytochemical and pharmacognostical screening of *Jasminum multiflorum* (Burm. F.) and *Jasminum mesnyi* (Hance) stems. Int J Pharm Sci & Res 2022; 13(10): 4035-43. doi: 10.13040/IJPSR.0975-8232.13(10).4035-43.

All © 2022 are reserved by International Journal of Pharmaceutical Sciences and Research. This Journal licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported License.

This article can be downloaded to **Android OS** based mobile. Scan QR Code using Code/Bar Scanner from your mobile. (Scanners are available on Google Playstore)