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EFFECT OF pH OF ENTERIC POLYER ON DISSOLUTION PROFILE OF DULOXETINE HCl DELAYED RELEASE PELLETS AT VARIOUS pH RANGES

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ABSTRACT: The main objective of the present study is to evaluate the effect of pH of enteric polymer [methacrylic acid ethyl acrylate copolymer dispersion (Eudragit L30 D-55)], on dissolution profile of Duloxetine HCl delayed release formulation with the T_{lag} of 2hrs. The said formulation was formulated using fluidized bed process in three separate layers, the drug layer, the barrier layer and the enteric layer, were coated on sugar spheres. The enteric coated pellets were top coated using a film coating material, filled in hard gelatin capsules. Eudragit L30-D55 is used as such and neutralized to different pH (pH 4.00, pH 4.25, pH 4.50, pH 4.75, pH 5.00 and pH 5.25), coated on barrier coated pellets. The formulation coated by using enteric polymer neutralized to pH 4.75 shows the maximum extent of dissolution (91.8%) in 0.1N HCl for 2 hrs followed by pH 5.5 phosphate buffer at 120mins.

INTRODUCTION: Enteric polymers are mainly applied as enteric-coatings to conventional solid dosage forms such as tablets, capsules or pellets, which are designed to be insoluble in gastric fluid and soluble in intestinal fluid. They consist of a long-chain polymer with ionizable carboxyl groups. In low-pH environment, the acidic groups are protonized and the polymer is lipophilic. A dramatic change in pH occurs when the dosage form is emptied into the duodenum. The application of an enteric coating to a solid dosage form is a well-established approach to prevent drug release in the stomach and allow release in the small intestine.



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It is used to preclude the degradation of acid-labile actives in the gastric environment or to protect the stomach from irritant compounds ¹.

The commonly used enteric coatings employ pH dependent polymers which contain carboxylic groups. These remain un-ionized in the low pH environment of the stomach, and become ionized in the higher pH conditions of the small intestine, thus allowing the dissolution of the coating and drug release

The *in vitro* dissolution performance of enteric coatings is usually assessed in compendial pH 6.8 phosphate buffer. In this medium, drug release is typically rapid ^{2, 3}.

However, neither does this reflect the *in vivo* performance of enteric coated products, nor it is sufficient to discriminate the dissolution behaviour between different enteric coatings.

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In vivo gamma scintigraphy studies have shown that there is a substantial time delay (up to 2 h) for such products to disintegrate in the human small intestine post gastric emptying, with different enteric polymer coatings exhibiting varying disintegration times ^{4, 5, 6}. Hence, the dissolution is recommended to be evaluated at lower pH like pH 6.0 phosphate buffer and pH 5.5 phosphate buffer, instead of pH 6.8 phosphate buffer ⁷.

The typical value of pH and mean residence time in various segments of the GI tract of young healthy volunteers is summarized in **table 1**.

TABLE 1:

TABLE I:		
Segment	pH (range)	MRT (Mean residence time for pellets)
Pre-prandial		
stomach		
Stomach	1.8 (1 - 3)	30 min
Duodenum	6.0 (4-7)	< 10 min
Upper jejunum	6.5 (5.5-7)	60 min
Lower jejunum	6.8 (6-7.2)	60 min
Upper ileum	7.2 (6.5 -7.5)	60 min
Lower ileum	7.5(7-8)	60 min
Proximal colon	5.5 - 6.5	4-12 hrs
Post-prandial		
stomach		
Stomach	4 (3-6)	2-4 hrs
Duodenum	5.0(4-7)	< 10 min
Upper jejunum	5.5 (5.5-7)	60 min
Lower jejunum	6.5(6-7.2)	60 min
Upper ileum	7.2(6.5 - 7.5)	60 min
Lower ileum	7.5 (7-8)	60 min
Proximal colon	5.5- 6.5	4-12 hrs

Orally administered duloxetine hydrochloride is well absorbed 8 . There is a median 2 hour lag until absorption begins (T_{lag}), with maximal plasma concentrations (C_{max}) of duloxetine occurring 6 hours post dose. Food does not affect the C_{max} of duloxetine, but delays the time to reach peak concentration from 6 to 10 hours and it marginally decreases the extent of absorption (AUC) by about 10%. There is a 3 hour delay in absorption and a one-third increase in apparent clearance of duloxetine after an evening dose as compared to a morning dose. Under post- prandial condition, the formulation will be at around pH 5.5.

MATERIALS AND METHODS:

Materials: The following chemicals were obtained from commercial suppliers and used as received:

Duloxetine Hydrochloride (Orchid chemicals and pharmaceuticals, Chennai), Sugar spheres (Werner, USA), HPMC 5cps (Dow chemicals), Eudragit L100-55 was purchased from Evonic Industries, Talc (Luzenac, Italy), triethyl citrate(Morflex, Greensboro, USA),New Delhi,. All chemicals were reagent grade or higher.

Digital weighing balance (C-220) (make: Saritorious), Remi mechanical propellant stirrer (RA124) (make: Remi), Fluid bed processor (GPCG 1.1) (make: pam glatt), Automatic capsule filling machine (AFT-Lab) (make: Pam machineries), Tray drier (make: Ganson engg), Double beam UV Visible spectrophotometer (Lambda 20) (make: Perkin Elmer), Digital Dissolution test apparatus (D58000) (lab India),

Methods:

Preparation of Pellets ^{8, 9}: Formulation of duloxetine delayed release pellets involves 4 stages

a) Stage – I : Drug layering

b) Stage - II: Barrier coating

c) Stage - III: Enteric coating

d) Stage - IV: Top coating

Stage I:

Preparation of Duloxetine HCl drug layered pellets:

- Binder solution was prepared by dissolving 42 g of povidone (PVP K30) in 1546g of purified water, under stirring using Remi propellant stirrer. Stirring was continued for 45 minutes to get clear solution.
- Duloxetine HCl was added to the above solution under stirring, mixed by using Remi propellant stirrer for 30 minutes to get uniform homogeneous dispersion.
- Corn starch, sodium chloride and talc are added to the above binder solution, mixed by using Remi propellant stirrer for 30 minutes to get uniform homogeneous dispersion.
- The resulting suspension was filtered through ASTM 60#.

- Sugar spheres (710-850μ) of 480g were loaded in Fluid bed processor, GPCG 1.1, with bottom spray assembly, drug layering suspension was coated on sugar spheres. The process parameters are tabulated in table 3.
- The solid content of drug layering suspension was 20% w/w.
- Each formula was having the batch size of 4000 units.
- To achieve 100% drug layering, the overage of 5% was used in the formulation.

Stage II:

Preparation of Duloxetine HCl barrier coated pellets:

- Barrier coating solution was prepared by dispersing Opadry white Y-1-7000 in purified water, mixed for 45 minutes, using Remi propellant stirrer.
- 10% w/v solution was prepared
- The resulting suspension was filtered using ASTM 60#, and coated on 217g of drug layered pellets using fluid bed processor (GPCG 1.1), to achieve 20% build up.
- During the preparation of coating solution the 10% of excess was prepared to recover the loss during practical work. And the coating solution was sprayed over drug layered pellets using Fluid bed coater until weight gain was achieved and % yield was calculated.
- The solid content of barrier coating suspension was 10% w/w
- The lot size for barrier coating was 1000 units.

Stage III:

Preparation of Duloxetine HCl Enteric coated pellets:

- Eudragit L30-D-55 is added to part quantity of purified water under stirring using Remi propellant stirrer, mixed for 10 minutes to get uniform suspension.
- The suspension is neutralized to different pH (4.00, 4.25, 4.50, 4.75, 5.00 & 5.25 respectively) using 0.1N Sodium hydroxide

- solution to the required pH, mixed for 30 minutes.
- Triethyl citrate was added to the above solution under stirring, mixed for 10 minutes.
- Talc was suspended in purified water separately, added to the above solution and mixed for 30 minutes.
- The resulting suspension was filtered through ASTM 60#, coated on barrier coated pellets using fluid bed processor (GPCG 1.1)
- During the preparation of coating solution the 10% of excess was prepared to recover the loss during practical work. And the coating solution was sprayed over barrier coated pellets using Fluid bed coater until weight gain was achieved and % yield was calculated.
- The solid content of enteric coating suspension was 15% w/w

Stage IV:

Preparation of Duloxetine HCl Top coated pellets:

- Top coating solution was prepared by dispersing Opadry white Y-1-7000 and Talc in purified water, mixed for 45 minutes, using Remi propellant stirrer.
- The resulting suspension was filtered using ASTM 60#, and coated on enteric coated pellets using fluid bed processor (GPCG 1.1)
- Barrier coating was performed in different lots, to the weight gain of 5% w/w.
- During the preparation of coating solution the 10% of excess was prepared to recover the loss during practical work. And the coating solution was sprayed over enteric coated pellets using Fluid bed coater until weight gain was achieved and % yield was calculated.
- The solid content of barrier coating suspension was 10% w/w
- The lot size for enteric coating was 1000 units.

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Encapsulation: The top coated pellets were cured for 2 hrs using tray drier, at 50°C. The cured pellets

were filled in to size "1" hard gelatin capsules, and evaluated for assay, dissolution and acid resistance.

TABLE 2: COMPOSITION OF DULOXETINE HCL ENTERIC COATED PELLETS

Duloxetine Drug layering										
S. No.	Ingredients	CE15	CE16	CE17	CE18	CE19	CE20	CE21		
1	1 Sugar spheres (710-850μ)		120	120	120	120	120	120		
2	Duloxetine HCl	67.3	67.3	67.3	67.3	67.3	67.3	67.3		
3	Povidone K-30	10	10	10	10	10	10	10		
5	Talc USP	5	5	5	5	5	5	5		
6	Corn starch NF	9.7	9.7	9.7	9.7	9.7	9.7	9.7		
7	Sodium chloride	5	5	5	5	5	5	5		
8	Purified water	qs	qs	qs	qs	qs	qs	qs		
	Sub total	217	217	217	217	217	217	217		
		Baı	rier coatin	g			•			
1	Opadry white Y-I-7000	43.4	43.4	43.4	43.4	43.4	43.4	43.4		
2 Purified water		qs	qs	qs	qs	qs	qs	qs		
	Sub total	260.4	260.4	260.4	260.4	260.4	260.4	260.4		
		Ent	teric coatin	g						
1	Eudragit L30-D55	133.54	133.54	133.54	133.54	133.54	133.54	133.54		
2	Triethyl citrate	4.01	4.01	4.01	4.01	4.01	4.01	4.01		
3	Talc USP	8.01	8.01	8.01	8.01	8.01	8.01	8.01		
3 1N sodium hydroxide solution		-	qs to pH 4.0	qs to pH 4.25	qs to pH 4.50	qs to pH 4.75	qs to pH 5.0	qs to pH 5.25		
4 Purified water		qs	qs	qs	qs	qs	qs	qs		
	Sub total	312.48	312.48	312.48	312.48	312.48	312.48	312.48		
Top coating										
1	Opadry white Y-I-7000	15.624	15.624	15.624	15.624	15.624	15.624	15.624		
2	Purified water	qs	qs	qs	qs	qs	qs	qs		
	Sub total	332.11	332.11	332.11	332.11	332.11	332.11	332.11		

TABLE 3: PROCESSING PARAMETERS AT VARIOUS STEPS

Processing parameters	Drug Coating	Barrier coating	Enteric coating	Top coating
Inlet Temperature (°C)	55-58	52-55	38-42	52-55
Exhaust Temperature (°C)	32-36	31-32	27-28	31-32
Product Temperature (°C)	40-44	41-43	32-33	41-43
Spray rate (g/min)	2.5-7	1.8-2	4-6	1.8-2
Atomization (bar)	1.1-1.5	1.1-1.3	1.1-1.3	1.1-1.3
Air flow (CFM)	52-73	52-70	52-68	52-70
Spray nozzle dia (mm)	1.0	0.8	1.0	0.8

Evaluation of Duloxetine HCl Enteric coated pellets Assay ^{9, 11}: Pellets from the capsule were dispersed in to 190 ml of pH 6.8 phosphate buffers by ultra-sonication for 30 minutes followed by 10 minutes stirring using magnetic stirrer. The solution was then filtered and the residues over filter paper were washed with 10 ml phosphate buffer. The solution was then diluted up to suitable

concentration and absorbance was measured using double beam UV-VIS Spectrophotometer at 289 nm.

Acid Resistance test ¹¹:

Principle: Residual Assay

Apparatus: USP Dissolution apparatus type I (basket)

Simulated Gastric fluid: 0.1N HCl (pH 1.2)

Volume of media: 1000 ml

Capsules were placed in the Basket and were rotated at 100 rpm at 37 ± 0.5 oC for 2 (Two) hours. After two hours drug content left in the pellets was assayed. Pellets left in the Basket after two hours were dissolved in 190ml 6.8 pH Phosphate buffer for 30 minute by Ultra sonicator followed by 10 min stirring using magnetic stirrer until pellets disintegrates completely.

The solution was filtered and the residues over filter paper were washed with 10 ml phosphate buffer. The solution was then diluted up to suitable concentration and absorbance was measured using double beam UV-VIS Spectro photometer at 289nm. Drug release in 0.1 N HCl was calculated using following equation;

Drug released in Gastric Fluid = Drug content of Capsule – Residual Assay..... (I)

In-vitro drug release study ¹²: Capsules were evaluated for *in-vitro* release study in 0.1 N HCl and phosphate buffer 6.8 pH, 0.1 N HCl and phosphate buffer 6.0 pH and 0.1 N HCl and phosphate buffer 5.5 pH. The drug dissolution test of Capsule was carried out using USP Dissolution apparatus type I (basket). Capsules were placed into the baskets and 1000 ml of 0.1 N HCl (pH 1.2) solution was filled in to the beaker. The baskets were rotated at 100 rpm.

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Buffer temperature was maintained at $37 \pm 0.5^{\circ}\text{C}$ for two hours. Then 0.1N HCl solution was replaced with 1000 ml of pH 6.8 phosphate buffer, and the baskets were rotated at 100 rpm and $37 \pm 0.5^{\circ}\text{C}$ buffer temperature. Then 10 ml of sample aliquots were collected at 90 minutes. The absorbance of sample was then measured using Double beam UV Visible spectrophotometer at 289 nm.

RESULTS & DISCUSSIONS: Assay: The filled capsules were evaluated for assay and acid resistance, and the results are tabulated below:

Batch Number ↓	Drug content*	Acid resistance*				
Datell Number \$	Di ug content	Residual Assay	% Drug Release in 0.1N HCl*			
Cymbalta (A525312)	100.7 ± 0.46	99.0 ± 0.36	1.7			
CE-15	99.7 ± 0.38	99.6 ± 0.42	0.0			
CE-16	99.7 ± 0.46	99.4 ± 0.47	0.3			
CE-17	99.3 ± 0.38	99.7 ± 0.5	0.0			
CE-18	99.2 ± 0.46	99.7 ± 0.5	0.1			
CE-19	100.5 ± 0.49	100.1 ± 0.2	0.4			
CE-20	100.8 ± 0.35	99.0 ± 0.31	1.8			
CE-21	100.8 ± 0.36	91.0 ± 1.36	9.8			

^{*}Listed value indicates mean value of results and Standard deviation (Where n=3)

In-vitro **Drug release study:** *In-vitro* drug release study was performed using USP apparatus-I (Basket type) for reference product (Cymbalta) and In-house formulations (CE15 to CE 21) in 0.1 N HCl for first two hrs followed by pH 6.8 phosphate buffer for 120 mins, 0.1 N HCl for first two hrs followed by pH 6.0 phosphate buffer for 120 mins and 0.1 N HCl for first two hrs followed by pH 5.5 phosphate buffer for 120 mins, the results are presented in table.

In-vitro Dissolution in 0.1N HCl for 2 hrs followed by pH 6.8 phosphate buffer for 120 minutes:

Apparatus : USP Type-I (Basket)

Medium : 0.1 HCl for 2 hrs followed pH 6.8

Phosphate buffer for 120 mins

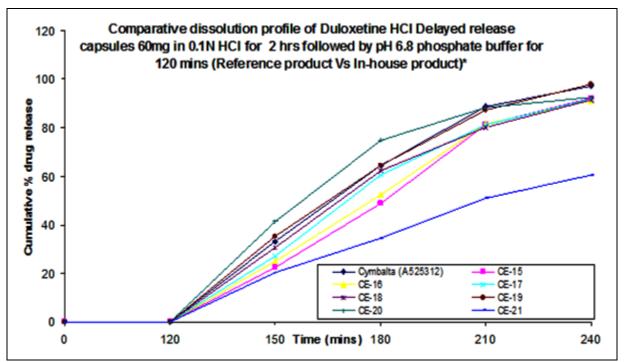
RPM : 100

Table: Dissolution of Duloxetine hydrochloride delayed release capsules in 0.1N HCl for 2 hrs followed by pH 6.8 phosphate buffer for 120 mins, USP-I, 1000ml, 100RPM

TABLE 6: COMPARATIVE DISSOLUTION PROFILE OF DULOXETINE HCI DELAYED RELEASE CAPSULES 60MG IN 0.1N HCl FOR 2 HRS FOLLOWED BY PH 6.8 PHOSPHATE BUFFER FOR 120 mins (Reference product Vs In-house product)*

Batch Number \rightarrow Time (mins) \downarrow	Cymbalta (A525312)	CE-15	CE-16	CE-17	CE-18	CE-19	CE-20	CE-21
30 mins	33.1±0.9	22.6 ± 0.9	24.9 ± 0.4	27.1 ± 0.5	30.5 ± 0.7	35.2 ± 1.4	41.2 ± 0.4	20.3 ± 0.9
60 mins	64.4 ± 2.0	48.8 ± 2.9	52.2 ± 0.8	60.7 ± 0.5	62.2 ± 0.6	64.5 ± 0.9	74.8 ± 0.6	34.6 ± 2.0
90 mins	$88.6 \pm~0.2$	81.3 ± 0.9	81.4 ± 1.0	81.1 ± 1.4	80.2 ± 0.3	87.1 ± 1.3	88.4 ± 0.2	50.9 ± 0.3
120 mins	97.1 ± 0.4	92.4 ± 0.3	91.2 ± 1.6	91.9 ± 0.5	91.5 ± 0.8	97.8 ± 1.1	92.5 ± 0.1	60.4 ± 0.7

^{*}Listed value indicates mean value of results and Standard deviation (Where n=3)



GRAPH 1: COMPARATIVE DISSOLUTION OF DULOXETINE HCI REFERENCE (CYMBALTA) Vs IN-HOUSE FORMULATION IN 0.1N HCI FOR 2 HRS FOLLOWED BY $_{
m PH}$ 6.8 PHOSPHATE BUFFER FOR 120 MINUTES

The dissolution profile indicates the formulations CE-18 & CE-19 are comparable to reference product. Whereas, the formulation CE-21 (Eudragit neutralized to pH 5.25), shows lower dissolution profile, and the extent is 60.4% is only achieved. The enteric polymer is not suitable for the formulation when it is neutralized to pH more than 5.0.

The formulation CE-15, CE-16, CE-17 & CE -20 are not having significance difference in dissolution profile and discrimination is not observed in the dissolution profile. Hence, it was required to evaluate the dissolution in 0.1N HCl for 2 hrs

followed by pH 5.5 phosphate buffer for 120 minutes.

In-vitro Dissolution in 0.1N HCl for 2 hrs followed by pH 5.5 phosphate buffer for 120 minutes:

Apparatus : USP Type-I (Basket)

Medium : 0.1 HCl for 2 hrs followed pH 5.5

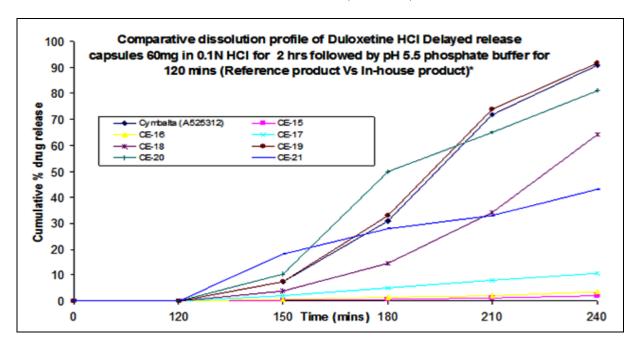
Phosphate buffer for 120 mins

RPM : 100

TABLE 7: COMPARATIVE DISSOLUTION PROFILE OF DULOXETINE HCI DELAYED RELEASE CAPSULES 60mg IN 0.1N HCl FOR 2 HRS FOLLOWED BY pH 5.5 PHOSPHATE BUFFER FOR 120 MINS (Reference product Vs In-house product)*

Batch Number \rightarrow Time (mins) \downarrow	Cymbalta (A525312)	CE-15	CE-16	CE-17	CE-18	CE-19	CE-20	CE-21
30 mins	7.5 ± 0.3	0.2 ± 0.1	0.5 ± 0.3	2.1 ± 0.6	4.0 ± 0.3	7.3 ± 0.9	10.5 ± 0.7	18.2 ± 1.5
60 mins	30.8 ± 1.0	0.6 ± 0.1	1.4 ± 0.2	$5.1 \pm~0.6$	$14.5 \pm \ 2.0$	33.0 ± 1.3	50 ± 1.2	28 ± 0.2
90 mins	71.9 ± 0.4	1.3 ± 0.1	2.2 ± 0.2	8.1 ± 0.6	34.2 ± 1.5	73.9 ± 1.1	65 ± 3.2	33 ± 1.0
120 mins	90.9 ± 0.6	$2.0 \pm \ 0.2$	3.5 ± 0.2	10.7 ± 0.5	64.2 ± 1.4	$91.8 \pm~0.6$	81 ± 0.8	43 ± 0.6

^{*}Listed value indicates mean value of results and Standard deviation (Where n=3)



The dissolution profile in pH 5.5 phosphate buffer indicates, the formulation CE-19 shows the extent of 91.8% dissolution and the dissolution profile is comparable to reference product. Formulation CE-15 to CE-17 are not having release in pH 5.5 phosphate buffer. CE-18, CE-20 & CE-21 are having lower dissolution profile, and the extent of 64.2%, 81% & 43% is released respectively. The discrimination in dissolution difference from pH 6.8 phosphate buffer and pH 5.5 phosphate buffer is high. It was decided to evaluate the dissolution in pH 6.0 phosphate buffer.

In-vitro Dissolution in 0.1N HCl for 2 hrs followed by pH 6.0 phosphate buffer for 120 minutes:

Apparatus : USP Type-I (Basket)

Medium : 0.1 HCl for 2 hrs followed pH 6.0

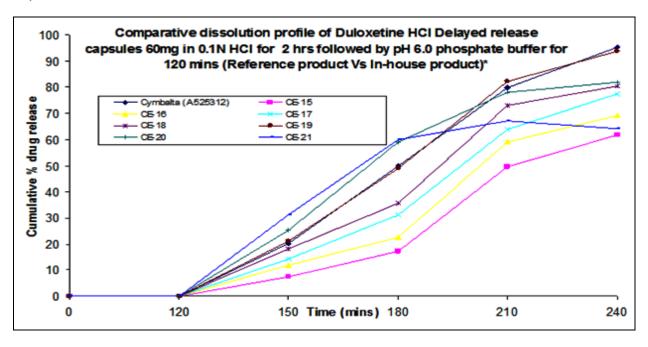
Phosphate buffer for 120 mins

RPM : 100

TABLE 8: COMPARATIVE DISSOLUTION PROFILE OF DULOXETINE HCl DELAYED RELEASE CAPSULES 60mg IN 0.1N HCl FOR 2 HRS FOLLOWED BY pH 6.0 PHOSPHATE BUFFER FOR 120 MINS (Reference product Vs In-house product)*

In-nouse product)**								
Batch Number →	Cymbalta (A525312)	CE-15	CE-16	CE-17	CE-18	CE-19	CE-20	CE-21
Time (mins)↓								
30 mins	20.2 ± 0.4	7.5 ± 0.8	12 ± 1.6	14.1 ± 0.3	18.1 ± 0.6	21 ± 0.4	25.1 ± 1.2	31.1 ± 0.5
60 mins	49.8 ± 0.3	17.3 ± 1.1	22.6 ± 0.9	31.2 ± 0.6	35.7 ± 1	49.1 ± 0.6	59 ± 0.4	60 ± 1
90 mins	79.8 ± 0.3	49.5 ± 2.6	59.1 ± 2.2	63.9 ± 1.4	73 ± 1.2	82.1 ± 0.8	78 ± 1.1	67 ± 5.5
120 mins	95.2 ± 0.4	61.6 ± 1.2	69.2 ± 0.7	77.4 ± 1.0	80.3 ± 1.0	93.9 ± 0.3	82 ± 0.5	64 ± 1.5

^{*}Listed value indicates mean value of results and Standard deviation (Where n=3)



The dissolution profile in pH 6.0 phosphate buffer indicates, the formulation CE-19 shows the extent of 93.9% dissolution and the dissolution profile is comparable to reference product. Whereas, the remaining formulations are having the extent of dissolution to the maximum of 82%. The formulation CE-19 is comparable to the reference product.

CONCLUSION: Duloxetine Hydrochloride delayed release pellets were evaluated dissolution at various pH to achieve the complete dissolution with the T_{lag} of 2 hrs. The formulation CE-19 (enteric polymer neutralized to pH 4.7) shown the better extent of dissolution in all the three dissolution media 0.1N HCl for 2hrs followed by pH 6.8 phosphate buffer for 120 minutes, 0.1N HCl for 2hrs followed by pH 5.5 phosphate buffer for 120 minutes and 0.1N HCl for 2hrs followed by pH 6.0 phosphate buffer for 120 minutes. The dissolution profile is also comparable to reference product.

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