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RP-HPLC METHOD FOR SIMULTANEOUS DETERMINATION OF ATORVASTATIN CALCIUM, OLMESARTAN MEDOXOMIL, CANDESARTAN, HYDROCHLOROTHIAZIDE AND CHLORTHALIDONE – APPLICATION TO COMMERCIALY AVAILABLE DRUG PRODUCTS

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ABSTRACT

A simple, precise and stability-indicating HPLC method was developed and validated for the simultaneous determination of anti-hypertensive drugs Atorvastatin Calcium, Olmesartan Medoxomil, Candesartan, diuretics Hydrochlorothiazide and Chlorthalidone. The separation was achieved on Cosmosil PAQ (Length 150 mm × Diameter 4.6 mm Particle size 5 μm) column with gradient flow. The mobile phase at a flow rate of 1.0 mL min⁻¹ consisted of 0.05 M sodium dihydrogen phosphate buffer and acetonitrile (Gradient ratio). The UV detection was carried out at 220 nm. The method was successfully validated in accordance to ICH guidelines. Further, the validated method was applied for commercially available pharmaceutical dosage form.

INTRODUCTION: The parent guideline on drug stability testing Q1A (R2) issued by International Conference on Harmonization (ICH) stipulates stress studies to be carried out on a drug in order to establish the drug's inherent stability characteristics¹⁻². Literature studies show various analytical methods reported for the estimation of individual, binary or tertiary combination of anti-hypertensive drugs or in combination with diuretics³⁻¹². Recently, HPLC method with fluorescence detection for simultaneous determination of chlorthalidone, valsartan and fluvastatin from human plasma has been reported¹³.

The HPLC-MS/MS method for simultaneous estimation of atenolol, bisoprolol, hydrochlorothiazide, chlorthalidone, salicylic acid, enalapril and its active metabolite enalaprilat, valsartan and fluvastatin is also reported¹⁴.

However, so far, no method was reported for the simultaneous determination in combination for Atorvastatin Calcium, Olmesartan Medoxomil, Candesartan, Hydrochlorothiazide, Chlorthalidone and its application to pharmaceutical samples.

An attempt was made in this study to develop a rapid, economical, precise and accurate stability-indicating assay method for simultaneous estimation of Atorvastatin Calcium, Olmesartan Medoxomil, Candesartan, Hydrochlorothiazide and Chlorthalidone in tablet formulation.

The proposed method is rapid, simple, accurate, and reproducible, and can be successfully employed in the routine analysis of both these drugs simultaneously, in tablet dosage form.

MATERIAL AND METHODS:

- **Chemicals and reagents:** Drug substances were provided by Sharon Biomedicine, India. All the chemicals and reagents sodium hydroxide, hydrochloric acid, monobasic sodium dihydrogen phosphate, hydrogen peroxide (30 %) was used of Analytical grade while, Acetonitrile was procured from Merck (Germany). A Millipore Milli Q plus water purification system (Milford, USA), was used to prepare distilled water ($>18 \mu\Omega$). The commercially available drug products were used as OLSAR 20 (Omlsartan - 20 mg, Unichem Laboratories, India), OLMAX H (Olmesartan Medoxomil - 20 mg and Hydrochlorothiazide - 12.5 mg; Glenmark, India), Atacor-5 (Atorvastatin Calcium 5 mg, Dr. Reddys, India).
- **Instruments:** Integrated HPLC system, Ultimate 3000 manufactured by Dionex (Germany) was used for method development and method validation. This system comprised of a quaternary gradient pump, auto sampler, column oven and a photodiode array detector. PC installed Chromeleon software was used to record and integrates the chromatograms. The analysis was carried out at ambient temperature. Photostability studies were performed in a photostability chamber, from Thermolab (India).
- **Chromatographic conditions:** Cosmosil PAQ (Length 150 mm \times Diameter 4.6 mm Particle size 5 μm) analytical column from Nacalai Tesque INC, Japan was used as a stationary phase. The flow rate was 1.0 mL min^{-1} and the detector was set at 220 nm. The volume of the sample solution injected was 10 μL . The gradient mobile phase consisted of 0.05 M sodium dihydrogen phosphate buffer and acetonitrile (Mobile Phase-B) with the gradient as mentioned in **Table 1**. A membrane filter of 0.45 μm porosity was used to filter and degas the mobile phase.
- **Standard and Test solutions:** Weighed accurately about 50 mg of each Atorvastatin Calcium, Olmesartan Medoxomil, Candesartan, Hydrochlorothiazide and Chlorthalidone reference standard into 50 ml volumetric flask. Added to it 20.0 ml methanol and sonicated to dissolve. Diluted this solution up to volume with diluent (Buffer: Methanol 30:70 v/v). Pipette out 5.0 ml of this solution into 50 ml volumetric flask and diluted to volume with diluent. ($100 \mu\text{g mL}^{-1}$ each of Atorvastatin Calcium, Olmesartan Medoxomil, Candesartan, Hydrochlorothiazide and Chlorthalidone). Similarly, the test solutions were prepared at same concentration using same diluents. ($100 \mu\text{g mL}^{-1}$ of each).

TABLE 1: MOBILE PHASE GRADIENT FOR CHROMATOGRAPHIC METHOD

Time (min)	% 0.05 M sodium dihydrogenphosphate buffer	% Acetonitrile
0	70	30
3	70	30
8	40	60
11	20	80
15	20	80
18	70	30

Method Development: A variety of mobile phases were investigated in the development of a stability-indicating LC method for the analysis of Atorvastatin Calcium, Olmesartan Medoxomil, Candesartan, Hydrochlorothiazide and Chlorthalidone drug substances. The suitability of mobile phase was decided on the basis of selectivity and sensitivity of the assay, stability studies and separation among impurities formed during forced degradation studies.

- **Wavelength Selection:** The individual drug substance solution at concentration of $100 \mu\text{g mL}^{-1}$ in diluent was scanned on PDA from 190 nm to 400 nm. The maximum wavelength were observed Hydrochlorothiazide (224 nm and 271 nm), Chlorthalidone (196 nm), Olmesartan Medoxomil (196 nm and 258 nm), Atorvastatin Calcium (194 nm and 246 nm), Candesartan (197 nm, 253 nm and 305 nm). However detection

was carried out at 220 nm on basis of higher

response (**Fig. 1 and Table 2**).

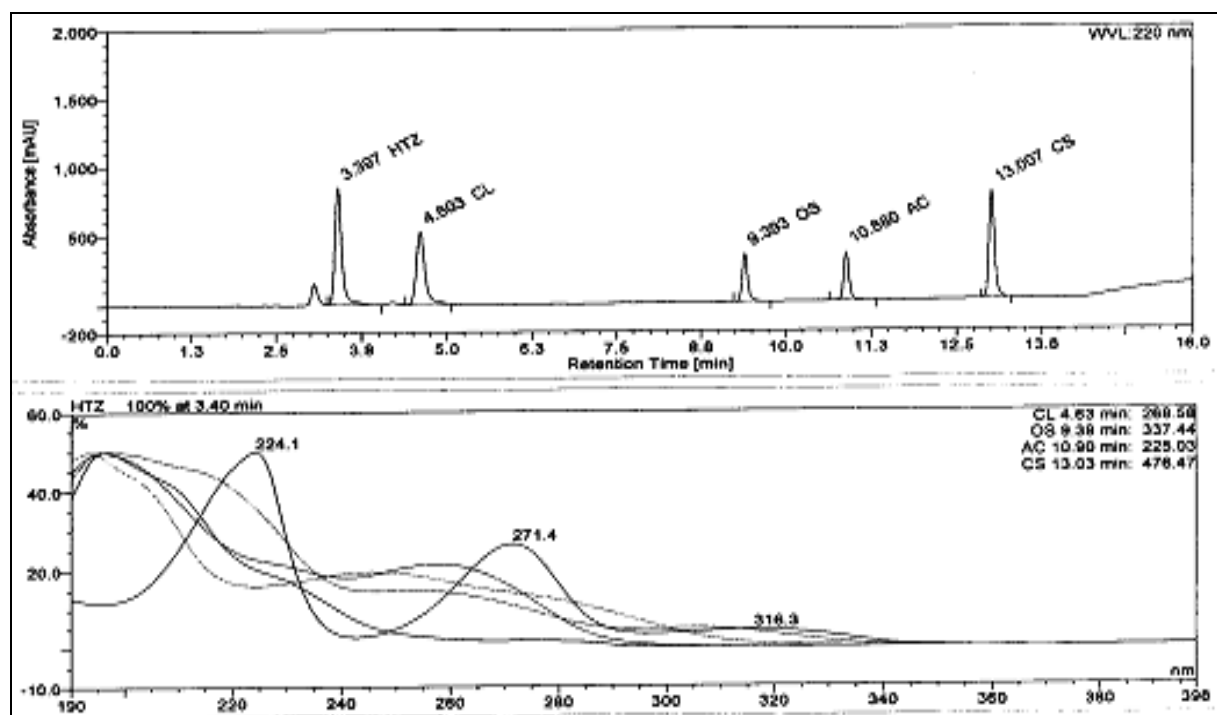


FIG. 1: UV SPECTRUM FOR DRUG SUBSTANCES IN DILUENTS

TABLE 2: AREA RESPONSE OF PEAKS AT DIFFERENT WAVELENGTHS

Wavelength (nm)	HTZ	CL	OS	AC	CS
220	99.732	76.133	31.913	31.504	72.216
230	60.948	54.561	25.560	30.871	48.029
240	7.573	29.388	23.006	34.822	28.576
250	9.9846	11.406	25.315	35.286	26.230
260	32.419	6.753	26.583	30.895	25.263
280	36.045	4.314	9.704	22.405	13.446
300	7.720	0.130	0.372	8.941	8.546

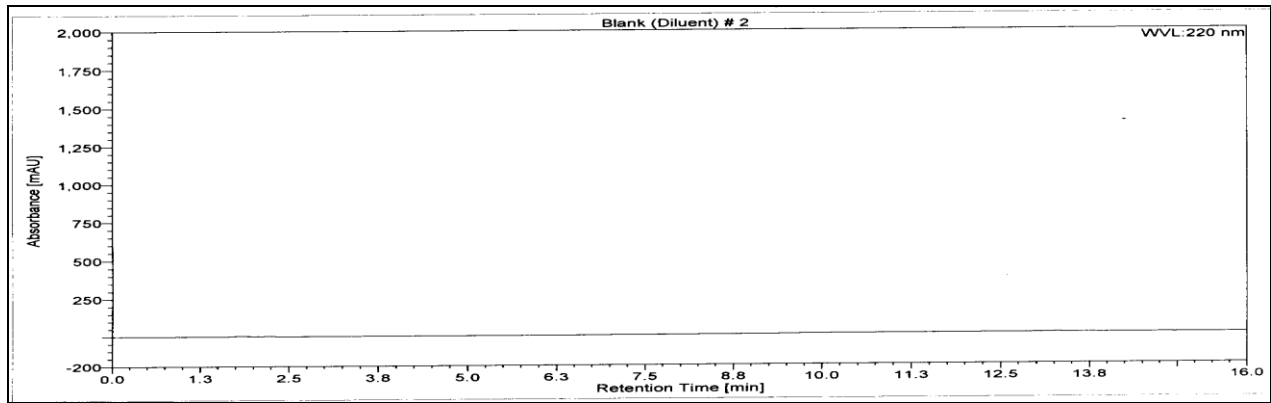
Method Validation: The optimized chromatographic conditions were validated by evaluating specificity-Forced degradation, linearity, precision, accuracy, robustness and system suitability parameters in accordance with the ICH guideline Q2 (R1) ².

Specificity- Forced Degradation Study:

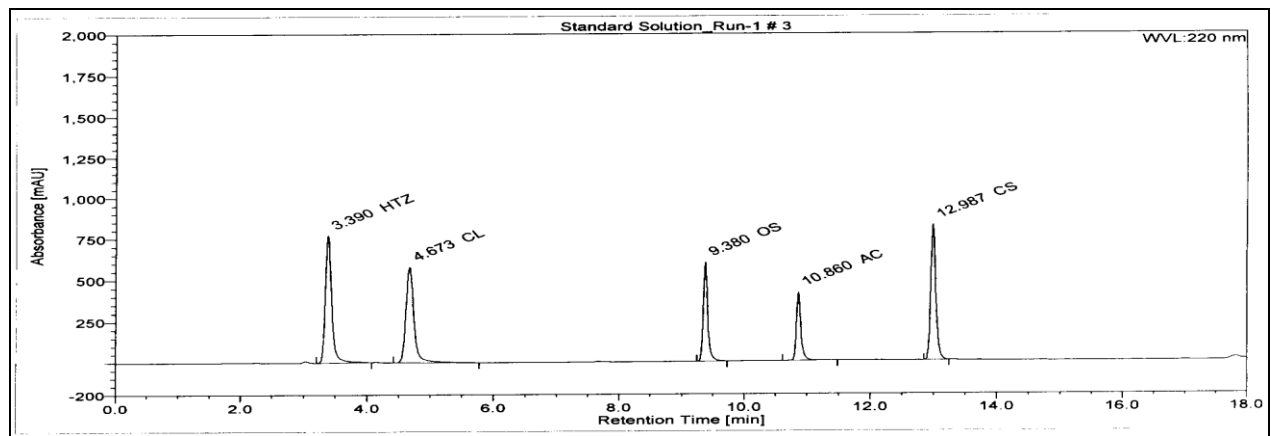
- **Acid Hydrolysis:** Forced degradation study was conducted on 5 ml stock solution of standard solution containing $1000 \mu\text{g mL}^{-1}$ of each drug substances by exposing with 5ml of 1N hydrochloric acid for 3 hours at room temperature. Then neutralized with base (when necessary) and dilute up to 50 ml with diluent.

- **Base Hydrolysis:** Forced degradation study was conducted on 5 ml stock solution of standard solution containing $1000 \mu\text{g mL}^{-1}$ of each drug substances by exposing with 5ml of 0.1N sodium hydroxide for 30 minutes at room temperature. Then neutralized with acid and dilute up to 50 ml with diluent.
- **Oxidation:** Forced degradation study was conducted on 5 ml stock solution of standard solution containing $1000 \mu\text{g mL}^{-1}$ of each drug substances by exposing with 5ml of 5% H_2O_2 for 30 minutes at room temperature. Then dilute up to 50 ml with diluent.
- **Thermal degradation:** Solid drugs powder was kept in dry oven at 105°C for 24 hours.
- **Photolysis:** Photolysis studies were carried out on solid drugs, their combination and their dosage form. The sample in a petri plate was spread as a thin layer (1 mm) and exposed to light in a photostability chamber. The method's analytical data were collected at a single wavelength of 220 nm. Additional PDA detector data were collected for the peak purity evaluation. The chromatograms were extracted

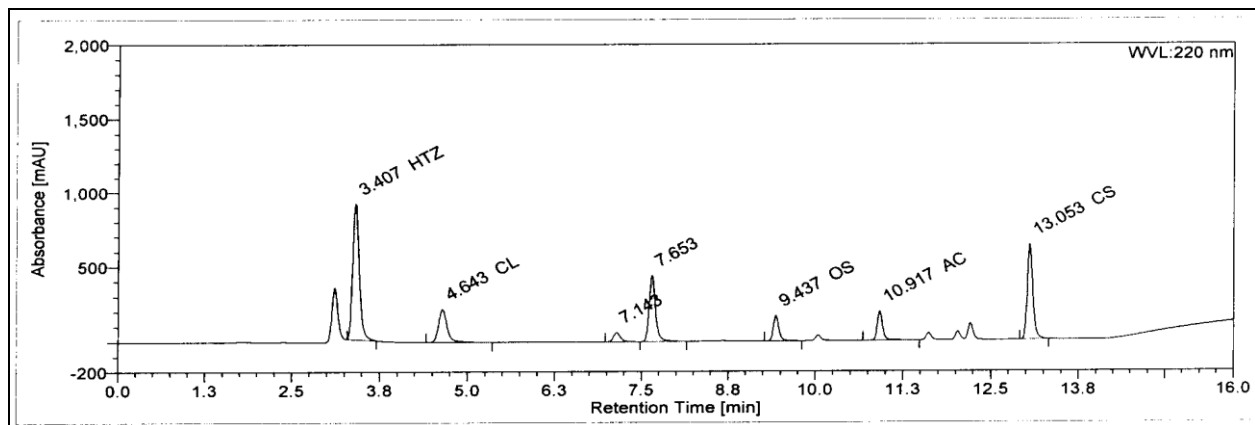
for Peak purity and demonstrated as in **Fig. 2a, b, c, d, e, f, g** and **Table 3.**



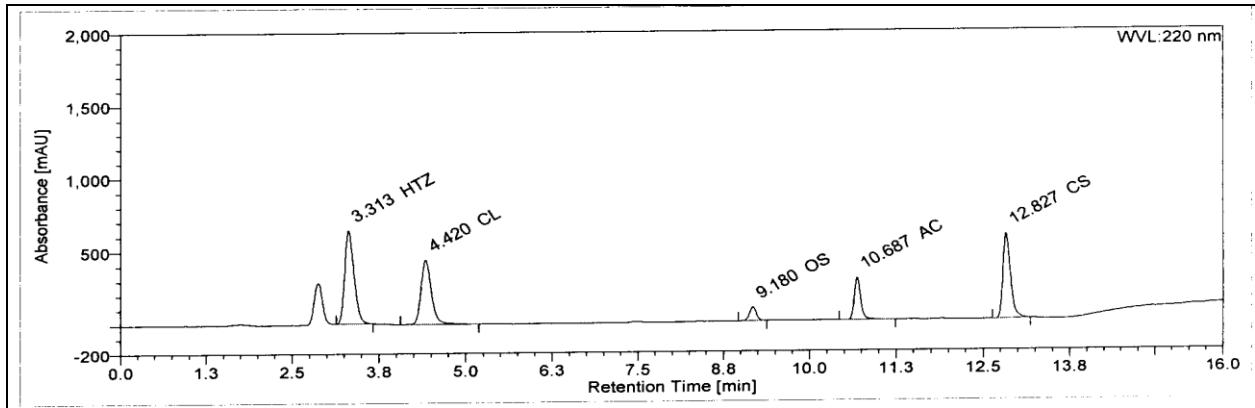
a) Blank solution



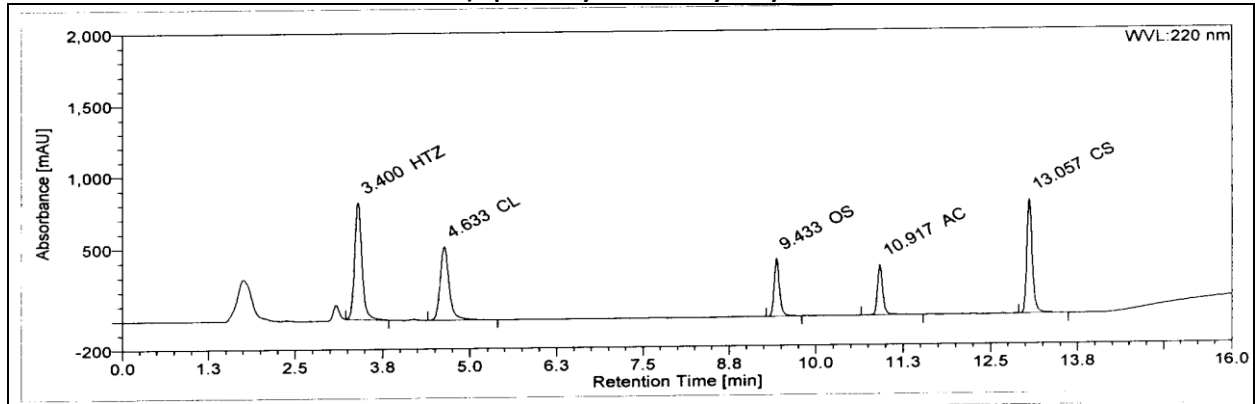
b) Standards solution



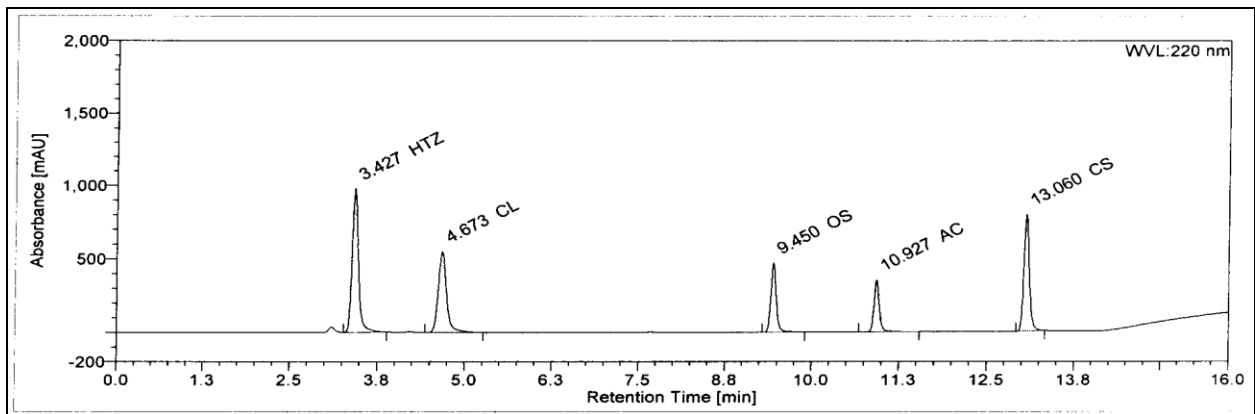
c) Specificity-Acid hydrolysis



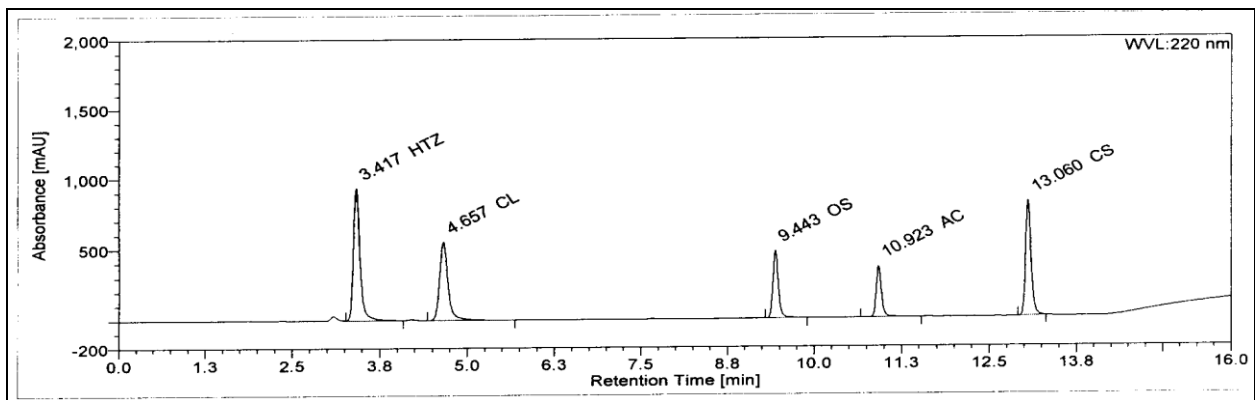
d) Specificity-Alkaline hydrolysis



e) Specificity-Peroxide oxidation



f) Specificity Photolytic degradation



g) Specificity-Thermal degradation

FIG. 2: CHROMATOGRAMS FOR STRESSED CONDITIONS

TABLE 3: RESULTS FOR FORCED DEGRADATION STUDY

Degradation Condition		HTZ	CL	OS	AC	CS
Acid hydrolysis	% Degradation	0.7	60.0	63.9	44.8	20.1
	Peak purity	1000	1000	1000	1000	1000
Alkaline hydrolysis	% Degradation	0.7	0.8	73.2	2.2	2.3
	Peak purity	1000	1000	1000	1000	1000
Peroxide Oxidation	% Degradation	4.8	1.6	14.9	2.4	1.1
	Peak purity	999	1000	1000	998	1000
Photolytic degradation	% Degradation	0.7	1.36	0.0	0.2	0.2
	Peak purity	1000	1000	1000	998	1000
Thermal degradation	% Degradation	1.2	0.0	0.0	0.0	0.0
	Peak purity	1000	1000	1000	998	1000

Linearity: Standard stock solution of the drug was diluted to prepare linearity standard solutions in the concentration range of 10 –150 $\mu\text{g mL}^{-1}$ for all Hydrochlorothiazide, Chlorthalidone, Olmesartan Medoxomil, Atorvastatin Calcium, and Candesartan. Three sets of such solutions were prepared. Each set was analyzed to plot a calibration curve. Slope, intercept and coefficient of determination (r^2) of the calibration curves were calculated to ascertain linearity of the method (Fig. 3 and Table 4).

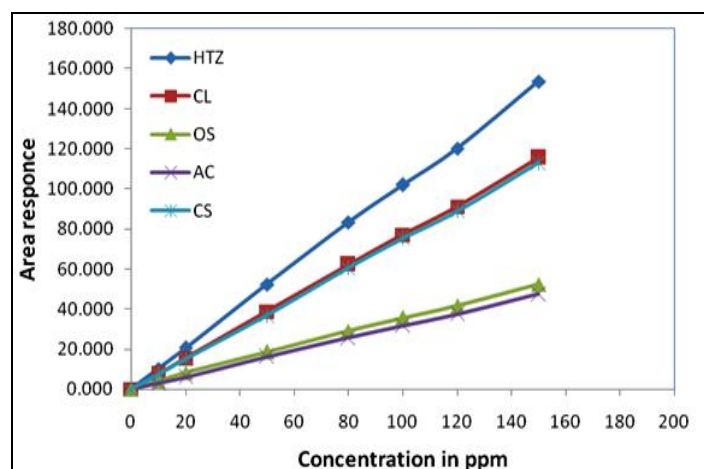


FIG. 3: LINEARITY CURVE WITH CORRELATION CO-EFFICIENT

Recovery: Recovery of the method was determined by analyzed the drug products and synthetic mixture of drug products with 50%, 100% and 150% levels. These

TABLE 4: SLOPE, INTERCEPT AND REGRESSION COEFFICIENT

Drug substance	Slope	Intercept	Regression(r^2)
Hydrochlorothiazide (HTZ)	1.016	0.548	1.000
Chlorthalidone (CL)	0.755	-0.225	1.000
Olmesartan Medoxomil (OS)	0.444	1.179	1.000
Atorvastatin Calcium (AC)	0.928	-1.202	1.000
Candesartan (CS)	2.370	0.053	1.000

TABLE 5: RECOVERY FROM COMMERCIALY AVAILABLE SAMPLES

mixtures were analyzed by the proposed method. The experiment was performed in triplicate and recovery (%); RSD (%) were calculated (Table 5 and 6).

Precision: The precision of the proposed method was evaluated by carrying out six independent assays of test samples. RSD (%) of six assay values obtained was calculated. Intermediate precision was carried out by analyzing the samples by a different analyst on another instrument (Table 7).

Robustness: The robustness was studied by evaluating the effect of small but deliberate variations in the chromatographic conditions. The conditions studied were wavelength (altered by ± 0.3 nm), column oven temperature (altered by ± 5 °C) and pH of buffer in mobile phase (altered by ± 0.2). These chromatographic variations were evaluated for resolution between all drug substances (Table 8 to 10).

Solution stability: To assess the solution stability, standard and test solutions were kept at 25 °C (laboratory temperature) for 24 hrs (Table 11).

System Suitability: The system suitability parameters with respect to theoretical plates, tailing factor, repeatability and resolution between peaks of all drug substances were defined (Table 12).

% Recovery	OLSAR 20 Tablets		OLMAX H Tablets		Atocor Tablets	
	OS 20mg (Unichem Lab Ltd)		OS 20mg and HTZ 12.5mg (Glenmark Pharma Ltd)		AC 5mg (Dr. Reddys Ltd)	
	Recovery (%) For OS		Recovery (%) For OS		Recovery (%) For HTZ	Recovery (%) For AC
50 %	99.45		98.56		102.37	101.08
	100.13		99.49		98.98	100.72
	98.65		98.70		101.13	99.30
100 %	102.00		98.24		101.10	98.70
	101.50		100.02		101.88	101.61
	101.93		98.40		101.21	100.90
150 %	101.08		98.91		99.58	101.68
	98.85		99.53		100.19	99.86
	101.76		100.87		100.80	101.91
Avg	100.60		99.02		100.86	100.64
RSD	1.34		0.87		1.06	1.11

TABLE 6: RECOVERY ON SYNTHETIC MIXTURE OF ALL FIVE DRUG SUBSTANCE

% Recovery	% Assay				
	HTZ	CL	OS	AC	CS
Level – 50%	101.27	100.14	100.72	100.59	99.75
	98.20	98.57	102.34	99.83	97.32
	99.83	99.68	99.96	102.18	99.41
Level – 100%	102.64	102.80	101.68	100.04	101.84
	99.82	97.87	100.46	99.72	100.43
	100.69	100.32	101.00	101.07	98.22
Level – 150%	99.96	100.88	100.71	100.28	101.08
	97.52	97.76	100.44	98.83	99.64
	99.25	100.80	99.63	100.80	97.99
Average	99.91	99.87	100.77	100.37	99.52
RSD	1.55	1.60	0.83	0.94	1.51

*Chlorthalidone and Candesartan drug substances spiked with Olmax H and Atocor drug products.

TABLE 7: PRECISION

Sr. Nos.	% Assay - Repeatability					% Assay - Intermediate Precision				
	HTZ	CL	OS	AC	CS	HTZ	CL	OS	AC	CS
1	98.04	100.74	101.19	100.46	99.99	101.76	98.20	99.78	98.31	99.13
2	99.82	101.89	99.50	100.21	100.20	99.72	100.11	99.73	101.20	99.80
6	100.02	100.15	99.65	100.17	100.14	99.70	100.02	99.83	99.88	100.01
4	98.57	99.60	98.66	99.94	98.35	99.98	100.17	101.65	101.92	101.08
5	100.11	100.33	99.73	98.51	101.96	100.04	99.95	98.42	99.89	98.47
6	99.97	100.38	101.77	99.10	100.13	101.04	99.94	98.46	100.35	99.84
Avg	99.42	100.52	100.08	99.73	100.13	100.39	99.73	99.65	100.26	99.72
RSD	0.89	0.77	1.16	0.76	1.14	0.82	0.76	1.19	1.24	0.88
Average for Precision and Intermediate precision						99.79	100.14	99.99	99.96	99.93
RSD for Precision and Intermediate precision						0.96	0.83	1.14	1.02	1.00

TABLE 8: CHANGE IN WAVELENGTH

Wavelength	% Assay				
	HTZ	CL	OS	AC	CS
217 nm	99.79	99.91	99.69	99.95	99.77
220 nm	99.93	100.03	99.80	100.33	100.25
223 nm	99.59	99.92	99.71	100.06	99.84

Average	99.77	99.95	99.73	100.11	99.95
RSD	0.17	0.07	0.06	0.20	0.26

TABLE 9: CHANGE IN COLUMN OVEN TEMPERATURE

Column oven temperature	% Assay				
	HTZ	CL	OS	AC	CS
20°C	99.98	100.14	99.76	100.16	100.08
25°C	99.93	100.03	99.80	100.33	100.25
30°C	99.89	100.07	99.74	100.35	100.29
Average	99.93	100.08	99.77	100.28	100.21
RSD	0.05	0.06	0.03	0.10	0.11

TABLE 10: CHANGE IN pH OF BUFFER SOLUTION IN MOBILE PHASE

pH of buffer	% Assay				
	HTZ	CL	OS	AC	CS
pH 4.0	99.93	100.32	99.77	100.30	100.32
pH 4.2	99.93	100.03	99.80	100.33	100.25
pH 4.4	99.90	100.17	99.79	100.17	100.06
Average	99.92	100.17	99.79	100.27	100.21
RSD	0.02	0.14	0.02	0.08	0.13

TABLE 11: RESULTS FOR SOLUTION STABILITY

Time (Hours)	% Assay				
	HTZ	CL	OS	AC	CS
Initial	99.87	100.13	99.79	99.97	100.06
3	99.88	100.52	99.30	99.58	100.11
6	99.73	99.97	79.29	99.25	100.57
12	96.90	98.77	62.90	97.38	100.62
18	96.46	97.04	55.31	95.19	100.84
24	96.08	95.66	53.75	94.75	96.56

TABLE 12: CHROMATOGRAPHIC PARAMETERS OF SYSTEM SUITABILITY

Drug substances	RT (min)	Theoretical plates	Symmetry	Resolution	Peak purity
HTZ	3.42	6612	1.42	-	1000
CL	4.65	7428	1.34	6.34	1000
OS	9.42	71558	1.20	26.82	999
AC	10.90	101530	1.24	10.67	999
CS	13.04	132362	1.22	15.26	1000

RESULTS AND DISCUSSION:

HPLC Method Development: The maximum absorption wavelength of the reference drug solution and of the forcefully degraded drug solution was found to be 220 nm. This was observed from the UV absorption spectra (Fig. 1) and was selected as detection wavelength for LC analysis. The main objective of this

chromatographic method was separation of degraded impurities from all drugs.

Forced degradation study revealed a critical separation of closely eluting impurity formed from the Hydrochlorothiazide, Chlorthalidone, Olmesartan Medoxomil, Atorvastatin Calcium and Candesartan peaks. PAQ (Length 150 mm × Diameter 4.6 mm, Particle size 5 µm) helped in resolving all peaks as the

column had carbon loading approx 11% against conventional ODS. This effect was observed by using the mobile phase 0.05 sodium hydrogen phosphate (pH 4.2) and acetonitrile in the gradient ratio.

Summary of Validation Parameters: The assay test method is validated for Specificity, Linearity, Precision, Accuracy (Recovery), Stability of Analytical Solution and Robustness and was found to be meeting the predetermined acceptance criteria.

The validated method is Specific, Linear, Precise, Accurate and Robust for determination of assay of Atorvastatin Calcium, Olmesartan Medoxomil, Candesartan, diuretics Hydrochlorothiazide and Chlorthalidone drug substances and drug products.

Hence, this method can be introduced into routine and stability analysis for the assay of Atorvastatin Calcium, Olmesartan Medoxomil, Candesartan, diuretics Hydrochlorothiazide and Chlorthalidone drug substances.

CONCLUSION: The stability indicating RP-HPLC assay method was developed and validated for simultaneous determination of Atorvastatin Calcium, Olmesartan Medoxomil, Candesartan, diuretics Hydrochlorothiazide and Chlorthalidone drug substances and drug products. The method was found to be simple, specific, Precise and Robust and can be applied for the routine and stability analysis for commercially available formulation.

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